

WestminsterResearch

<http://www.westminster.ac.uk/westminsterresearch>

**The distinctive hepatoprotective activity of turmeric kombucha
(*Curcuma longa*) induced by diethylnitrosamine in Balb/C mice
Elok Zubaidah, Ike Susanti, Hidayat Sujuti, Erryana Martati, Aldilla
Putri Rahayu, Ignatius Srianta and Ihab Tewfik**

NOTICE: this is the authors' version of a work that was accepted for publication in Food Bioscience. Changes resulting from the publishing process, such as peer review, editing, corrections, structural formatting, and other quality control mechanisms may not be reflected in this document. Changes may have been made to this work since it was submitted for publication. A definitive version was subsequently published in Food Bioscience, volume 55, October 2023, 103043.

The final definitive version in Food Bioscience is available online at:

<https://doi.org/10.1016/j.fbio.2023.103043>

© 2023. This manuscript version is made available under the CC-BY-NC-ND 4.0 license

<https://creativecommons.org/licenses/by-nc-nd/4.0/>

The WestminsterResearch online digital archive at the University of Westminster aims to make the research output of the University available to a wider audience. Copyright and Moral Rights remain with the authors and/or copyright owners.

Journal Pre-proof

The distinctive hepatoprotective activity of turmeric kombucha (*Curcuma longa*) induced by diethylnitrosamine in Balb/C mice

Elok Zubaidah, Ike Susanti, Hidayat Sujuti, Erryana Martati, Aldilla Putri Rahayu, Ignatius Srianta, Ihab Tewfik



PII: S2212-4292(23)00694-6

DOI: <https://doi.org/10.1016/j.fbio.2023.103043>

Reference: FBIO 103043

To appear in: *Food Bioscience*

Received Date: 27 May 2023

Revised Date: 10 August 2023

Accepted Date: 12 August 2023

Please cite this article as: Zubaidah E., Susanti I., Sujuti H., Martati E., Rahayu A.P., Srianta I. & Tewfik I., The distinctive hepatoprotective activity of turmeric kombucha (*Curcuma longa*) induced by diethylnitrosamine in Balb/C mice, *Food Bioscience* (2023), doi: <https://doi.org/10.1016/j.fbio.2023.103043>.

This is a PDF file of an article that has undergone enhancements after acceptance, such as the addition of a cover page and metadata, and formatting for readability, but it is not yet the definitive version of record. This version will undergo additional copyediting, typesetting and review before it is published in its final form, but we are providing this version to give early visibility of the article. Please note that, during the production process, errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

© 2023 Published by Elsevier Ltd.

1 **The distinctive hepatoprotective activity of turmeric kombucha (*Curcuma longa*) induced**
2 **by diethylnitrosamine in Balb/C mice**

3
4 Elok Zubaidah^a, Ike Susanti^a, Hidayat Sujuti^b, Erryana Martati^a, Aldilla Putri Rahayu^c, Ignatius
5 Srianta^d, and Ihab Tewfik^e
6

7 ^aDepartment of Food Science and Technology, Faculty of Agricultural Technology, Brawijaya
8 University, Jalan Veteran, Malang, 65145, East Java, Indonesia

9 ^bDepartment of Biomedical, Faculty of Medicine, Brawijaya University, Jalan Veteran, Malang,
10 65145, East Java, Indonesia

11 ^cDepartment of Agronomy, Faculty of Agriculture, Brawijaya University, Jalan Veteran, Malang,
12 65145, East Java, Indonesia

13 ^dDepartment of Food Technology, Faculty of Agricultural Technology, Widya Mandala Surabaya
14 Catholic University, Jalan Dinoyo, 42-44, Surabaya, 60265, Indonesia

15 ^eSchool of Life Sciences, University of Westminster, 115 New Cavendish Street, London, W1W
16 6UW, UK
17

18 Corresponding author:

19 Elok Zubaidah

20 elzoeba@yahoo.com, elok@ub.ac.id

21 Brawijaya University, Jalan Veteran, Malang, 65145, East Java, Indonesia

22 Tel: +62-341-551611 ext: 126
23

24 **Abstract**

25

26 This study aims to investigate the potential hepatoprotective activity of turmeric kombucha before
27 and after fermentation and to compare such distinctive activity in turmeric kombucha versus
28 turmeric essence beverage (turmeric beverage without fermentation). Liquid chromatography-
29 mass spectrometer (LC-MC) analyses revealed the presence of bioactive compounds in turmeric
30 kombucha and turmeric essence beverages. *In vivo* tests appraised the levels of alanine
31 transaminase (ALT), aspartate transaminase (AST), malondialdehyde (MDA) in Balb/C mice and
32 the histology of their livers was determined. Upon successful fermentation process new
33 compounds such as: tetrahydrocurcumin, ferulic acid, glucuronidated curcumin, cyclofenil, acetic
34 acid, glucuronic acid, and D-saccharic acid-1,4-lactone were produced in turmeric kombucha,
35 which were not found in non-turmeric kombucha. The positive effect of fermentation has boosted
36 the hepatoprotective activity of turmeric kombucha through the release of compounds and the
37 production of new bioactive compounds. Therefore, fermented turmeric kombucha had a greater
38 effect on the hepatoprotective activity compared to turmeric essence beverage in Balb/C mice.

39

40 **Keywords:** Turmeric, kombucha, fermentation, hepatoprotective, diethylnitrosamine

41

42

43 1. Introduction

44 The liver is the main organ that plays a part in the metabolism of drugs and toxic chemicals.
45 Excessive exposure to toxins can cause hepatotoxicity (Maran et al., 2022). Several factors that
46 contribute to liver toxicity include genetic, carcinogenic, and interactions with drugs and alcohol
47 (Malaguarnera et al., 2012). Exposure to chemicals such as diethylnitrosamine (DEN) can induce
48 liver damage and cause oxidative stress, inflammation, and deoxyribonucleic acid (DNA)
49 destruction (Al-Rejaie et al., 2009). Liver damage is triggered when enzymes in the liver undergo
50 lysis and are released into the blood. Compounds that can maintain and repair liver damage are
51 called hepatoprotectives (Maran et al., 2022).

52 Turmeric is a medicinal plant with functional biological properties and benefits for human
53 health. The bioactive compounds contained in turmeric are curcuminoids, essential oils, tannins,
54 and minerals. It was reported that 2%-5% of turmeric essential oils consisted of phenylpropane
55 turmerone derivatives (aryl-turmerone, alpha turmerone, and beta turmerone) (Goenka et al.,
56 2021). Curcumin has been known to have antioxidant activity, as a radical scavenger, and as a
57 catalyst for the formation of hydroxyl radicals (Bimonte et al., 2013). However, the bioavailability
58 of active compound in curcumin is relatively low due to binding to other compounds.

59 The fermentation process is one of the food processing methods in which large substrates are
60 broken down into simpler ones assisted by the action of microorganisms. Kombucha is a traditional
61 drink from the fermentation process of sweet tea with a mixed culture of bacteria and yeast. The
62 mixed culture is commonly known as SCOBY (symbiotic culture of bacteria and yeast) which
63 produces a floating biofilm known as microbial cellulose layer or 'nata' (Zailani & Adnan, 2022).
64 The substrate often used is steeped tea, so 'nata' is also known as "tea mushroom" or tea fungus"
65 (Battikh et al., 2012). Zubaidah et al. (2021) has explored the chemical, microbiological, and
66 antibacterial characteristics of turmeric kombucha, concluding that turmeric can be processed as a
67 kombucha with notable microbiological and antibacterial activity. There have been no research on
68 turmeric kombucha as a hepatoprotective by the time this article was written. This study was
69 conducted to determine the potential hepatoprotective property of turmeric kombucha.

70

71 2. Materials and methods

72 2.1. Materials

73 Turmeric (*Curcuma longa*) was obtained from a local traditional market in Malang, East Java,
74 Indonesia. Commercial kombucha starter (SCOBY), sugar, and chemicals were obtained from
75 local distributors. SCOBY consists of acetic acid bacteria (AAB) Acetobacteraceae and osmophilic
76 yeast (Filippis et al., 2018). DEN was obtained from Tokyo Chemical Industry Co., Ltd. (Tokyo,
77 Japan). Ketamine HCl injection (Bernofarm; anesthesia) was obtained from Bioscience Institute
78 Universitas Brawijaya (Malang, East Java, Indonesia). Thirty male Balb/c mice as the
79 experimental animals (6 wks old, 20-30 g). Water and feed were given ad libitum during 1-week
80 period of acclimatization.

81

82 2.2. Kombucha turmeric and turmeric essence beverage solution preparation and analysis

83 Kombucha preparation and analysis was done according to previous research by Zubaidah et
84 al., 2021. Turmeric was peeled and washed, sliced to ± 1 cm thick, dried in a dry cabinet at 70°C
85 for 12 h, and grinded using a blender (Philips, Amsterdam, Netherlands). Turmeric powder was
86 brewed in hot water with a ratio of 1:10 (5 g of powder in 500 mL of water) for 5 min, 10% of
87 sugar was added, and after cooling, 10% (v/v) of SCOBY starter was added. The mouth of the jar
88 was covered with a cloth and tied. The jar was placed in a room that was not exposed to direct
89 sunlight and at room temperature (30°C) to ferment for 12 d. Non-fermented turmeric essence
90 beverage was prepared with a concentration of 1.2% (6 g of powder in 500 mL of water) and was
91 run through the same procedures as turmeric kombucha, without the addition of a kombucha
92 starter.

93

94 **2.3. Identification of the components of turmeric kombucha bioactive compound**

95 LC-MS analysis was carried out using a high-performance liquid chromatography-mass
96 spectrometer (LC-20A, Shimadzu Corporation, Kyoto, Japan) equipped with a Waters 2695
97 preconditioner pump (Waters Corporation, MA, USA). The MS calibration used was Kromtekindo
98 PRO\ACQUDB/Mass. MS scan was carried out with an initial mass of 50.0/s and final mass of
99 1200.0, scan time was 5.00, interscan time was 0.10 s, start time was 0.0 min, and end time was
100 50 min. The storage volume used was 50 L, flow ramp was 0.10, flow was 0.20 mL/min, stop time
101 was 35 min, column temperature was 40°C, column temperature limit was 10°C, minimum
102 pressure was 0.0 Bar, maximum pressure was 300 Bar, pre-column volume was 0 L, column type
103 2, with a size of 1 mm x 100 mm. Solvent 'A' was 10% methanol, solvent 'B' was 90% water,
104 solvent 'C' was 0 formic acid, and solvent 'D' was 0 acetonitrile. Draw speed; needle depth was
105 1/mm, sample temperature was 20°C, and sample limit temperature was 20°C.

106

107 **2.4. Animal experiment and analysis**

108 Testing of hepatoprotective activity was carried out using the *in vivo* method with 30 male
109 Balb/c mice (6 wks old, 20-30 g). The research was approved by Brawijaya University Research
110 Ethics Committee (Ethical Clearance No. 104-KEP-UB-2021). Grouping of the mice was carried
111 out according to the experimental design with 10 treatments (Table 1). Turmeric kombucha and
112 turmeric essence beverage were given daily for 3 wks, with the induction carried out only after
113 then. DEN with a dose of 100 mg/kg was given through an intraperitoneal injection process at the
114 rate of 1 injection/wk for 2 wks. During the DEN injection treatment, turmeric kombucha and
115 turmeric essence beverage were still being given, with an incubation period of 1 wk. Mice without
116 DEN injections were treated according to the grouping. On the 49th day, surgery was performed
117 after fasting for 24 h from the last day of treatment. An anesthesia process was used during the
118 induction of 0.2 mL ketamine (50 mg/kg). During surgery, blood serum samples were taken from
119 the heart and liver. Parameters observed were alanine transaminase (ALT) activity, aspartate
120 transaminase (AST), malondialdehyde (MDA), and liver histology (Fig. 1).

121

122 **2.4.1. ALT and AST enzyme (Modification Devaraj et al., 2014)**

123 The clotted blood samples were centrifuged at 3000 rpm (3461 x g in a EBA 200, Andreas
124 Hettich GmbH & Co. KG, Tuttlingen, Germany) at room temperature (30°C) for 15 min to separate
125 the cell nucleus from the blood serum. Blood serum was then taken and biochemically tested for
126 the amount of AST and ALT enzymes.

127

128 **2.4.2. MDA enzyme (Modification Devaraj et al., 2014)**

129 As much as 10% of the liver homogenate was mixed into 0.1 M Tris-HCl buffer pH 7.4 at
130 40°C. The sample was homogenized (VELP Scientifica Srl, Usmate, Italy) with at 1000 rpm for 2
131 min. The homogenate was centrifuged at 1000 rpm at 40°C for 10 min to separate the nucleus and
132 cell solids. The supernatant was tested for the amount of MDA to see the level of liver oxidation.

133

134 **2.4.3. Histopathological observation (Modification Jantararussamee et al., 2020)**

135 Histopathological observations of mice were carried out by taking some liver samples from
136 each group by dissection. The results of the dissection were dehydrated with 50%-100% ethanol
137 and given paraffin with a thickness of 5 cm. The paraffin-treated sections were then stained with
138 hematoxylin and eosin (HE) to color the cell parts and observed under a light microscope
139 (Olympus Corporation, Tokyo, Japan) with magnifications of 40x, 100x, and 400x.

140

141 **2.5. Statistical analysis**

142 The statistical analysis was carried out by comparison of all the groups. Analysis of variance
143 (ANOVA) was used and followed by Fisher's exact test at $p < 0.05$. All statistical analyses were
144 carried out using Minitab software (17.0 version, Minitab, LLC).

145

146

147 **3. Result and discussion**

148 **3.1. Turmeric kombucha and turmeric essence beverage characteristics**

149 Turmeric kombucha and turmeric essence beverage were used as treatments to mice.
150 Physicochemical and microbiological analysis were conducted prior to the *in vivo* procedures. The
151 higher the concentration of turmeric, then the lower the microbe total and acid total obtained. The
152 higher the total phenol concentration of turmeric, then the higher the antioxidant activity. The best
153 treatment results were obtained with 1% of turmeric kombucha concentration (Zubaidah et al.,
154 2021).

155 The characteristics of turmeric kombucha and turmeric essence beverage found by Zubaidah
156 et al. (2021) are shown on Table 2. Turmeric kombucha showed higher total phenolic content and
157 antioxidant activity elevation compared to turmeric essence. Turmeric kombucha also had a higher
158 total of titratable acid, lower pH, and an increase of the AAB total. This was due to the addition of
159 kombucha starter. Kombucha starter mainly comprised of bacteria and yeast, which led to them
160 influencing the microbial characteristics of turmeric and black tea kombucha. According to
161 Zubaidah et al. (2021), black tea kombucha recorded 1.3×10^8 CFU/mL of total microbes on day-
162 14, higher than the turmeric kombucha with 2.0×10^7 CFU/mL. Microbial activity results in the

163 breakdown of turmeric bioactive compounds. Turmeric kombucha showed an increase of total
164 titratable acid, decrease of pH, higher total phenolic content, and lower IC₅₀ value which enabled
165 better free radical degradation than turmeric essence beverage. This was due to the existence of
166 organic acids produced by microorganisms during fermentation. This proved that kombucha
167 fermentation increased total phenolic content and antioxidant activity of turmeric.

168

169 **3.2. Components of bioactive compounds in turmeric kombucha and turmeric essence**

170 **beverage**

171 Identification of chemical compounds contained in turmeric kombucha and turmeric essence
172 beverage using LC-MS (Table 3) revealed that they contained phenolic compounds, curcumin,
173 demethoxycurcumin, bisdemethoxycurcumin, several compounds derived from curcumin, and
174 organic acids. Chemical compounds detected in the phenolic group were nitrophenol, phenol, and
175 quinoline. Phenol compounds are secondary metabolites of plant metabolism that attach to metal
176 ions which can fight free radicals and increase antimicrobial activity (Cavalcanti et al., 2012).
177 Phenolic compounds have functional abilities such as cardiovascular inhibition, anticancer, and
178 chronic disease prevention (Soto-Quintero et al., 2019).

179 Chemical compounds detected in the curcuminoids group were curcumin,
180 bisdemethoxycurcumin, and demethoxycurcumin. The derivative components of the curcumin
181 compounds consisted of ferulic acid, acetylsalicylic acid, guaiacol, eugenol, licopyranocoumarin,
182 and phenyl. Ferulic acid is an acid consisting of trans-cinnamic acid which has methoxy and
183 substitution of hydroxyl on the phenyl ring. Ferulic acid has bioactivities as an antioxidant, anti-
184 inflammatory, inhibitor of apoptosis, and cardioprotective prevention. Ferulic acid is a chemical
185 compound that is commonly found in plants, belonging to a group of secondary metabolites that
186 bind to esters, glycosides, components of lignin, and tannins (Mattila & Kumpulainen, 2002).
187 Based on the chemical structure, it can be divided into benzoic acid derivatives by substitution of
188 hydroxyl and methoxy groups and phenolic acids. Ferulic acids such as caffeic, p-coumaric,
189 sinapic acid, and vanillin acid are cinnamic acid derivatives (Bezerra et al., 2017).

190 Acetylsalicylic acid is a chemical compound that functions as an analgesic drug or pain
191 reliever. Acetylsalicylic acid can bind and acetylate serine residues in cyclooxygenase (COX),
192 resulting in decreased prostaglandin synthesis, platelet aggregation, and inflammation.
193 Acetylsalicylic acid has analgesic, antipyretic, and anticoagulant properties. Research conducted
194 by Purpura et al. (2018) reported that curcumin significantly reduced pain in the legs of
195 experimental rats. Prostaglandins are known to reduce pain receptors through the COX and
196 lipoxygenase (LOX) pathways. Conditions like this can suppress COX-2 and 5-LOX which are
197 enzymes that cause pain. Curcumin showed a significant antipyretic effect with decreasing rectal
198 temperature. The decrease in temperature can be caused by the presence of acetylsalicylic acid
199 which can inhibit prostaglandins (Hatcher et al., 2008).

200

201 **3.3. Hepatoprotective activity**

202 **3.3.1. Alanine transaminase**

203 ALT is an enzyme present in the cytosol of liver parenchyma cells and thus is a more specific
204 parameter to analyze liver damage. If there was damage to the liver, the cell would undergo lysis
205 and ALT enzymes would come out of the cells and be carried in the blood circulation. This
206 indicated that the ALT enzyme was detected in the analysis of blood serum (Jilkova et al., 2019).
207 Treatment with DEN can affect the activity of ALT in the blood serum of mice. The blood serum
208 of the positive control group (normal diet + DEN) showed higher values than the negative control
209 group (normal diet). ALT activity decreased after the administration of turmeric essence beverage
210 and turmeric kombucha of various concentrations. The administration of turmeric kombucha with
211 a concentration of 0.5 mL/20 g BW showed the largest decrease among the DEN-induced groups,
212 which was 20.851 U/L (Table 3). The normal diet group with DEN induction had the highest ALT
213 value, where there was an increase in the ALT value to 41.147 U/L. DEN damages liver cells,
214 causing lysis and triggering liver cell death. DEN can be metabolized in dysenterylabular
215 hepatocytes followed by oxidative DNA damage reactions (Jilkova et al., 2019). After DEN
216 induction and administration of turmeric essence beverage at a dose of 0.5 mL/20 g BW, the ALT
217 value was reduced to a value of 30.451 U/L. The treatment with turmeric kombucha had lower
218 ALT activity than the turmeric essence beverage treatment. Turmeric kombucha with various
219 concentrations had a higher ability to reduce ALT activity in mouse blood serum. The difference
220 in dosages of turmeric kombucha and turmeric essence beverage showed a significant difference
221 in decreasing ALT activity ($p < 0.05$). The reduction of the ALT enzyme in blood serum was
222 because the ability of curcumin to fight free radicals and induce arachidonic acid metabolism
223 through the COX and LOX pathways (Ak & Gülçin, 2008). The results of research conducted by
224 Bimonte et al. (2013) showed that curcumin can prevent liver toxicity and reduce ALT levels
225 caused by methotrexate induction.

226

227 **3.3.2. Aspartate transaminase**

228 AST is an enzyme found in the cytosol and mitochondria of liver cells, cardiac muscle cells,
229 striated muscles, and kidneys (Jilkova et al., 2019). This indicates that high AST values are not
230 only caused by damage to liver cells but can also occur due to the presence of AST in other cells.
231 If liver cells are lysed, the enzyme will be carried out in the blood circulation so that it can be
232 detected in blood serum analysis (Castro et al., 2015). Treatment with DEN can affect AST
233 activity in the blood serum of mice. The blood serum of the positive control group (normal diet +
234 DEN) shows a higher value than the negative control group (normal diet). AST activity decreased
235 after the administration of turmeric essence beverage and turmeric kombucha of various
236 concentrations. The administration of turmeric kombucha resulted in a higher reduction activity
237 than turmeric essence beverage (Table 4). The increase in the value of AST activity in the positive
238 control group was 40.739 U/L. After the administration of turmeric kombucha and turmeric
239 essence beverage of various concentrations, there was a decrease in AST activity in the blood
240 serum of mice. Notwithstanding, the decrease in AST value in turmeric essence beverage was
241 within normal limits with the lowest value at a dose of 0.5 mL/20 g BW which was 30.077 U/L,

242 while in turmeric kombucha the lowest value was 20.110 U/L within normal limits. This indicates
243 that the higher the dose given, the lower the value of AST activity in the blood serum of mice.

244 The normal AST value for mice is 8-40 U/L. Curcumin is a compound found in turmeric
245 with functions as a hepatoprotective, such as antioxidant activity, anti-inflammatory,
246 antimicrobial, and anticarcinogenic (Karimian et al., 2017). Curcumin and curcumin derivatives
247 such as 5-benzo [1,3] dioxol-5-il-1-phenyl-penta-2,4-dien-1 have the ability as hepatoprotectives
248 to protect and repair damaged liver cells. According to research conducted by Kapelle et al. (2020),
249 the increase in turmeric hepatoprotective activity was due to microbial activity during the
250 kombucha fermentation process. According to Acosta-Cota et al. (2019), yeast and
251 *Gluconacetobacter* sp. formed glucuronic acid during the fermentation of kombucha.
252 Identification with LC-MS of turmeric kombucha found compounds of organic acids which were
253 glucuronic acid and 1,4-lactone D-saccharic acid (DSL). Glucuronic acid can bind to toxic
254 metabolites or compounds that will be eliminated from the body so these compounds are more
255 water soluble and their toxic activity is reduced. DSL in kombucha tea is a hepatoprotective
256 detoxifier and can curatively maintain liver pathophysiology. In addition to glucuronic acid, it has
257 the potential to clear hepatotoxins caused by toxins such as acetaminophen, carbon tetrachloride,
258 hydrocarbon carcinogens, nitrosamines, and aromatic amines (Bhattacharya et al., 2011).

259

260 3.3.3. Lipid peroxidation level

261 MDA is a product of fat oxidation. The high the levels of MDA the high the levels of fat
262 oxidation in the body. Lipid peroxidation has a role in the pathogenesis of tissue injury, especially
263 in damage caused by several toxic substances (Dzoyem et al., 2014). Normal diet + DEN induction
264 treatment showed the highest MDA levels. Turmeric kombucha and turmeric essence beverage
265 decreased MDA levels. The lowest MDA levels were found in 0.5 mL/20 g BW turmeric
266 kombucha induced mice (Table 4). This was due to turmeric kombucha containing more bioactive
267 compounds, organic acids, and microorganisms compared to turmeric essence beverage. There
268 were several bioactive compounds derived from curcuminoids that have the functional properties
269 of preventing liver damage. In addition, several organic acid compounds in turmeric kombucha
270 could prevent liver damage such as glucuronic acid and DSL, these compounds were not found in
271 turmeric essence beverage. The higher the dose of kombucha, the lower the MDA levels in the
272 mice's serum. This increase in antioxidant activity reduced lipid peroxidation and prevented the
273 formation of MDA (Sobhani et al., 2020). Organic acids such as acetic acid and glucuronic acid
274 have high antioxidant activity. Kombucha was able to reduce liver damage caused by oxidative
275 stress (Gharib, 2009). Glucuronic acid is a bioactive compound in kombucha with high antioxidant
276 activity as a detoxifier in the liver through the glucuronidation process. Glucuronidation is a
277 xenobiotic conjugation process such as; acetylaminofluorene (carcinogenic), aniline, benzoic acid,
278 and steroid compounds. The conjugation process with glucuronyl transferase enzyme is catalyzed
279 by UDP-glucanoyltransferase (Alvarenga et al., 2018; Coton et al., 2017).

280

281 3.4. Liver Histology

282 Liver histology was performed to determine the condition of the cells in the liver,
283 observations were made using preparations from the liver. Liver damage is characterized by the
284 occurrence of inflammatory cell damage, fibrosis, and the formation of acidophilic
285 bodies/apoptotic bodies. The negative control group/normal diet (Fig. 2a) shows that the liver cells
286 looked normal, where the condition of the cells stained with HE had purple cytoplasm, the cell
287 nucleus was clear and had a dark purplish color, the boundaries between the cells were visible, and
288 the central blood vessels were visible. Normal histology has a brownish-red color, shiny, sharp
289 edges, a smooth texture, good cytoplasmic conditions, a prominent nucleus, and sinusoidal spaces.
290 It also has liver lobules and a uniform pattern around polyhedral hepatocytes from the central vein
291 to the periphery (Jantararussamee et al., 2021; Jeyadevi et al., 2019; Mondal et al., 2019). Normal
292 diet groups fed with turmeric essence beverage and turmeric kombucha display similar liver
293 histology to the negative control group (Fig. 2c and 2g). The positive control group (normal diet +
294 DEN) shows the histology of a damaged liver due to the toxicity of DEN (Fig. 2b). Cells had a
295 light pink color and some cells did not have a cell nucleus. The boundaries between liver cells
296 were not clearly visible. Liver cells underwent degradation and inflammation occurred in some
297 cells. Induction of DEN can cause hydropic degradation, mitosis, pseudo-nucleus, apoptosis, and
298 liver necrosis (Santos et al., 2017). The treatment of turmeric kombucha and turmeric essence
299 beverage showed changes in liver histology for the better. The 0.1 mL/20 g BW dose from turmeric
300 essence beverage and kombucha improved cell boundaries and nucleus prominence (Fig. 2d and
301 2h), then doses of 0.3 mL/20 g BW and 0.5 mL/20 g BW produced almost normal liver histology
302 (Fig. 2e, 2f, 2i, and 2j).

303

304 **3.5.Total cell damage**

305 Turmeric is a rhizome that contains curcumin as an anti-inflammatory bioactive.
306 Administration of turmeric kombucha and turmeric essence beverage can reduce and prevent
307 inflammation. Curcumin can inhibit proliferation and reduce inflammation. In addition, it can also
308 reduce levels of MDA, glutathione, nitric oxide (NO), and tumor necrosis factor (TNF) and
309 increase catalase, superoxide dismutase (SOD), and glutathione transferase (GST) activity in the
310 liver (Tokaç et al., 2013). Based on the histology data, the results show liver cell damage due to
311 DEN induction through several damaged and dead cells (Table 5).

312

313 **4. Conclusion**

314 This study shows that the fermentation process can produce other compounds in turmeric
315 kombucha that are not detected in turmeric essence beverage. Fermentation affects the
316 hepatoprotective activity of turmeric through the release of compounds and the production of new
317 bioactive compounds. Therefore, fermented turmeric kombucha offers greater effect on the
318 hepatoprotective activity compared to turmeric essence beverage in experimental animal.

319

320 **Conflict of interest**

321 The authors confirm that they have no conflicts of interest with respect to the work described
322 in this manuscript.

323

324 **Acknowledgments**

325 Thank you to the Universitas Brawijaya Rector for the professorial grant to fully support
326 the current study.

327

328 **References**

329 Acosta-Cota, S.J., Aguilar-Medina, E.M., Ramos-Payán, R., Ruiz-Quiñónez, A.K., Romero-

330 Quintana, J.G., Montes-Avila, J., Rendón-Maldonado, J.G., Sánchez-López, A., Centurión,

331 D., & Osuna-Martínez, U. (2019). Histopathological and biochemical changes in the

332 development of nonalcoholic fatty liver disease induced by high-sucrose diet at different

333 times. *Canadian Journal of Physiology and Pharmacology*, 97(1), 23–36.

334 Aggarwal, B.B., Yuan, W., Li, S., & Gupta, S.C. (2013). Curcumin-free turmeric exhibits anti-

335 inflammatory and anticancer activities: Identification of novel components of turmeric.

336 *Molecular Nutrition and Food Research*, 57(9), 1529–1542.

337 Ak, T., & Gülçin, I. (2008). Antioxidant and radical scavenging properties of curcumin. *Chemico-*

338 *Biological Interactions*, 174(1), 27–37.

339 Al-Rejaie, S.S., Aleisa, A.M., Al-Yahya, A.A., Bakheet, S.A., Alsheikh, A., Fatani, A.G., Al-

340 Shabanah, O.A., & Sayed-Ahmed, M.M. (2009). Progression of diethylnitrosamine-induced

341 hepatic carcinogenesis in carnitine-depleted rats. *World Journal of Gastroenterology*, 15(11),

342 1373–1380.

343 Alvarenga, L.A., Leal, V.O., Borges, N.A., Aguiar, A.S., Faxén-Irving, G., Stenvinkel, P.,

344 Lindholm, B., & Mafra, D. (2018). Curcumin - A promising nutritional strategy for chronic

345 kidney disease patients. *Journal of Functional Foods*, 40, 715–721.

346 Ashok, P.K., & Upadhyaya, K. (2013). Evaluation of Analgesic and Anti-inflammatory Activities

347 of Aerial Parts of *Artemisia vulgaris* L. in Experimental Animal Models. *Journal of*

- 348 *Biologically Active Products from Nature*, 3(1), 101-105.
- 349 Battikh, H., Bakhrouf, A., & Ammar, E. (2012). Antimicrobial effect of Kombucha analogues.
350 *LWT - Food Science and Technology*, 47(1), 71–77.
- 351 Bauer-Petrovska, B., & Petrushevska-Tozi, L. (2000). Mineral and water soluble vitamin content
352 in the Kombucha drink. *International Journal of Food Science and Technology*, 35(2), 201–
353 205.
- 354 Bezerra, G.S.N., Pereira, M.A.V., Ostrosky, E.A., Barbosa, E.G., de Moura, M.F.V., Ferrari, M.,
355 Aragão, C.F.S., & Gomes, A.P.B. (2017). Compatibility study between ferulic acid and
356 excipients used in cosmetic formulations by TG/DTG, DSC and FTIR. *Journal of Thermal*
357 *Analysis and Calorimetry*, 127(2), 1683–1691.
- 358 Bhattacharya, S., Manna, P., Gachhui, R., & Sil, P.C. (2011). Protective effect of Kampuchea tea
359 against tertiary butyl hydro peroxide induced cytotoxicity and cell death in murine
360 hepatocytes. *Indian Journal of Experimental Biology*, 49(7), 511–524.
- 361 Bimonte, S., Barbieri, A., Palma, G., Luciano, A., Rea, D., & Arra, C. (2013). Curcumin inhibits
362 tumor growth and angiogenesis in an orthotopic mouse model of human pancreatic cancer.
363 *BioMed Research International*.
- 364 Bimonte, S., Barbieri, A., Palma, G., Rea, D., Luciano, A., D’Aiuto, M., Arra, C., & Izzo, F.
365 (2015). Dissecting the role of curcumin in tumour growth and angiogenesis in mouse model
366 of human breast cancer. *BioMed Research International*, 16–20.
- 367 Castro, C.A., Dias, M.M.S., Silva, K.A., Reis, S.A., Conceição, L.L., Marcon, L.N., Moraes,
368 L.F.S., & Peluzio, M.C.G. (2015). *Biomarkers in Liver Disease*. Dordrecht: Springer.
- 369 Cavalcanti, Y.V., Brelaz, M.C.A., Neves, J.K., Ferraz, J.C., & Pereira, V.R. (2012). Role of TNF-
370 alpha, IFN-gamma, and IL-10 in the development of pulmonary tuberculosis. *Pulmonary*

- 371 *Medicine*, 745483.
- 372
- 373 Coton, M., Pawtowski, A., Taminiou, B., Burgaud, G., Deniel, F., Coulloume-Labarthe, L., Fall,
374 A., Daube, G., & Coton, E. (2017). Unravelling microbial ecology of industrial-scale
375 Kombucha fermentations by metabarcoding and culture based methods. *FEMS Microbiology*
376 *Ecology*, 93(5), 1–41.
- 377 Devaraj, S., Ismail, S., Ramanathan, S., & Yam, M.F. (2014). Investigation of antioxidant and
378 hepatoprotective activity of standardized *Curcuma xanthorrhiza* rhizome in carbon
379 tetrachloride-induced hepatic damaged rats. *Scientific World Journal*, 2014, 353128.
- 380 Dzoyem, J., Kuete, V., & Eloff, J. (2014). Biochemical parameters in toxicological studies in
381 africa: Significance, principle of methods, data interpretation, and use in plant screenings. In
382 V. Kuete (Ed), *Toxicological Survey of African Medicinal Plants*. Amsterdam: Elsevier. p
383 659–715.
- 384 Filippis, F., Troise, A.D., Vitaglione, P., & Ercolini, D. (2018). Different temperatures select
385 distinctive acetic acid bacteria species and promotes organic acids production during
386 Kombucha tea fermentation. *Food Microbiology*, 73, 11–16.
- 387 Gharib, O.A. (2009). Effects of Kombucha on oxidative stress induced nephrotoxicity in rats.
388 *Chinese Medicine*, 4, 2–7.
- 389 Goenka, S., Johnson, F., & Simon, S.R. (2021). Novel chemically modified curcumin (Cmc)
390 derivatives inhibit tyrosinase activity and melanin synthesis in b16f10 mouse melanoma cells.
391 *Biomolecules*, 11(5).
- 392 Hatcher, H., Planalp, R., Cho, J., Torti, F.M., & Torti, S.V. (2008). Curcumin: From ancient
393 medicine to current clinical trials. *Cellular and Molecular Life Sciences*, 65(11), 1631–1652.

- 394 Hoehle, S.I., Pfeiffer, E., Sólyom, A.M., & Metzler, M. (2006). Metabolism of curcuminoids in
395 tissue slices and subcellular fractions from rat liver. *Journal of Agricultural and Food*
396 *Chemistry*, 54(3), 756–764.
- 397 Ireson, C.R., Jones, D.J.L., Boocock, D.J., Farmer, P.B., Gescher, A.J., Orr, S., Coughtrie,
398 M.W.H., Williams, M.L., & Steward, W.P. (2002). Metabolism of the cancer
399 chemopreventive agent curcumin in human and rat intestine. *Cancer Epidemiology*
400 *Biomarkers and Prevention*, 11(1), 105–111.
- 401 Jakubczyk, K., Kałduńska, J., Kochman, J., & Janda, K. (2020). Chemical profile and antioxidant
402 activity of the kombucha beverage derived from white, green, black and red tea. *Antioxidants*,
403 9(5), 447.
- 404 Jantararussamee, C., Rodniem, S., Taweechotipatr, M., Showpittapornchai, U., & Pradidarcheep,
405 W. (2021). Hepatoprotective Effect of Probiotic Lactic Acid Bacteria on Thioacetamide-
406 Induced Liver Fibrosis in Rats. *Probiotics and Antimicrobial Proteins*, 13(1), 40–50.
- 407 Jayabalan, R., Malbaša, R.V., Lončar, E.S., Vitas, J.S., & Sathishkumar, M. (2014). A review on
408 kombucha tea-microbiology, composition, fermentation, beneficial effects, toxicity, and tea
409 fungus. *Comprehensive Reviews in Food Science and Food Safety*, 13(4), 538–550.
- 410 Jeyadevi, R., Ananth, D.A., & Sivasudha, T. (2019). Hepatoprotective and antioxidant activity of
411 *Ipomoea staphylina* Linn. *Clinical Phytoscience*, 5(1) [insert page number is available].
- 412 Jilkova, Z. M., Kurma, K., and Decaens, T. (2019). Animal models of hepatocellular carcinoma:
413 The role of immune system. *Cancers*, 11, 1–12.
- 414 Kapelle, I.B.D., Manalu, W., Mainassy, M.C., Renur, N.M., & Joris, S.N. (2020). The
415 hepatoprotection effect of the asymmetric curcumin analogue synthetic product in male rat
416 abstract (*Rattus norvegicus* L.). *Systemic Reviews in Pharmacy*, 11(10), 766–771.

- 417 Karimian, M.S., Pirro, M., Majeed, M., & Sahebkar, A. (2017). Curcumin as a natural regulator
418 of monocyte chemoattractant protein-1. *Cytokine and Growth Factor Reviews*, 33, 55–63.
- 419 Malaguarnera, G., Cataudella, E., Giordano, M., Nunnari, G., Chisari, G., & Malaguarnera, M.
420 (2012). Toxic hepatitis in occupational exposure to solvents. *World Journal of*
421 *Gastroenterology*, 18(22), 2756–2766.
- 422 Maran, B. A. V., Iqbal, M., Gangadaran, P., Ahn, B. C., Rao, P. V., & Shah, M. D. (2022).
423 Hepatoprotective potential of malaysian medicinal plants: A review on phytochemicals,
424 oxidative stress, and antioxidant mechanisms. *Molecules*, 27, 1533.
- 425 Martínez-Leal, J., Ponce-García, N., & Escalante-Aburto, A. (2020). Recent evidence of the
426 beneficial effects associated with glucuronic acid contained in kombucha beverages. *Current*
427 *Nutrition Reports*, 9(3), 163–170.
- 428 Mattila, P., & Kumpulainen, J. (2002). Determination of free and total phenolic acids in plant-
429 derived foods by HPLC with diode-array detection. *Journal of Agricultural and Food*
430 *Chemistry*, 50(13), 3660–3667.
- 431 Ming, J., Ye, J., Zhang, Y., Xu, Q., Yang, X., Shao, X., Qiang, J., & Xu, P. (2020). Optimal dietary
432 curcumin improved growth performance, and modulated innate immunity, antioxidant
433 capacity and related genes expression of NF- κ B and Nrf2 signaling pathways in grass carp
434 (*Ctenopharyngodon idella*) after infection with *Aeromonas hydrophila*. *Fish and Shellfish*
435 *Immunology*, 97, 540–553.
- 436 Mondal, M., Hossain, M.M., Rahman, M.A., Saha, S., Uddin, N., Hasan, M.R., Kader, A., Wahed,
437 T.B., Kundu, S.K., Islam, M.T., & Mubarak, M.S. (2019). Hepatoprotective and antioxidant
438 activities of *Justicia gendarussa* leaf extract in carbofuran-induced hepatic damage in rats.
439 *Chemical Research in Toxicology*, 32(12), 2499–2508.

- 440 Mughal, M.H. (2019). Turmeric polyphenols: A comprehensive review. *Integrative Food,*
441 *Nutrition and Metabolism*, 6(6), 1–6.
- 442 Pereira, D.M., Valentão, P., Pereira, J.A., & Andrade, P.B. (2009). Phenolics: From chemistry to
443 biology. *Molecules*, 14(6), 2202–2211.
- 444 Purpura, M., Lowery, R.P., Wilson, J.M., Mannan, H., Münch, G., & Razmovski-Naumovski, V.
445 (2018). Analysis of different innovative formulations of curcumin for improved relative oral
446 bioavailability in human subjects. *European Journal of Nutrition*, 57(3), 929–938.
- 447 Ramezani, M., Hatamipour, M., & Sahebkar, A. (2018). Promising anti-tumor properties of
448 bisdemethoxycurcumin: A naturally occurring curcumin analogue. *Journal of Cellular*
449 *Physiology*, 233(2), 880–887.
- 450 Rocha-Ramírez, L., Pérez-Solano, R., Castañón-Alonso, S., Guerrero, S.M., Pacheco, A.R.,
451 Garibay, M.G., & Eslava, C. (2017). Probiotic *Lactobacillus* strains stimulate the
452 inflammatory response and activate human macrophages. *Journal of Immunology Research*,
453 4607491.
- 454 Santos, N.P., Colaço, A.A., & Oliveira, P.A. (2017). Animal models as a tool in hepatocellular
455 carcinoma research: A Review. *Tumor Biology*, 39(3) [insert page number is available].
- 456 Sayed, M.M., & El-Kordy, E.A. (2014). The protective effect of curcumin on paracetamol-induced
457 liver damage in adult male rabbits: Biochemical and histological studies. *Egyptian Journal of*
458 *Histology*, 37(4), 629–639.
- 459 Sim, Y.Y., Ong, W.T.J., & Nyam, K.L. (2019). Effect of various solvents on the pulsed ultrasonic
460 assisted extraction of phenolic compounds from *Hibiscus cannabinus* L. leaves. *Industrial*
461 *Crops and Products*, 140(1), 111708.
- 462 Sobhani, M., Farzaei, M.H., Kiani, S., & Khodarahmi, R. (2020). Immunomodulatory; anti-

- 463 inflammatory/antioxidant effects of polyphenols: A comparative review on the parental
464 compounds and their metabolites. *Food Reviews International*, 37(8), 759–811.
465 <https://doi.org/10.1080/87559129.2020.1717523>
- 466 Soto-Quintero, A., Guarrotxena, N., García, O., & Quijada-Garrido, I. (2019). Curcumin to
467 promote the synthesis of silver NPs and their self-assembly with a thermoresponsive polymer
468 in core-shell nanohybrids. *Scientific Reports*, 9(1), 1–14.
- 469 Sun, T.Y., Li, J.S., & Chen, C. (2015). Effects of blending wheatgrass juice on enhancing phenolic
470 compounds and antioxidant activities of traditional kombucha beverage. *Journal of Food and*
471 *Drug Analysis*, 23(4), 709–718.
- 472 Tahri, K., Tiebe, C., El Bari, N., Hübert, T., & Bouchikhi, B. (2016). Geographical provenience
473 differentiation and adulteration detection of cumin by means of electronic sensing systems
474 and SPME-GC-MS in combination with different chemometric approaches. *Analytical*
475 *Methods*, 8(42), 7638–7649.
- 476 Tokaç, M., Taner, G., Aydın, S., Özkardeş, A.B., Dündar, H.Z., Taşlipinar, M.Y., Arikök, A.T.,
477 Kiliç, M., Başaran, A.A., & Basaran, N. (2013). Protective effects of curcumin against
478 oxidative stress parameters and DNA damage in the livers and kidneys of rats with biliary
479 obstruction. *Food and Chemical Toxicology*, 61, 28–35.
- 480 Wang, Y., Ji, B., Wu, W., Wang, R., Yang, Z., Zhang, D., & Tian, W. (2014). Hepatoprotective
481 effects of kombucha tea: Identification of functional strains and quantification of functional
482 components. *Journal of the Science of Food and Agriculture*, 94(2), 265–272.
- 483 Zailani, N. S., & Adnan, A. (2022). Substrates and metabolic pathways in symbiotic culture of
484 bacteria and yeast (SCOBY) fermentation: A mini review. *Jurnal Teknologi*, 84(5), 155-165.
- 485 Zhao, Z. J., Sui, Y. C., Wu, H. W., Zhou, C. B., Hu, X. C., & Zhang, J. (2018). Flavour chemical

486 dynamics during fermentation of kombucha tea. *Emirates Journal of Food and Agriculture*,
487 30(9), 732–741.

488 Zheng, X., Ma, W., Sun, R., Yin, H., Lin, F., Liu, Y., Xu, W., & Zeng, H. (2018). Butaselen
489 prevents hepatocarcinogenesis and progression through inhibiting thioredoxin reductase
490 activity. *Redox Biology*, 14, 237–249.

491 Zubaidah, E., Nisak, Y.K., Wijayanti, S.A., & Christianty, R.A. (2021). Characteristic of
492 microbiological, chemical, and antibacterial activity of turmeric (*Curcuma longa*) kombucha.
493 *IOP Conference Series: Earth and Environmental Science*, 924, 012080.

494

495

496 **Tables**

497

498 **Table 1.** List of formulations to test the hepatoprotective property of turmeric kombucha and
 499 turmeric essence beverages

500

Treatment Group	Description
Control negative (P0)	Normal mice by feeding healthy mice/mice
Control positive (P1)	Mice Normal diet + DEN
P2	Mice non-DEN+ Turmeric essence beverage dose 0.3 mL/20 g BW
P3	Mice DEN + Turmeric essence beverage dose 0.1 mL/20 g BW
P4	Mice DEN + Turmeric essence beverage dose 0.3 mL/20 g BW
P5	Mice DEN + Turmeric essence beverage dose 0.5 mL/20 g BW
P6	Mice non-DEN + Turmeric kombucha dose 0.3 mL/20 g BW
P7	Mice DEN + Turmeric kombucha dose 0.1 mL/20 g BW
P8	Mice DEN + Turmeric kombucha dose 0.3 mL/20 g BW
P9	Mice DEN + Turmeric kombucha dose 0.5 mL/20 g BW

501

502

503

504 **Table 2.** Turmeric kombucha and turmeric essence beverage characteristics (Zubaidah et al.,
505 2021)

Parameter	Turmeric kombucha	Turmeric essence beverage
pH	3.8	7.4
Total titratable acid	1.24%	Not detected
Total phenolic content	137.28 mg GAE/mL	94.25 mg GAE/mL
IC ₅₀ antioxidant activity	76.16 ppm	106.59 ppm
Total microbial cells	2.70 x 10 ⁷ CFU/mL	Not detected

506

507 **Table 3.** Identified chemical compounds in turmeric kombucha and turmeric essence beverage
 508 using LC-MS

Component	MW (g/mol)	Retention time	Turmeric kombucha	Turmeric essence beverage	Benefit
Tetrahydrocurcumin(1,7-bis(4-hydroxy-3-methoxyphenyl)	372.4	2.563	√	N/D	Anti-inflammation, Anti-cancer and anti-bacterial
Ferulic acid (4-Hydroxy-3-methoxy cinnamic acid)	194.18	1.277	√	√	Antioxidant, anti-inflammation, apoptosis, and cardioprotective (Bezerra et al., 2017; Mattila & Kumpulainen, 2002)
Acetylsalicylic acid (2-acetyl benzoic acid)	180.31	2.563	√	N/D	Analgesic (Ashok & Upadhyaya, 2013)
Methoxyphenol	124.14	1.772	√	√	Anti-carcinogenic and antioxidant (Sun et al., 2015).
Eugenol	127.39	2.735	√	√	Anti-bacterial, antioxidant, and analgesic
Bisdemethoxycurcumin	308.3	6.127	√	√	Anti-inflammation and lower expression NF- κ B (Ramezani <i>et al.</i> , 2018).
Demethoxycurcumin	308.3	17.871	√	√	Anti-inflammation and anti-neoplastic (Hoehle et al., 2006)
Curcumin glucuronide	544.5	3.182	√	N/D	Immunosuppressive, antioxidant, anti-neoplastic, cytotoxic, anti-cancer, and anti-tumor (Ming et al., 2020)
Cyclofenil	364.4	1.565	√	√	Ovulation induction, infertility anti-virus (Sayed & El-Kordy, 2014), and inhibition of MCF cell proliferation in breast cancer (Mughal, 2019)
Acetic acid	60.05	14.183	√	N/D	Antioxidant, anti-microbial, toxicity (Jakubczyk et al., 2020; Jayabalan et al., 2014; Tahri et al., 2016), and anti-

D-saccharic acid-1,4-lactone (DSL)	192.12	18.721	√	N/D	inflammation (Bimonte et al., 2015) Antioxidant, anti-inflammation, and heart damage (Ireson et al., 2002), anti-diabetes, cytotoxic, hepatotoxic, and hepatoprotective (Wang et al., 2014)
Glucuronic acid	397.17	17.234	√	N/D	Antioxidant, hepatoprotective, and anti-inflammation (Martínez-Leal et al., 2020)
Carboxylic acid	477.4	3.182	√	N/D	Prevent liver damage (Rocha-Ramírez et al., 2017) and immunomodulator (Bauer-Petrovska & Petrushevska-Tozi, 2000)
Chloroacetyl-dl-phenylalanine	241.67	1.600	N/D	√	Bacterial xenobiotic metabolites (Aggarwal et al., 2013; Zhao et al., 2018)
Phenyl	364.4	2.356	√	√	Anti-microbial (Pereira et al., 2009), cardiovascular, and anti-cancer (Sim et al., 2019)
Pyrazine	979.0	2.735	√	√	Analgesic, anti-inflammation, antioxidant, anti-cancer, and anti-microbial
Quinazoline	1033.2	2.941	√	√	Anti-inflammation, anti-cancer, anti-inflammation, and anti-microbial (Bimonte et al., 2015; Hatcher et al., 2008)

509

510

511

512 **Table 4.** Hepatoprotective activity of turmeric kombucha and turmeric essence beverage

Treatment group	ALT (U/L)	AST (U/L)	MDA (nanomole/mL)
Normal diet	25.770 ^d ± 1.0	20.199 ^e ± 0,8	4.319 ^{ab} ± 0,7
Normal diet + DEN	41.147 ^a ± 0.4	40.739 ^a ± 1,1	5.292 ^a ± 0,8
Normal diet + turmeric essence beverage	20.445 ^e ± 1.2	21.347 ^d ± 1,3	3.812 ^b ± 0,3
DEN + dose 0.1 mL/20 g BW	32.510 ^b ± 0.4	31.958 ^b ± 3,7	4.322 ^{ab} ± 0,1
DEN +dose 0.3 mL/20 g BW	31.355 ^{bc} ± 0.4	31.107 ^{bc} ± 0,9	4.076 ^{ab} ± 0,6
DEN + dose 0.5 mL/20 g BW	30.451 ^c ± 1.1	30.077 ^c ± 3,3	4.032 ^{ab} ± 0,7
Normal diet + turmeric kombucha	20.884 ^e ± 0.8	20.454 ^{de} ± 0,5	3.858 ^{ab} ± 0,3
DEN + dose 0.1 mL/20 g BW	21.962 ^e ± 0.1	21.738 ^d ± 0,5	4.079 ^{ab} ± 0,2
DEN + dose 0.3 mL/20 g BW	21.040 ^e ± 0.8	21.341 ^{de} ± 1,2	3.807 ^b ± 0,4
DEN + dose 0.5 mL/20 g BW	20.851 ^e ± 0.8	20.110 ^e ± 0,3	3.761 ^b ± 0,1

513 Note: ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; MDA: Malonaldehyde;
514 different notations show a real difference ($\alpha=0.05$); data obtained from the average of 3
515 replications ± SD

516
517

518 **Table 5.** Total cell damage

	Treatment	Total of dead cells
	Normal diet	17 ^c ± 0.8
	Normal diet + DEN	49 ^a ± 0.8
	Normal diet + Turmeric Essence Beverage	16 ^e ± 1.2
	DEN + dose 0.1 mL/20 g BW	32 ^b ± 1.6
	DEN + dose 0.3 mL/20 g BW	30 ^{bc} ± 0.8
	DEN + dose 0.5 mL/20 g BW	28 ^{cd} ± 0.5
	Normal diet + Turmeric kombucha	15 ^e ± 0.5
	DEN + dose 0.1 mL/20 g BW	30 ^b ± 0.9
	DEN + dose 0.3 mL/20 g BW	27 ^d ± 1.2
	DEN + dose 0.5 mL/20 g BW	26 ^d ± 0.5

519 Note: Different notations show a real difference ($\alpha=0.05$); data obtained from the average of three
 520 replications \pm SD

521

522 **Figure legend**

523

524 **Fig. 1.** Experimental animal treatment procedures, modified (Zheng et al., 2018)

525

526 **Figure 2.** Mice liver histology

527 (a) Normal diet; (b) Normal diet + DEN; (c) Normal diet + turmeric essence beverage; (d) DEN +

528 0.1 mL/g turmeric essence beverage; (e) DEN + 0.3 mL/g turmeric essence beverage; (f) DEN +

529 0.5 mL/g turmeric essence beverage; (g) Normal diet + turmeric kombucha; (h) DEN + 0.1 mL/g

530 turmeric kombucha; (i) DEN + 0.3 mL/g turmeric kombucha; (j) DEN + 0.5 mL/g turmeric

531 kombucha; magnification 400x

532 I: inflammation; A: apoptotic body; F: fibrosis; CV: central vein

533

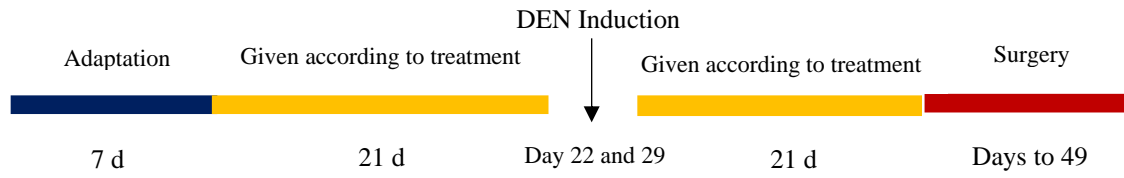
534

535 **Figures (colors should be used)**

536

537 **Fig. 1.**

538



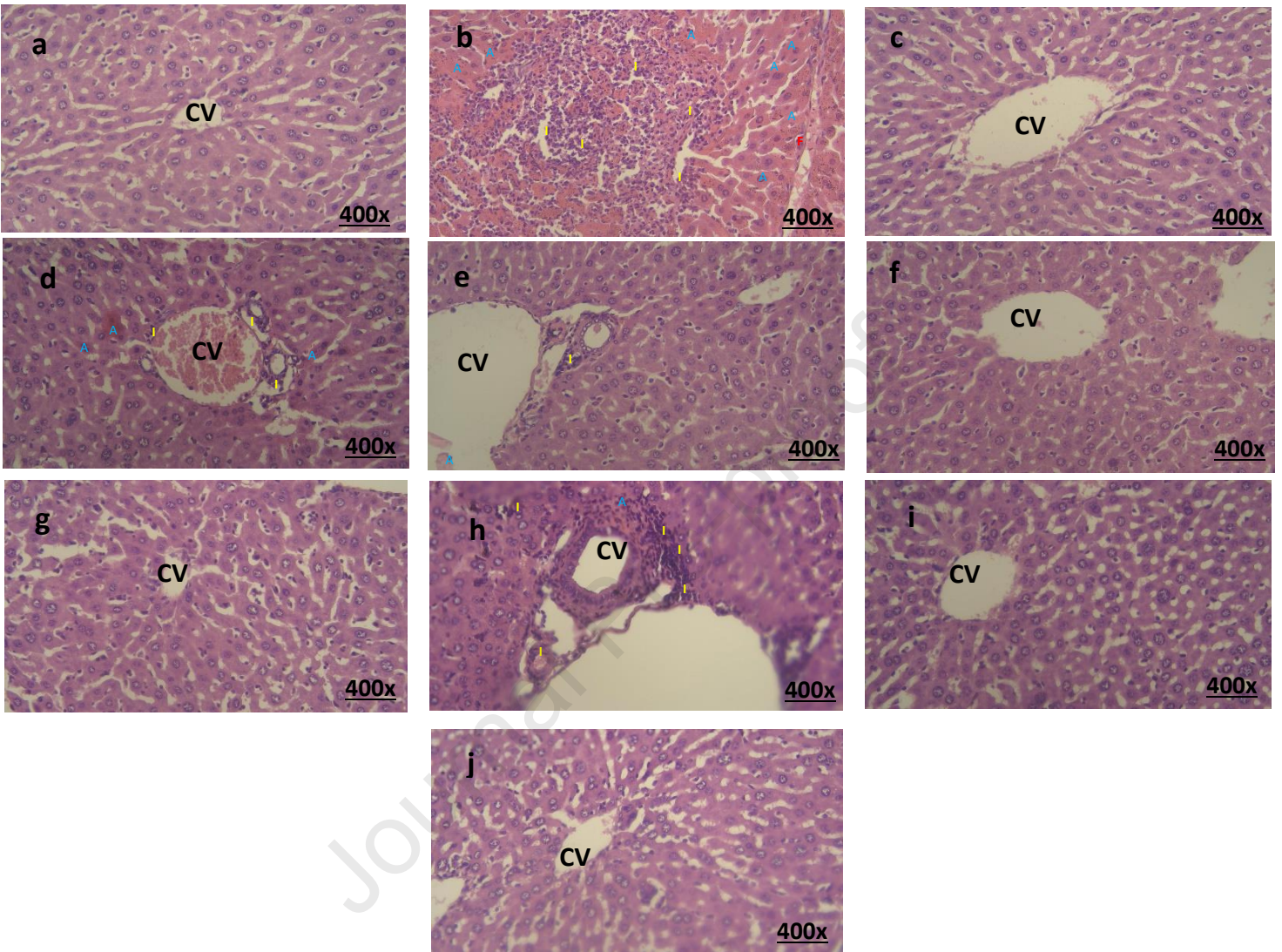
539

540

Journal Pre-proof

541 **Fig. 2.**

542



Author statement

We hereby declare that all the authors of “The distinctive hepatoprotective activity of turmeric kombucha (*Curcuma longa*) induced by diethylnitrosamine in Balb/C mice” have approved the newly revised manuscript to be re-submitted to Food Bioscience. There are no conflicts of interests.

Journal Pre-proof

Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

Journal Pre-proof