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# Green tea polyphenols in cardiometabolic health: A critical appraisal on Phytogenomics towards personalized green tea

Logesh Rajan<sup>1#</sup>, Arun Radhakrishnan<sup>2#</sup>, Keshav Narayan Alagarsamy<sup>3</sup>, Abhay Srivastava<sup>3</sup>, Sanjiv Dhingra<sup>3</sup>, Anthony Booker<sup>4</sup>, Viven Rolfe<sup>5</sup>, Dhanabal Palaniswamy<sup>1</sup> and Suresh Kumar Mohankumar<sup>6\*</sup>

- 1. TIFAC CORE in Herbal Drugs, Department of Pharmacognosy, JSS College of Pharmacy, JSS Academy of Higher Education & Research, Ooty, Nilgiris, Tamilnadu, India.
- Department of Pharmaceutics, JSS College of Pharmacy, JSS Academy of Higher Education & Research, Ooty, Nilgiris, Tamilnadu, India.
- Regenerative Medicine Program, Institute of Cardiovascular Sciences, St. Boniface Hospital Albrechtsen Research Centre, Department of Physiology and Pathophysiology, Max Rady College of Medicine, Rady Faculty of Health Sciences, University of Manitoba, MB R3T 2N2, Winnipeg, Canada.
- 4. Research Centre for Optimal Health, School of Life Sciences, College of Liberal Arts and Sciences, University of Westminster, London, United Kingdom
- 5. Pukka herbs Ltd, Block C, The chocolate factory, Keynsham, BS31 2GN, United Kingdom.
- Swansea University Medical School, Institute of Life Sciences, Swansea University, Singleton Park, Swansea, Wales SA2 8PP, United Kingdom.

#Authors contributed equally

# \* Corresponding author

### Suresh K. Mohankumar, M.Pharm, Ph.D, PGCHE, FHEA

Associate Professor, Pharmacy-Pharmacology Swansea University Medical School Institute of Life Sciences Swansea University Singleton Park Swansea Wales SA2 8PP Phone +44 07760 164415 Mail id: s.k.mohankumar@swansea.ac.uk

#### Author contributions

Logesh Rajan#: Content writing & data collection Arun Radhakrishnan#: Content writing & data collection Keshav Narayan Alagarsamy: Literature review and content writing Abhay Srivastava: Literature review and draft preparation Sanjiv Dhingra: Content design & development Anthony Booker: Content development and editing Viven Rolfe: Content development and editing Dhanabal Palaniswamy: Content review and editing Suresh Kumar Mohankumar: Framework planning, Concept development, Guidance

#### Abstract

Cardiovascular disease is a chronic multifactorial health complication that is either directly or indirectly associated with pathophysiological mechanisms, including pro-oxidation, proinflammation, vascular and endothelial dysfunction, impaired platelet function, thrombosis, and others. The therapeutic options to circumvent cardiovascular complications include several phytomedicines, including green tea polyphenols. However, while many experimental and clinical studies report distinct mechanisms by which the polyphenols of green tea elicit a beneficial role in cardiometabolic health, the translation and applications of green tea polyphenols in clinics have yet to gain their optimal use on the broader population. This review critically appraises the various reported mechanisms of green tea polyphenols in modulating cardio-metabolic health and associated phyto-genomic challenges. Further, our review highlights the probability of gene polymorphic associated therapeutic variations in individuals using green tea for cardio-metabolic effects and the necessity to personalize green tea for clinical use, thereby improvising the risk-benefit ratio.

#### **Keywords:**

Green tea polyphenols, cardiovascular disease, inter-individual variability, genetic Polymorphism, phyto-genomic.

#### Abbreviations

- ACE Angiotensin-converting enzyme
- ADMA Asymmetric dimethylarginine
- ADP Adenosine diphosphate
- ADSC Adipose-derived stem cells
- AGEs Advanced glycation end products
- BMI Body mass index
- BW Body weight
- CAD Coronary artery disease
- COMT Catechol-O-methyltransferase
- CRB Catechins rich beverage
- CRP C-reactive protein
- CVDs Cardiovascular disease
- DBP Diastolic blood pressure
- ECs Endothelial cells
- EDHFs-Endothelium-derived hyperpolarizing factors
- EDRF Endothelium-derived relaxing factor
- EGC Epigallo catechin
- EGCG Epigallocatechin gallate
- eNOS Endothelial nitric oxide synthase
- FBF Forearm blood flow
- FMD Fibromuscular dysplasia
- GTC Green tea catechins
- GTE Green tea extract
- HDLC-High-density lipoproteins cholesterol
- HOMA-IR-Homeostasis model assessment of insulin resistance
- ICAM-1 Intercellular adhesion molecule-1
- IEs Idiopathic environmental intolerances
- IHD Ischemic heart disease
- iNOS Inducible nitric oxide synthase
- NOS Nitric oxide synthase

- LDL Low density lipoprotein
- LDL-C-Low density lipoprotein cholesterol
- MCP-1 Monocyte chemo attractant protein-1
- MC-SF Macrophage colony-stimulating factor
- MMPs Matrix metalloproteinases
- NO Nitric oxide
- PDE-5 Phosphodiesterase type 5
- PDGF Platelet-derived growth factor
- PI3-K Phosphatidylinositol 3-kinase
- PKC Protein kinase-C
- PL Phospholipids
- RCM Restrictive cardiomyopathy
- ROS Reactive oxygen species
- SBP Systolic blood pressure
- SNP Single nucleotide polymorphisms
- TAS Total anti-oxidant status
- TC Total Cholesterol
- TG Triglycerides
- TNF-α Tumor necrosis factor
- TXA2 Thromboxane A2
- VCAM-1-Vascular cell adhesion molecule-1
- VED Vascular endothelial dysfunction
- VSMCs Vascular smooth muscle cells
- WC Waist circumference

#### Introduction

Cardiovascular disease (CVD) is a chronic severe multifactorial disease that adversely affects an individual's health, well-being, and lifestyle [1]. The World Health Organisation (WHO) projected cardiovascular diseases (CVDs) as the primary cause of death globally. In 2019, around 17.9 million (32%) people died due to CVDs, and it is estimated to increase exponentially in the future [2-5]. Thus, preventing the risk of CVD is a timely task. Plant-based

bioactivities hold enormous interest in a safe alternative among various therapeutic strategies to combat CVD.

Since ancient times, tea has been consumed as a common drink by humans worldwide. The tea plant *Camellia sinensis* (L.) Kuntze Theaceae, is commonly known as tea, and it likely originates from South-West China, Indo-Burma, Tibet region, having been domesticated from the wild Camellia taliensis (W.W.Sm) Melch [6, 7]. While tea has several varieties, green tea has gained immense therapeutic interest because of its potent anti-oxidant, anti-inflammatory, anti-hypertensive, anti-diabetic, and other valuable activities [8, 9]. The primary chemical constituents of green tea are catechins, consisting of 80-90% of the total polyphenols. Among the various catechins reported in green tea, the epigallocatechin gallate (EGCG) constitutes more than 50% of total catechins. The polyphenols of green tea have shown significant cardioprotective effects under experimental conditions in cell lines, animal models, and clinical studies [10-15], suggesting its therapeutic potential in combating CVD. Of note, many epidemiological studies report that regular consumption of green tea catechins (GTC) reduces the risk of CVD [16-22]. However, despite the several pieces of evidence supporting GTC's therapeutic implications in CVD, the clinical outcomes of GTC remain mixed.

Emerging reports indicate that the effects of GTC vary with individuals due to ethnic, gender, lifestyle, cultural, environmental, and genetic differences [23-25]. This was further justified by various researchers by their experimental pieces of evidence. In 2015 a Japan-based prospective study revealed that lifestyle change, i.e., smoking, dietary habits, etc., can alter the effect of green tea in CVD management. Later in 2016, green tea failed to establish its ability to control hyperlipidemia in a clinical trial on the Chinese population[26]. However, in the same year, a clinical trial conducted on the American hyperlipidemic population established positive corelation, which indicates the region, race associated altered effect of green tea. Inter-ethnic related differences in the pharmacokinetic profile of green tea were also explained in a systematic literature study carried out in 2021[27]. Apart from these positive outcomes, the frequency and degree of ADRs also change according to the individual variation. Even though the number of ADRs was associated with green tea consumption, liver injury is considered significant adverse effects. A systematic review on green tea supplements revealed the hepatotoxic effect of green tea and it further concluded that the adverse effect is majorly based

on patient factors [28]. A randomized, placebo-controlled, double-blinded phase II clinical trial studied the adverse effects of sustained high dose oral green tea extract. The study strongly indicated the effect of green tea extract in liver enzyme elevation [28]. The European Food Safety Association (EFSA] Panel on Food Additives and Nutrient Sources added to Food suggested that the intake of EGCG equal to or above 800mg/day may increase the serum transaminases [29]. Likewise, few published reports indicate the population-based hepatotoxic nature of green tea. Despite these ADRs, several scientific studies reported GTC consumption's health benefits; the possible associations with gene polymorphisms were less addressed [30]. A few ancillary analysis studies suggested that the selective genes such as *SLCO1B1\*1B* (rs4149056), *UGT1A1* (rs8175347), *SOD1A251G* (rs2070424), and others could influence the molecular mechanisms by which GTC elicit cardio-metabolic effects [31, 32]. Thus, it is pivotal to understand the specific genetic polymorphisms and their association with cardiovascular benefits of GTC.

Collectively, this review collates the effective mechanisms associated with cardio-metabolic health benefits of GTC and subsequently appraises the genomic factors that affect the therapeutic responses of GTCs. These genes are selectively proposed to alter the dynamics and kinetics of GTC in humans, based on the population pharmacokinetic data constructed from conventional drugs, which exhibit their action via similar pathways [33, 34].

### GTCs-mediated cardiometabolic health

#### Conventional cardioprotective action

Many human clinical trials and cohort studies show the association between green tea consumption to CVD health [35-39]. For example, a Japanese elderly cohort study with the patient population of 14001 reported that intake of green tea of about five or more cups daily significantly reduced CVD mortality [40, 41]. Additionally, various cohort studies carried out in China and Japan further strongly explained the positive effect of green tea on CVD [42]. However, the risk of CAD was reduced only in males, whereas it was not observed in female participants [43]. Likewise, multiple clinical studies showed that consuming green tea flavonoids significantly decreased endothelium-dependent vascular reactivity and CVD parameters [44-46].

On the contrary, some studies on specific people have also shown that green tea fails to meet the expected cardio-related positive effects. In **table 1** population-based positive, negative co-relation and the adverse effects of green tea towards various cardio-parameters were extensively reviewed.

Table 1. Pop	oulation-based	studies on green	tea consumptions	s and its cardiom	etabolic outcomes.

Co-relation type	Objective	Results& Conclusion	Population	Year
Positive correlation Tea consumption	BP, cholesterol, coronary and total mortality	Decreased SBP, coronary mortality rate	Norway (9,856 men and 10,233 women)	1992 [47]
effects on	Endothelial disfunction	Reversed endothelial dysfunction.	USA (coronary artery disease patients)	2001 [48]
	BP, serum lipids, oxidative stress, and inflammatory	Significant decrease in LDL c, HDL c.	China (hypercholesterol emic patients)	2003 [49]
S	4-O-methylgallic acid and hypertension	Long term intake of tea has a favorable effect on BP Reduction in SBP and DBP	Australia (greater than 70 years old women)	2003 [50]
	Hypertension	Reduction in the risk of developing hypertension	China (711 men and 796 women with no hypertension)	2004 [51]
	Anti-oxidant activity, oxidative resistance to DNA, and lipid profile	GT improved overall antioxidative status	Italy	2005 [52]
	Postprandial lipid response	Catechins reduce TG level after fat loaded	Japan (subjects with hyper	2005 [53]

		food consumption	triacylglycerolae mia)	
	Insulin resistance and systemic inflammation	A slight decrease in BW, SBP and DBP, blood glucose, hemoglobin A1C, and insulin levels	Japan (53 males and 13 females with mild diabetes)	2005 [54]
	Endothelial function and endothelial progenitor cells in smokers	Increase in FMD and endothelial progenitor cells, prevent the cardiovascular events in chronic smokers	Korea (20 young healthy smokers)	2006 [55]
	Endothelial dysfunction	The level of O- methylase affects the improvement of FMG	Australia	2006 [56]
	Endothelial dysfunction and blood pressure	Flavonoid supplementation improved endothelial function.		2006 [57]
	Oxidized plasma LDL and lipid profile	Significantly decreased oxidized LDL without change in plasma LDL	Japan (10 men, 30 women)	2007 [58]
	Cardiovascular risk and body fat reduction	Reduced BW, BMI, body fat ratio, WC, body fat mass, SBP, and LDL.	Japan (women and men with visceral fat-type obesity)	2007 [59]
S	Susceptibility of LDL oxidation	Decreased LDL, prevent atherosclerosis progression	Japan	2008 [60]

Nitric oxide and endothelin 1 Acute and chronic administration of high doses (580mg)	Enhancing the NO and decreasing the ET-1 plasma concentrations Improvement in the forearm blood flow in male smokers	Australia (Healthy men)	2008 [61]
Endothelial function [Flow-mediated vasodilation)	Increased FMD, Acute beneficial effect on endothelial function	Greece (Male)	2008 [62]
Vascular function and lipid peroxidation	Significant increase in brachial artery diameter, reduction in oxidized LDL and anti-oxidized LDL IgM antibodies level	Spain (Female)	2008 [63]
Type 2 diabetes patients	Decreased WC and increased Adiponectin, useful in reducing obesity.	Japan (Patients with type 2 diabetes)	2009 (64]
BW, lipids and lipids peroxidation	Green tea improved the metabolic syndrome in obese patients	USA (35 subjects with obesity and metabolic syndrome)	2010 [65]
Evaluate the effects of green tea on cardiovascular disorder markers	Anti-atherosclerotic action on dysfunctional vessels in smokers	Japan (smokers)	2010 [66]
TC and LDL and HDl	Reduced LDL cholesterol	USA	2011 [67]
LDL, glucose and hormone level in post- menopausal women	Significant decrease in LDL.	USA (post- menopausal women)	2012 [68]

	Insulin resistance and cardiovascular risks in obese hypertensive subjects	GTE influences blood pressure, TC, LDL-C, HOMA-IR, CRP, TNF- $\alpha$ , and TAS	Poland (obese, hypertensive patients)	2012 [69]
	Prevention of CVD	Black tea: Reduced LDL, SBP, and DBP Green tea: Reduced TC	UK	2013 [70]
	Insulin resistance and glucagon-like peptide 1 in type 2 diabetes patients	Decrease in insulin resistance and TG and HOMA-IR index. Increased HDL	China	2014 [71]
	Weight reduction, lipid profile, and hormone peptides (obesity-related]	Significant reduction in BW reduced LDL and TC	China (women with central obesity]	2015 [72]
	Ischemic heart disease (IHD) and cancer	Reduced risk of IHD and major coronary events	China	2017 [73]
	CVD	Reduced BP, LDL, support vascular functioning and reduced inflammation	UK	2018 [74]
	Dose response relation between green tea consumption and risk of CVDs	4% lower risk of stroke and CVD mortality, 2% lower risk of CVD events	USA	2020 [75)
S	Effect of green tea consumption on blood pressure	Significant reduction in SBP and DBP	China	2020 [76]
Negative correlation	Arterial stiffness and inflammation in type 2 diabetes patients	No reduction in inflammatory, lipid profiles, blood glucose, insulin resistance	South Korea	2005 [77]
Tea consumption effects on	Inhibition of platelet aggregation, LDL oxidation, and matrix metalloproteinases (MMPs) activity	No reduction in the concentration of MMPs, urine 8-epi- PGF(2alpha), and platelet aggregation	Japan	2005 [78]

	Blood parameters and metabolic parameters when consumed with low energy diet	Modest weight loss	Netherlands	2006 [79]
	Body fat and CVD and its safety in obese children	No significant weight loss. Genetic variation, dietary habits, and body composition alters the results	Eastern and western population	2008 [80]
	Obese women and related hormones	No significant reduction in BW, BMI, and WC	Taiwan (Obese women)	2008 [81]
	Insulin resistance and associated metabolic risk factors	No significant effect on insulin resistance. A modest reduction in diastolic BP	UK (obese patients: 40-65 years)	2009 [82]
	Serum lipids	No significant LDL lowering	Netherlands (67 men and 35 women)	2010 [83]
	CVD	No anti-inflammatory, anti-oxidant, antiproliferative effects.	USA	2011 [84]
	Obese, type 2 diabetic patients (EGCG 856 mg)	No significant difference between the placebo and treatment group in EC and insulin level	Taiwan	2011 [85]
Adverse effects Tea consumption effects on	TC, LDL, and HDL	Significant reduction in TC and LDL Adverse effects: Skin rashes, abdominal bloating, and gastric upset	China (1136 subjects)	2011 [86]
	Cholesterol	Visceral fat loss Adverse effects: Gastrointestinal symptoms (Bloating,	China (Adults)	2012 [87]

	change)		
Blood pressure	Reduced SBP, TC, and LDL Adverse effects: High doses are associated with side effects	UK	2014 [71]
Insulin resistance and glucagon-like peptides in type II diabetes Mellitus and lipid abnormality patients	Improvedinsulindecreasedthe level ofglucagon-likepeptide 1.Adverseeffects:Epigastricdullnessmildconstipation	China (92 subjects)	2014 [88]
Weight loss	No significant weight reduction. Adverse effects: Mild to moderate hypertension and constipation	Canada	2014 [89]
Blood lipids	Reduced LDL and TC.No reduction in HDLAdverseeffects:Abdominalbloating,gastricupsetandskinrashes	China	2020 [90]

These reported data show that green tea fails to reach its positive claim in some studies and, up to some extent, leads to adverse effects. Also, multiple studies reported that regardless of age, gender, sex, etc., genetic predisposition plays a significant role in inter-individual variability. Extensive studies were carried out by analyzing genes irrespective of age, gender, sex, etc. Genetic predisposition plays a vital role in inter-individual variability. Comprehensive studies were carried out by analyzing gene age, gender, sex, etc. Genetic predisposition plays a significant role in inter-individual variability. Extensive studies were carried out by analyzing gene age, gender, sex, etc. Genetic predisposition plays a significant role in inter-individual variability. Extensive studies were carried out by analyzing gene age, gender, sex, etc. Genetic predisposition plays a significant role in inter-individual variability. Extensive studies were carried out by analyzing gene age, gender, sex, etc. Genetic predisposition plays a significant role in inter-individual variability. Extensive studies were carried out by analyzing gene-related population changes with positive results [91]. An individual's genetic makeup

remains a constant variable throughout their lifespan and shows a potential impact on kinetic profile [92]. Even though various factors determine drug efficacy, genetic predisposition is considered feasible, accurate, and highly influencing compared to other factors' heterogeneity among individuals [93]. Based on the extensive referred reports and shreds of evidence, it was firmly believed that green tea exhibits significant inter-individual variability on cardio-metabolic health. These individual variabilities may result in an unexpected effect that may or may not benefit a person already at risk of cardiac health. In association with many inconsistent reported clinical interventions, the "one size fits for all" approach is a failed strategy in green tea consumption. Therefore, this helps us focus more on inter-individual variability, which can predict the efficacy of green tea usage in the treatment of cardio-metabolic disorders. This further paved the path of research towards molecular mechanism based genetic Polymorphism associated inter-individual variability

# Molecular mechanisms by which GTC combats cardiometabolic diseases

S. No.	Mechanism	Activity	Reference
1.	Anti-oxidant effects	• Increased anti-oxidant enzymes	Dhalla et al., 2000[94];
		decreased pro-oxidant enzymes	Paravicini and Touyz,
		• Free radicals scavenging	2008[95]; Park et al.,
		• Chelation of redox ions	2011[96]; Zinkevich and
		• Decreased OxLDL and NOX-1	Gutterman, 2011[97]; A
	3	enzymes	Islam, 2012[98]
2.	Anti-inflammatory	• Decreased cytokines,	Marui et al., 1993[99];
	activity	chemokines, VCAM-1,	Lin and Lin, 1997[100];
		adhesion molecules, MCP-1	Gu et al., 1998 [101];
		expression, leucocyte	Gerszten et al.,
		penetration, p38 MAPK, and	1999[102]; Kevil et al.,
		NF-kappa B	2001 [103]; Ludwig et
			al., 2004 [104]; Libby,
			2006[105]; Suzuki et al.,

 Table 2. Distinct mechanisms of GTCs to combat CVD.

			2007[106]; A Islam,
			2012 [39]
3.	Vascular	• Increased NO, eNOS, cGMP,	Dimmeler et al.,
	endothelial	PI3K/Akt and ET-1 activation	1999[107]; Schiffrin,
	dysfunction		2001 [108]; Galley and
			Webster, 2004[109];
			Kobayashi et al.,
			2005[110]; Hadi and
			Carr, 2005[111];
			Forstermann and
		C)	Munzel, 2006[112];
			Basu and Lucas, 2007
			[113]; Balakumar et al.,
			2009[114]; Islam,
			2012[98]
			D 1 01
4.	Anti-proliferative	• Decreased VSMCs	Ross and Glomset,
	activity	proliferation, PDGF expression,	1973[115]; Lindner and
	(	degradation of matrix protein,	Reidy, 1991[116];
		MMP-2 and MMP-9 expression	Schwartz et al.,
		•	1995[117]; Yamamoto
			et al., 2000[118]; Visse
			and Nagase, 2003 [119];
			Ouyang et al.,
			2004[120]; De Donatis
			et al., 2008[121]; Yuan
			et al., 2011[122]; Yang
			et al., 2011[123]; A
			Islam, 2012[98]
5.	Anti-platelet and	• Decreased Ca <sup>2+</sup> utilization	Born, 1965 [124]: Kang
	anti-thrombotic	inositol 1 4 5-triphosphate	et al 2001 [125]:
	activity	fibrinogen-GPIIh/IIIa hinding	Ueno et al
		nhosnholinase Cv2	2011[126]: A
			2011[120], 11

		phosphorylation, protein	Islam, 2012[98]
		tyrosine phosphorylation,	
		arachidonic acid, and cellular	
		prostaglandin D2 levels	
6.	Reduce the	• Significant down regulation of	Luo K et al., 2020 [127]
	production of	Apolipoprotein B and	
	VLDL and induce	microsomal triglyceride transfer	
	LDL expression	protein reduce VLDL	<u>S</u>
		Activate peroxisome	
		proliferator-activated receptor $\delta$	
		and regulatory element-binding	
		protein 2 thereby increase the	
		level of LDL c	
7.	Reduction of	Targeting Low-Density	Pamela Mason et al.,
	cholesterol	Lipoprotein Receptor and down	2021 [128]
	absorption from	regulating Microsomal	
	intestine	Triglyceride Transfer Protein	

# Genomic factors responsible for the inter-individual variability

Genetic polymorphism and altered activity in the human body differentiate individuals' drug responses. Any polymorphism that influences the mediators in a drug action pathway or the drug-metabolizing enzymes determines the drug response. The inter-individual variability constructed the path towards genomic variation-based drug action science. The concept of pharmacogenomics has existed since ancient times. The Greek philosopher Pythagoras reported that only a micro-set of people suffered from fatal hemolytic anemia [129]. Later, modernization of science-supported his claim by identifying genetic polymorphism associated with glucose-6-phosphate dehydrogenase. This initiation, along with the human genome project, widened the scope of pharmacogenomics in medical research.

Pharmacogenomic factors are regarded as critical in cardiovascular therapy. For instance, one of the metabolizing enzymes, *CYP2C19*, was 40% low functional in the Hawaiian population. Such polymorphisms influenced clopidogrel's therapeutic outcome in the Hawaiian population [130].

Most cardiometabolic medicines are predominantly affected by ABCB1 [efflux pump) polymorphism, specifically C3435T, which results in an average 72% risk when comparing individuals with CT/CC genotype. [131-134]. In addition, various other genes *VKORC1*, *CYP2C19\*2*, *CYP3A4*, *HMGCR13*, *SLCO1B1*, *NEDD4L*, *CACNA1C*, *CACNB2*, and *KCNMB1*, were related to change in cardiovascular drug PK-PD profile [135]. Figure 1 shows the association of molecular mechanisms of green tea polyphenols with genetic polymorphism-linked inter-individual variations in protecting cardio-metabolic health.



Figure 1. Association of molecular mechanisms of green tea polyphenols with genetic polymorphism-linked inter-individual variations in protecting cardio-metabolic health (insert here)

Some direct correlation studies on green tea catechins pharmacogenomic profiles were performed on various populations, which provided a favorable report on inter-individual variability. Investigation revealed that the individuals with polymorphisms in the gene T allele of *MRP2 -24C* present in the *ABCC2* gene reduced the efficacy of EGC. Bruhn and co-workers studied the polymorphisms of drug transporters ABCB1, ABCG2, ABCC2, and ABCC3 and

their impact on drug bioavailability with respect to the clinical relevance of EGC [136]. It was also found that SLCO1B1 gene coding for OATP1B1 has been involved in EGCG and EGC clearance. There was a 30% reduction in clearance for homozygous individuals with the C allele of SLCO1B1 521T>C SLCO1B1\*5 and a 35% reduction in wild-type clearance when compared to the SLCO1B1 388A>G allele. Konig et al., reported that the clearance of EGC was reduced in the wild type compared to the variant allele SLCO1B1\*1B found in individuals [137]. Gong and his co-authors worked on the impact of genetic variation in OATP transporters on EGC disposition and response [138]. While considering the phase II enzymes in the biotransformation of green tea catechins SULT1A1, COMT, UGT1A1, there was less impact on the kinetics of EGC. Twenty-four human volunteers, 14 women and 10 men with COMT SNPs, were studied in Berlin to determine the effect of EGCG on COMT. 24% lesser bioavailability of EGC was identified with individuals having low activity genotype Met/Met of COMT. 26% clearance reduction in the UGT1A1(rs8175347) wild-type carriers were identified. The activity of catechol-O-methyltransferase was not impaired by high doses of epigallocatechin-3- gallate EGCG in vivo [139]. In a randomized controlled cross-over trial, the influence of COMT Val/Met genotype was observed. The homozygous group with high activity G allele had a lesser urinary accumulation of EGC and 4'-O-methyl EGC. In contrast, the individuals with a minimum of one low activity A allele had a Higher EGC and 4'-O-methyl EGC in urine [140]. A population-based case-control study performed on Asians from Los Angeles suggested the COMT genotype influences the change in green tea's polyphenol bioavailability. The results revealed that the low-activity COMT genotype carriers excreted fewer polyphenols in urine, implying that they may retain more tea polyphenols in their bodies and get more health advantages[141, 142]. On the other side, a study conducted in the United Kingdom with twenty participants recruited 10 of each homozygous COMT genotype. A preliminary investigation of the impact of COMT genotype on the absorption and metabolism of green tea catechins was studied on the population and demonstrated no significant variation in absorption and elimination of EGCG in COMT polymorphism [143]. A population nutrikinetic study conducted in German people, 84 healthy volunteers on the effect of EGCG and EGC, indicated the use of oral contraceptives and inherent genetic variations in MRP2 and OATP1B1 impact their pharmacokinetics [144]. Whereas the genes MRP2, MRP2, ABCB, ABCC, and ABCG are related to a transmembrane protein specifically involved in the efflux of drug molecules, including EGCG and EGC. Hence,

polymorphism in this protein will enhance the elimination rate and thereby reduce the efficacy of green tea. On the other hand, the genes *OATP1B1* and *SLCO1B* are related to influx protein, which takes the drug for hepatic metabolism. Polymorphism in these genes can alter the pharmacokinetic profile of EGCG and EGC [145]. Similarly, various studies have suggested significant pathways involved in the metabolism of catechins are glucuronidation (4''-O-glucuronidation, 3'-O-glucuronidation, 7-O-glucuronidation), sulfation (O-sulfation), o-methylation (4'-O-methylation, O-methylation), and ester hydrolysis. The isozymes involved in the enzymatic hydrolysis are SULT1A1, SULT1A3, UGT1A8, UGT1A9, UGT1A3. Polymorphism in these enzyme-related genes may also alter the pharmacokinetics of catechins [146].

The significant genetic polymorphism involved in altering the effect of GTCs is shown in **table 3**.

S. No	Genetic	Drug	Effect	Year	Reference
	Polymorphism		·		
1.	SLCO1B1	EGCG and EGC	Involvement in clearance	2000	König et al., 2000[137]
2.	MRP1, MRP2 and Pgp	Green tea catechin	Involvement in the absorption and excretion of GTCs	2008	Kadowaki et al., 2008[147]
3.	COMT	EGCG	Not impaired by high doses of EGCG	2010	Lorenz et al., 2014[139]
4.	COMT	tea polyphenols	Influence in bioavailability.	2010	Inoue-Choi et al., 2010 [39]
5.	SLCO1B1*1B	EGC	Reduced clearance	2012	Gong and Kim, 2012 [138]
6.	COMT SNPs	EGC, EGCG	Reduced bioavailability	2010	Inoue-Choi et al., [141]

Table 3. Genetic Polymorphism	involved in GTCs.
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7.	MRP2-24C, ABCC2,	EGC	Reduced efficacy of	2014	Bruhn and
	ABCB1, ABCG2,		EGC		Cascorbi,
	ABCC2 and ABCC3				2014[136]
8.	MRP2 and OATP1B1	EGCG and	Impact in	2018	Scholl et
		EGC	pharmacokinetics		al.,
					2018[144]

At present surplus, clinical-cum-pharmacogenomic information is available on FDA-approved synthetic molecules. Based on this evidence, we are hypothesizing the EGCG associated cardiometabolic genetic polymorphism. EGCG action in the body happens through NOS, CAT, and GPX. EGCG inhibits inducible nitric oxide synthase (iNOS), Lipoxygenase, cyclooxygenase, xanthine oxidase, etc. Hence the SNPs in these components affect the activity of EGCG in an individual.

The induction of NOS is one of the mechanisms by which EGCG produces its anti-oxidant activity. A cohort study conducted in 2015 reported the influence of nitric oxide synthase NOS polymorphisms NOS2A -2.5 kb(CCTTT)<sub>n</sub>, Ser608Leu and NOS3 -786(T>C) of nitrite/nitrate levels in inflammatory disorders accounting to 107 multiple chemical sensitivity, 89 fibromyalgia/chronic fatigue syndrome, 108 suspected multiple chemical sensitivity patients and 196 healthy subjects. NOS3 -786(TT)was associated with higher nitrite/nitrate levels in study subjects with idiopathic environmental intolerances IEs. In a study performed in 2017, polymorphic effects of NOS and nitric oxide levels in chronic periodontitis, rather than the polymorphic variations, gender was the contributing factor for the differences in nitric oxide concentration. Concomitantly in Brazilian individuals, SNPs in -1026(AC) and +2087(AG)SNPs had a significantly higher level of nitric oxide but were not associated with chronic periodontitis (148]. These polymorphisms determine the levels of NOS in individuals, hence the effect of a drug acting through NOS mechanisms. Another enzyme involved in the pathway of EGCG is Lipoxygenase. Lipoxygenase catalyzes the conversion of arachidonic acid to leukotriene A4. Polymorphisms in Lipoxygenase extensively influence the action of Leukotriene receptor antagonists and leukotriene synthesis inhibitors[149]. Around 2002, US-based researchers came up with the concept, ser529 gene polymorphic COX-1, PLA1/A2 polymorphic gene-related glycoprotein GP Ia/IIa may result in the maximized chance of resistance in aspirin action and

thereby enhanced ischemic cardiac events [150]. EGCG produces its anti-oxidant effects, reduces oxidative stress and inflammation by inhibiting the enzymes such as inducible nitric oxide synthase (iNOS), Lipoxygenase, cyclooxygenase, xanthine oxidase, etc and thereby supporting cardio-health. Pharmacokinetically higher the inhibition better the action, but the above discussed pharmacogenomic variation will affect EGCG binding affinity which further results in inter-individual variations in EGCG action. Similarly, in a clinical trial it was found that the variability in drug response is also associated with non-steroidal anti-inflammatory drugs due to COX-2 polymorphisms. In 2017, Lee et. al. found celecoxib produced inhibitory action in 2h in all subjects, the area under the curve was significantly less for rs689466 GG genotype compared to other groups [151]. SOD, GPX, and CAT are the major anti-oxidant enzymes and the anti-oxidant activity of EGCG is through the induction of these enzymes. SOD1 A251G(rs2070424) polymorphism was observed in 56.3% of females and 43.7% males in a study of 494 Turkish subjects in 2017 [152]. The association of CAT enzyme expression during a pathological condition depends on various factors such as TNF-a, PPARy, CpG islands hypermethylation, and p53 protein. Polymorphisms in the CAT gene contribute to variation in the enzyme levels and activity during pathological conditions, such as asthma, insulin resistance, hypertension, dyslipidemia, vitiligo, and hypertension. Polymorphisms -262 C/T and -844 A/G influence the gene expression and its activity. Physical activity, age, seasonal variations, and several chemical compounds further influence the expression of CAT [153]. In 2018, the effect of polymorphism in CAT, SOD, and GPX enzymes in clinical, anthropometric, and biochemical outputs was studied in obese patients under dietary intervention in a clinical trial. There was a significant difference in outcomes between genotypes. Genotypes rs7943316 SNPs-21A>T CAT and rs4880 47C>T SOD2 significantly affected clinical, anthropometric, and biochemical outcomes compared to other groups [154]. Apart from CAT, SOD, GPX enzymes, it was identified that catechin also exhibits its action by inhibiting MMP-2; this paved the pathway for analyzing gene Polymorphism associated with MMP-2. Around 2017, Andrea R et al., worked on MMP-2 polymorphism associated with drug resistant hypertension in the Brazilian population in a cross sectional study and concluded the result stating GCC and GCT haplotype of -735C/T MMP-2 may critically be associated with drug failure in hypertension, and this justifies the chances of genetic level change in MMP-2 can further affect catechin action [155].

EGCG induces vasorelaxation by increasing NO production by activating eNOS, which is responsible for the highest production of NO and has an extensive impact on the cardiovascular system. eNOS polymorphisms impact the response of drugs angiotensin II receptor antagonists, beta-blockers, statins, diuretics, angiotensin-converting enzyme inhibitors acting via NO signaling [156]. A clinical study by Silva et al., in 2013 presented anti-hypertensive elanapril response better in patients with rs2070744 polymorphism. But there were responders and nonresponders to the drug even though they the possessed same4b/4a and Glu298Asp polymorphism. Therefore, the 2996A/G Polymorphism in eNOS was responsible for enhancing beta-blockers and angiotensin II receptor blockers [157]. Diuretic hydrochlorthiazide action is modulated by the effect eNOS(Glu298Asp) polymorphism. Glu polymorphic individuals are significantly more responsive to hydrochlorothiazide than individuals with ASP polymorphism [158]. A study in 2003 demonstrated the effect of polymorphism in eNOS inhibitors, the Phosphodiesterase type 5 PDE-5 which are used in the treatment of erectile dysfunction since the drug acts by increasing cGMP levels in tissues in the absence of NO. Homozygous subjects of Glu298Asp Polymorphism are less responsive to sildenafil [159]. Calcium channel blockers such as nifedipine can enhance the NO bioavailability and endothelial function [160, 161]. A randomized clinical trial conducted in 2012 reported the polymorphic effects of NO<sub>3</sub> with stroke, and the glu298asp G>T variant resulted in mortality while comparing amlodipine with lisinopril [162]. Further, EC maintains the angiotensin- II and thereby balances the vascular tone, but in 1999, researchers conducted a clinical experiment with losartan, revealing a change in drug action associated with Ang II type 1 receptor genetic polymorphism. The study was conducted on 66 healthy Caucasian populations, para amino Hippurate and inulin clearance were used as a tool to measure renal hemodynamic function [87]. This analysis specified that EC following angiotensin mechanism may face gene-based variation in drug action similar to losartan.

The drug examples specified in the above sections were acting via one of the paths followed by GTCS to produce its cardiometabolic action. Hence there exists a possibility either the same polymorphic effects will affect the response of green tea. These possibilities will necessitate the personalization of green tea use to achieve maximum benefits. Further on, understanding green tea's challenges and opportunities from industrial perspectives will broaden the scope for real-time implementation for the benefit of the society.

#### **Real-Time implementation: Challenges and opportunities**

Globally, tea is one of the most highly consumed beverages. Multiple tea-producing countries and processing industries economically support this vast market size. Based on a 2014 survey, the global cultivation area of tea was around 3.8 million hectares. The productivity was about 5.6 million metric tons, including multiple verities and species of tea worth billions of dollars[163]. This growing market supported certain tea varieties to transit from commercial drink to health drink, and at present organic tea, white tea, etc., are typically considered a nutraceutical health drink. Based on these statistics, we can forecast the growth of the green tea market as a nutraceutical benefiting both producers and consumers. Hence, integrating the phytopharmacological knowledge with pharmacogenomics is regarded as an important way forward as it could advance the personalization of green tea for enhanced cardiometabolic outcomes. However, phyto-genomic approach towards green tea personalization also faces numerous challenges. The non-specific cardiometabolic pathway of GTCS makes it challenging to predict and narrow down the influencing gene. Gene-related experiments are expensive, which in turn may increase the product cost, stringent regulations on phytochemical-based clinical trials, lack of extensive clinical data, etc. Transdisciplinary approaches with the support of phytochemical industries, clinical researchers, and the technology sector can resolve these challenges, resulting in next-generation personalized green tea for a specified population.

#### Conclusion

Green tea and its polyphenols such as catechins and EGCG especially have gained attention in targeting CVDs. The mechanism by which green tea exerts cardio-metabolic effects includes anti-oxidant, anti-inflammatory, vascular endothelial dysfunction, anti-proliferative, antiplatelet and anti-thrombotic activities. Apart from its application towards CVDs, green tea also possesses various other applications in diabetes, cancer, and other minor benefits such as headaches and better digestion etc., towards human health. However, in this review, we have extensively collated published literature on green tea with its role in preventing CVDs; most clinical studies suggest a complete reduction in CVDs. On the other hand, some interventional studies possess the hepatotoxic effect of green tea consumption.

Further, there are also mixed data about the effects of the consumption of green tea on cardiac risk factors from observational and intervention studies. We firmly believe that green tea exhibits significant inter-individual variability on cardio-metabolic health based on many heterogeneously reported clinical interventions. This further paved the research to focus on genetic polymorphism associated with inter-individual variability. Polymorphisms of *NOS, COX 1, COX 2, SOD, CAT*, etc. have been modifying the conventional drug's efficacy via cardiometabolic pathways. This evidence indicates the probability of polymorphic effects in individuals using green tea for cardio-metabolic effects and the necessity to individualize green tea therapy.

# **Conflict of Interest**

The authors declare no conflict of interest.

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I certify that no financial and/or material support was received for this research and/or the creation of this work

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# **References:**

- 1. Nathan DM. Diabetes: advances in diagnosis and treatment. Jama. 2015;314(10):1052-62.
- 2. Organization WH. Health topics: Cardiovascular diseases. See http://www.who int/topics/cardiovascular\_diseases/en/(last checked July 2012). 2013.
- 3. Organization WH. World health statistics 2009: World Health Organization; 2009.
- 4. Ausloos M, Brugha TS, Collaborators G. Global, regional, and national disability-adjusted lifeyears (DALYs) for 359 diseases and injuries and healthy life expectancy (HALE) for 195 countries and territories, 1990-2017: a systematic analysis for the Global Burden of Disease Study 2017. 2018.
- 5. Kyu HH, Abate D, Abate KH, Abay SM, Abbafati C, Abbasi N, et al. Global, regional, and national disability-adjusted life-years (DALYs) for 359 diseases and injuries and healthy life

expectancy (HALE) for 195 countries and territories, 1990–2017: a systematic analysis for the Global Burden of Disease Study 2017. The Lancet. 2018;392(10159):1859-922.

- 6. Booker A, Zhai L, Gkouva C, Li S, Heinrich M. From traditional resource to global commodities:—a comparison of Rhodiola species using NMR spectroscopy—metabolomics and HPTLC. Frontiers in pharmacology. 2016;7:254.
- 7. Meegahakumbura MK, Wambulwa MC, Li M-M, Thapa KK, Sun Y-S, Möller M, et al. Domestication origin and breeding history of the tea plant (Camellia sinensis) in China and India based on nuclear microsatellites and cpDNA sequence data. Frontiers in plant science. 2018;8:2270.
- 8. Poirier P, Giles TD, Bray GA, Hong Y, Stern JS, Pi-Sunyer FX, et al. Obesity and cardiovascular disease: pathophysiology, evaluation, and effect of weight loss: an update of the 1997 American Heart Association Scientific Statement on Obesity and Heart Disease from the Obesity Committee of the Council on Nutrition, Physical Activity, and Metabolism. Circulation. 2006;113(6):898-918.
- 9. Cabrera C, Artacho R, Giménez R. Beneficial effects of green tea—a review. Journal of the American College of Nutrition. 2006;25(2):79-99.
- 10. Venkatakrishnan K, Chiu H-F, Cheng J-C, Chang Y-H, Lu Y-Y, Han Y-C, et al. Comparative studies on the hypolipidemic, anti-oxidant and hepatoprotective activities of catechin-enriched green and oolong tea in a double-blind clinical trial. Food & function. 2018;9(2):1205-13.
- 11. Goszcz K, Duthie GG, Stewart D, Leslie SJ, Megson IL. Bioactive polyphenols and cardiovascular disease: chemical antagonists, pharmacological agents or xenobiotics that drive an adaptive response? British journal of pharmacology. 2017;174(11):1209-25.
- 12. Arbeláez LFG, Pardo AC, Fantinelli JC, Schinella GR, Mosca SM, Ríos J-L. Cardioprotection and natural polyphenols: an update of clinical and experimental studies. Food & function. 2018;9(12):6129-45.
- 13. Ibrahim MA, Bakhaat GA, Tammam HG, Mohamed RM, El-Naggar SA. Cardioprotective effect of green tea extract and vitamin E on Cisplatin-induced cardiotoxicity in mice: toxicological, histological and immunohistochemical studies. Biomedicine & Pharmacotherapy. 2019;113:108731.
- 14. Albuquerque BR, Heleno SA, Oliveira MBP, Barros L, Ferreira IC. Phenolic compounds: current industrial applications, limitations and future challenges. Food & Function. 2021;12(1):14-29.
- 15. Tsoupras A, Lordan R, Harrington J, Pienaar R, Devaney K, Heaney S, et al. The effects of oxidation on the antithrombotic properties of tea lipids against PAF, thrombin, collagen, and ADP. Foods. 2020;9(4):385.
- 16. Li G, Zhang Y, Thabane L, Mbuagbaw L, Liu A, Levine MA, et al. Effect of green tea supplementation on blood pressure among overweight and obese adults: a systematic review and meta-analysis. Journal of hypertension. 2015;33(2):243-54.
- 17. Katanasaka Y, Miyazaki Y, Sunagawa Y, Funamoto M, Shimizu K, Shimizu S, et al. Kosencha, a polymerized catechin-rich green tea, as a potential functional beverage for the reduction

of body weight and cardiovascular risk factors: a pilot study in obese patients. Biological and Pharmaceutical Bulletin. 2020;43(4):675-81.

- 18. Landini L, Rebelos E, Honka M-J. Green Tea from the Far East to the Drug Store: Focus on the Beneficial Cardiovascular Effects. Current Pharmaceutical Design. 2021.
- 19. Ikeda A, Iso H, Yamagishi K, Iwasaki M, Yamaji T, Miura T, et al. Plasma tea catechins and risk of cardiovascular disease in middle-aged Japanese subjects: The JPHC study. Atherosclerosis. 2018;277:90-7.
- 20. Xing L, Zhang H, Qi R, Tsao R, Mine Y. Recent advances in the understanding of the health benefits and molecular mechanisms associated with green tea polyphenols. Journal of agricultural and food chemistry. 2019;67(4):1029-43.
- Yang CS, Wang H, Sheridan ZP. Studies on prevention of obesity, metabolic syndrome, diabetes, cardiovascular diseases and cancer by tea. journal of food and drug analysis. 2018;26(1):1-13.
- 22. Arts IC, Hollman PC, Feskens EJ, Bueno de Mesquita HB, Kromhout D. Catechin intake might explain the inverse relation between tea consumption and ischemic heart disease: the Zutphen Elderly Study. The American journal of clinical nutrition. 2001;74(2):227-32.
- 23. Jówko E. Green tea catechins and sport performance. SPORT NUTRITION. 2015:123.
- 24. Williams SN, Shih H, Guenette DK, Brackney W, Denison MS, Pickwell GV, et al. Comparative studies on the effects of green tea extracts and individual tea catechins on human CYP1A gene expression. Chemico-biological interactions. 2000;128(3):211-29.
- 25. Iwasaki M, Inoue M, Sasazuki S, Sawada N, Yamaji T, Shimazu T, et al. Green tea drinking and subsequent risk of breast cancer in a population to based cohort of Japanese women. Breast Cancer Research. 2010;12(5):1-10.
- 26. Liu J, Liu S, Zhou H, Hanson T, Yang L, Chen Z, et al. Association of green tea consumption with mortality from all-cause, cardiovascular disease and cancer in a Chinese cohort of 165,000 adult men. European journal of epidemiology. 2016;31(9):853-65.
- 27. Olafuyi O, Parekh N, Wright J, Koenig J. Inter-ethnic differences in pharmacokinetics—is there more that unites than divides? Pharmacology research & perspectives. 2021;9(6):e00890.
- 28. Mazzanti G, Di Sotto A, Vitalone A. Hepatotoxicity of green tea: an update. Archives of toxicology. 2015;89(8):1175-91.
- 29. Additives EPoF, Food NSat, Younes M, Aggett P, Aguilar F, Crebelli R, et al. Scientific opinion on the safety of green tea catechins. EFSA Journal. 2018;16(4):e05239.
- 30. Ghodke Y, Joshi K, Patwardhan B. Traditional medicine to modern pharmacogenomics: Ayurveda Prakriti type and CYP2C19 gene polymorphism associated with the metabolic variability. Evidence-Based Complementary and Alternative Medicine. 2011;2011.
- 31. Stangl V, Dreger H, Stangl K, Lorenz M. Molecular targets of tea polyphenols in the cardiovascular system. Cardiovascular research. 2007;73(2):348-58.
- 32. Suzuki J-i, Isobe M, Morishita R, Nagai R. Tea polyphenols regulate key mediators on inflammatory cardiovascular diseases. Mediators of inflammation. 2009;2009.

- 33. Tomalik-Scharte D, Lazar A, Fuhr U, Kirchheiner J. The clinical role of genetic polymorphisms in drug-metabolizing enzymes. The pharmacogenomics journal. 2008;8(1):4-15.
- 34. Brown S-A, Pereira N. Pharmacogenomic impact of CYP2C19 variation on clopidogrel therapy in precision cardiovascular medicine. Journal of personalized medicine. 2018;8(1):8.
- 35. Tokunaga S, White IR, Frost C, Tanaka K, Kono S, Tokudome S, et al. Green tea consumption and serum lipids and lipoproteins in a population of healthy workers in Japan. Annals of epidemiology. 2002;12(3):157-65.
- 36. Hollman PC, Feskens EJ, Katan MB. Tea flavonols in cardiovascular disease and cancer epidemiology. Proceedings of the Society for experimental Biology and Medicine. 1999;220(4):198-202.
- 37. Kuriyama S. The relation between green tea consumption and cardiovascular disease as evidenced by epidemiological studies. The Journal of nutrition. 2008;138(8):1548S-53S.
- 38. Kuriyama S. Green tea consumption and prevention of coronary artery disease. Circulation Journal. 2010;74(2):248-9.
- 39. Sasazuki S, Kodama H, Yoshimasu K, Liu Y, Washio M, Tanaka K, et al. Relation between green tea consumption and the severity of coronary atherosclerosis among Japanese men and women. Annals of Epidemiology. 2000;10(6):401-8.
- 40. Suzuki E, Yorifuji T, Takao S, Komatsu H, Sugiyama M, Ohta T, et al. Green tea consumption and mortality among Japanese elderly people: the prospective Shizuoka elderly cohort. Annals of epidemiology. 2009;19(10):732-9.
- 41. Nakachi K, Matsuyama S, Miyake S, Suganuma M, Imai K. Preventive effects of drinking green tea on cancer and cardiovascular disease: epidemiological evidence for multiple targeting prevention. Biofactors. 2000;13(1-4):49-54.
- 42. Zhao L-G, Li H-L, Sun J-W, Yang Y, Ma X, Shu X-O, et al. Green tea consumption and causespecific mortality: Results from two prospective cohort studies in China. Journal of epidemiology. 2017;27(1):36-41.
- 43. Wang Q-M, Gong Q-Y, Yan J-J, Tang J-J, Wang M-W, Yang Z-J, et al. Association between green tea intake and coronary artery disease in a Chinese population. Circulation Journal. 2010:0912140554-.
- 44. Miller RJ, Jackson KG, Dadd T, Mayes AE, Brown AL, Minihane AM. The impact of the catechol-O-methyltransferase genotype on the acute responsiveness of vascular reactivity to a green tea extract. British journal of nutrition. 2011;105(8):1138-44.
- 45. Frank J, George TW, Lodge JK, Rodriguez-Mateos AM, Spencer JP, Minihane AM, et al. Daily consumption of an aqueous green tea extract supplement does not impair liver function or alter cardiovascular disease risk biomarkers in healthy men. The Journal of nutrition. 2009;139(1):58-62.
- 46. Grassi D, Aggio A, Onori L, Croce G, Tiberti S, Ferri C, et al. Tea, flavonoids, and nitric oxide-mediated vascular reactivity. The Journal of nutrition. 2008;138(8):1554S-60S.

- 47. Stensvold I, Tverdal A, Solvoll K, Foss OP. Tea consumption. Relationship to cholesterol, blood pressure, and coronary and total mortality. Preventive medicine. 1992;21(4):546-53.
- 48. Duffy SJ, Keaney Jr JF, Holbrook M, Gokce N, Swerdloff PL, Frei B, et al. Short-and longterm black tea consumption reverses endothelial dysfunction in patients with coronary artery disease. Circulation. 2001;104(2):151-6.
- 49. Nantz MP, Rowe CA, Bukowski JF, Percival SS. Standardized capsule of Camellia sinensis lowers cardiovascular risk factors in a randomized, double-blind, placebo-controlled study. Nutrition. 2009;25(2):147-54.
- 50. Hodgson JM, Devine A, Puddey IB, Chan SY, Beilin LJ, Prince RL. Tea intake is inversely related to blood pressure in older women. The Journal of nutrition. 2003;133(9):2883-6.
- 51. Yang Y-C, Lu F-H, Wu J-S, Wu C-H, Chang C-J. The protective effect of habitual tea consumption on hypertension. Archives of internal medicine. 2004;164(14):1534-40.
- 52. Erba D, Riso P, Bordoni A, Foti P, Biagi PL, Testolin G. Effectiveness of moderate green tea consumption on antioxidative status and plasma lipid profile in humans. The Journal of nutritional biochemistry. 2005;16(3):144-9.
- 53. Unno T, Tago M, Suzuki Y, Nozawa A, Sagesaka YM, Kakuda T, et al. Effect of tea catechins on postprandial plasma lipid responses in human subjects. British Journal of Nutrition. 2005;93(4):543-7.
- 54. Fukino Y, Shimbo M, Aoki N, Okubo T, Iso H. Randomized controlled trial for an effect of green tea consumption on insulin resistance and inflammation markers. Journal of nutritional science and vitaminology. 2005;51(5):335-42.
- 55. Kim W, Jeong MH, Cho SH, Yun JH, Chae HJ, Ahn YK, et al. Effect of green tea consumption on endothelial function and circulating endothelial progenitor cells in chronic smokers. Circulation Journal. 2006;70(8):1052-7.
- 56. Hodgson JM, Puddey IB, Burke V, Croft KD. Is reversal of endothelial dysfunction by tea related to flavonoid metabolism? British journal of nutrition. 2006;95(1):14-7.
- 57. Hodgson J. Effects of tea and tea flavonoids on endothelial function and blood pressure: a brief review. Clinical and experimental pharmacology and physiology. 2006;33(9):838-41.
- 58. Inami S, Takano M, Yamamoto M, Murakami D, Tajika K, Yodogawa K, et al. Tea catechin consumption reduces circulating oxidized low-density lipoprotein. International Heart Journal. 2007;48(6):725-32.
- 59. Nagao T, Hase T, Tokimitsu I. A green tea extract high in catechins reduces body fat and cardiovascular risks in humans. Obesity. 2007;15(6):1473-83.
- 60. Gomikawa S, Ishikawa Y, Hayase W, Haratake Y, Hirano N, Matuura H, et al. Effect of ground green tea drinking for 2 weeks on the susceptibility of plasma and LDL to the oxidation ex vivo in healthy volunteers. Kobe J Med Sci. 2008;54(1):62-72.
- 61. Loke WM, Hodgson JM, Proudfoot JM, McKinley AJ, Puddey IB, Croft KD. Pure dietary flavonoids quercetin and (–)-epicatechin augment nitric oxide products and reduce endothelin-1 acutely in healthy men. The American journal of clinical nutrition. 2008;88(4):1018-25.

- 62. Alexopoulos N, Vlachopoulos C, Aznaouridis K, Baou K, Vasiliadou C, Pietri P, et al. The acute effect of green tea consumption on endothelial function in healthy individuals. European Journal of Preventive Cardiology. 2008;15(3):300-5.
- 63. Tinahones F, Rubio M, Garrido-Sanchez L, Ruiz C, Gordillo E, Cabrerizo L, et al. Green tea reduces LDL oxidability and improves vascular function. Journal of the American College of Nutrition. 2008;27(2):209-13.
- 64. Nagao T, Meguro S, Hase T, Otsuka K, Komikado M, Tokimitsu I, et al. A catechin-rich beverage improves obesity and blood glucose control in patients with type 2 diabetes. Obesity. 2009;17(2):310-7.
- 65. Basu A, Sanchez K, Leyva MJ, Wu M, Betts NM, Aston CE, et al. Green tea supplementation affects body weight, lipids, and lipid peroxidation in obese subjects with metabolic syndrome. Journal of the American College of Nutrition. 2010;29(1):31-40.
- 66. Bhardwaj P, Khanna D. Green tea catechins: defensive role in cardiovascular disorders. Chinese journal of natural medicines. 2013;11(4):345-53.
- 67. Kim A, Chiu A, Barone MK, Avino D, Wang F, Coleman CI, et al. Green tea catechins decrease total and low-density lipoprotein cholesterol: a systematic review and meta-analysis. Journal of the American Dietetic Association. 2011;111(11):1720-9.
- 68. Wu AH, Spicer D, Stanczyk FZ, Tseng C-C, Yang CS, Pike MC. Effect of 2-month controlled green tea intervention on lipoprotein cholesterol, glucose, and hormone levels in healthy postmenopausal women. Cancer Prevention Research. 2012;5(3):393-402.
- 69. Bogdanski P, Suliburska J, Szulinska M, Stepien M, Pupek-Musialik D, Jablecka A. Green tea extract reduces blood pressure, inflammatory biomarkers, and oxidative stress and improves parameters associated with insulin resistance in obese, hypertensive patients. Nutrition research. 2012;32(6):421-7.
- Hartley L, Flowers N, Holmes J, Clarke A, Stranges S, Hooper L, et al. Green and black tea for the primary prevention of cardiovascular disease. Cochrane Database of Systematic Reviews. 2013(6).
- Liu C-Y, Huang C-J, Huang L-H, Chen I-J, Chiu J-P, Hsu C-H. Effects of green tea extract on insulin resistance and glucagon-like peptide 1 in patients with type 2 diabetes and lipid abnormalities: a randomized, double-blinded, and placebo-controlled trial. PLoS One. 2014;9(3):e91163.
- 72. Chen I-J, Liu C-Y, Chiu J-P, Hsu C-H. Therapeutic effect of high-dose green tea extract on weight reduction: A randomized, double-blind, placebo-controlled clinical trial. Clinical Nutrition. 2016;35(3):592-9.
- 73. Yang CS, Zhang J. Studies on the prevention of cancer and cardiometabolic diseases by tea: Issues on mechanisms, effective doses, and toxicities. Journal of agricultural and food chemistry. 2018;67(19):5446-56.
- Etheridge C, Bond T, Derbyshire E. Effects of tea consumption on measures of cardiovascular disease: A systematic review of meta-analysis studies and randomised controlled trials. J Nutr Food Sci. 2018;8(724):2.

- 75. Chung M, Zhao N, Wang D, Shams-White M, Karlsen M, Cassidy A, et al. Dose–response relation between tea consumption and risk of cardiovascular disease and all-cause mortality: a systematic review and meta-analysis of population-based studies. Advances in Nutrition. 2020;11(4):790-814.
- 76. Xu R, Yang K, Ding J, Chen G. Effect of green tea supplementation on blood pressure: A systematic review and meta-analysis of randomized controlled trials. Medicine. 2020;99(6).
- 77. Ryu O, Lee J, Lee K, Kim H, Seo JA, Kim SG, et al. Effects of green tea consumption on inflammation, insulin resistance and pulse wave velocity in type 2 diabetes patients. Diabetes research and clinical practice. 2006;71(3):356-8.
- 78. Hirano-Ohmori R, Takahashi R, Momiyama Y, Taniguchi H, Yonemura A, Tamai S, et al. Green tea consumption and serum malondialdehyde-modified LDL concentrations in healthy subjects. Journal of the American College of Nutrition. 2005;24(5):342-6.
- 79. Diepvens K, Kovacs E, Vogels N, Westerterp-Plantenga M. Metabolic effects of green tea and of phases of weight loss. Physiology & behavior. 2006;87(1):185-91.
- 80. Matsuyama T, Tanaka Y, Kamimaki I, Nagao T, Tokimitsu I. Catechin safely improved higher levels of fatness, blood pressure, and cholesterol in children. Obesity. 2008;16(6):1338-48.
- Hsu C-H, Tsai T-H, Kao Y-H, Hwang K-C, Tseng T-Y, Chou P. Effect of green tea extract on obese women: a randomized, double-blind, placebo-controlled clinical trial. Clinical nutrition. 2008;27(3):363-70.
- 82. Brown AL, Lane J, Coverly J, Stocks J, Jackson S, Stephen A, et al. Effects of dietary supplementation with the green tea polyphenol epigallocatechin-3-gallate on insulin resistance and associated metabolic risk factors: randomized controlled trial. British journal of nutrition. 2008;101(6):886-94.
- 83. Trautwein EA, Du Y, Meynen E, Yan X, Wen Y, Wang H, et al. Purified black tea theaflavins and theaflavins/catechin supplements did not affect serum lipids in healthy individuals with mildly to moderately elevated cholesterol concentrations. European journal of nutrition. 2010;49(1):27-35.
- Deka A, Vita JA. Tea and cardiovascular disease. Pharmacological research. 2011;64(2):136-45.
- 85. Hua C, Liao Y, Lin S, Tsai T, Huang C, Chou P. Does supplementation with green tea extract improve insulin resistance in obese type 2 diabetics? A randomized, double-blind, and placebocontrolled clinical trial. Alternative Medicine Review. 2011;16(2):157-63.
- 86. Zheng X-X, Xu Y-L, Li S-H, Liu X-X, Hui R, Huang X-H. Green tea intake lowers fasting serum total and LDL cholesterol in adults: a meta-analysis of 14 randomized controlled trials. The American journal of clinical nutrition. 2011;94(2):601-10.
- 87. Zhang X, Lynch AI, Davis BR, Ford CE, Boerwinkle E, Eckfeldt JH, et al. Pharmacogenetic association of NOS3 variants with cardiovascular disease in patients with hypertension: the GenHAT study. PLoS One. 2012;7(3):e34217.

- Onakpoya I, Spencer E, Heneghan C, Thompson M. The effect of green tea on blood pressure and lipid profile: a systematic review and meta-analysis of randomized clinical trials. Nutrition, Metabolism and Cardiovascular Diseases. 2014;24(8):823-36.
- 89. Jurgens T, Whelan AM. Can green tea preparations help with weight loss? Canadian Pharmacists Journal. 2014;147(3):159.
- 90. Xu R, Yang K, Li S, Dai M, Chen G. Effect of green tea consumption on blood lipids: a systematic review and meta-analysis of randomized controlled trials. Nutrition journal. 2020;19:1-15.
- 91. Zhou S-F, Ming Di Y, Chan E, Du Y-M, Chow VD-W, Xue CC, et al. Clinical pharmacogenetics and potential application in personalized medicine. Current drug metabolism. 2008;9(8):738-84.
- 92. Goldstein DB, Tate SK, Sisodiya SM. Pharmacogenetics goes genomic. Nature Reviews Genetics. 2003;4(12):937-47.
- 93. Evans WE, Relling MV. Pharmacogenomics: translating functional genomics into rational therapeutics. science. 1999;286(5439):487-91.
- 94. Dhalla NS, Temsah RM, Netticadan T. Role of oxidative stress in cardiovascular diseases. Journal of hypertension. 2000;18(6):655-73.
- 95. Paravicini TM, Touyz RM. NADPH oxidases, reactive oxygen species, and hypertension: clinical implications and therapeutic possibilities. Diabetes care. 2008;31(Supplement 2):S170-S80.
- 96. Park J, Lee J, Choi C. Mitochondrial network determines intracellular ROS dynamics and sensitivity to oxidative stress through switching inter-mitochondrial messengers. PloS one. 2011;6(8):e23211.
- 97. Zinkevich NS, Gutterman DD. ROS-induced ROS release in vascular biology: redox-redox signaling. American journal of physiology-heart and circulatory physiology. 2011;301(3):H647-H53.
- 98. A Islam M. Cardiovascular effects of green tea catechins: progress and promise. Recent Patents on Cardiovascular Drug Discovery (Discontinued). 2012;7(2):88-99.
- 99. Marui N, Offermann MK, Swerlick R, Kunsch C, Rosen CA, Ahmad M, et al. Vascular cell adhesion molecule-1 (VCAM-1) gene transcription and expression are regulated through an antioxidant-sensitive mechanism in human vascular endothelial cells. The Journal of clinical investigation. 1993;92(4):1866-74.
- Lin Y-L, Lin J-K. (–)-Epigallocatechin-3-gallate blocks the induction of nitric oxide synthase by down-regulating lipopolysaccharide-induced activity of transcription factor nuclear factorκB. Molecular pharmacology. 1997;52(3):465-72.
- 101. Gu L, Okada Y, Clinton SK, Gerard C, Sukhova GK, Libby P, et al. Absence of monocyte chemoattractant protein-1 reduces atherosclerosis in low density lipoprotein receptor–deficient mice. Molecular cell. 1998;2(2):275-81.

- 102. Gerszten RE, Garcia-Zepeda EA, Lim Y-C, Yoshida M, Ding HA, Gimbrone MA, et al. MCP-1 and IL-8 trigger firm adhesion of monocytes to vascular endothelium under flow conditions. Nature. 1999;398(6729):718-23.
- Kevil CG, Patel RP, Bullard DC. Essential role of ICAM-1 in mediating monocyte adhesion to aortic endothelial cells. American Journal of Physiology-Cell Physiology. 2001;281(5):C1442-C7.
- 104. Ludwig A, Lorenz M, Grimbo N, Steinle F, Meiners S, Bartsch C, et al. The tea flavonoid epigallocatechin-3-gallate reduces cytokine-induced VCAM-1 expression and monocyte adhesion to endothelial cells. Biochemical and biophysical research communications. 2004;316(3):659-65.
- 105. Libby P. Inflammation and cardiovascular disease mechanisms. The American journal of clinical nutrition. 2006;83(2):456S-60S.
- 106. Suzuki Ji, Ogawa M, Futamatsu H, Kosuge H, Sagesaka YM, Isobe M. Tea catechins improve left ventricular dysfunction, suppress myocardial inflammation and fibrosis, and alter cytokine expression in rat autoimmune myocarditis. European journal of heart failure. 2007;9(2):152-9.
- 107. Dimmeler S, Fleming I, Fisslthaler B, Hermann C, Busse R, Zeiher AM. Activation of nitric oxide synthase in endothelial cells by Akt-dependent phosphorylation. Nature. 1999;399(6736):601-5.
- 108. Schiffrin EL, Touyz RM. From bedside to bench to bedside: role of renin-angiotensinaldosterone system in remodeling of resistance arteries in hypertension. American Journal of Physiology-Heart and Circulatory Physiology. 2004;287(2):H435-H46.
- 109. Galley HF, Webster NR. Physiology of the endothelium. British journal of anaesthesia. 2004;93(1):105-13.
- Kobayashi T, Matsumoto T, Kamata K. The PI3-K/Akt pathway: roles related to alterations in vasomotor responses in diabetic models. Journal of Smooth Muscle Research. 2005;41(6):283-302.
- 111. Hadi HA, Carr CS, Al Suwaidi J. Endothelial dysfunction: cardiovascular risk factors, therapy, and outcome. Vascular health and risk management. 2005;1(3):183.
- 112. Forstermann U, Munzel T. Endothelial nitric oxide synthase in vascular disease: from marvel to menace. Circulation. 2006;113(13):1708-14.
- 113. Basu A, Lucas EA. Mechanisms and effects of green tea on cardiovascular health. Nutrition reviews. 2007;65(8):361-75.
- 114. Balakumar P, Chakkarwar VA, Krishan P, Singh M. Vascular endothelial dysfunction: a tug of war in diabetic nephropathy? Biomedicine & Pharmacotherapy. 2009;63(3):171-9.
- 115. Ross R, Glomset JA. Atherosclerosis and the arterial smooth muscle cell. Science. 1973;180(4093):1332-9.
- 116. Lindner V, Reidy M. Proliferation of smooth muscle cells after vascular injury is inhibited by an antibody against basic fibroblast growth factor. Proceedings of the National Academy of Sciences. 1991;88(9):3739-43.

- 117. Schwartz S. deBlois D., O'Brien ER The intima. Soil for atherosclerosis and restenosis. Circ Res. 1995;77(3):445-65.
- 118. Yamamoto M, Acevedo-Duncan M, Chalfant CE, Patel NA, Watson JE, Cooper DR. Acute glucose-induced downregulation of PKC-βII accelerates cultured VSMC proliferation. American Journal of Physiology-Cell Physiology. 2000;279(3):C587-C95.
- 119. Visse R, Nagase H. Matrix metalloproteinases and tissue inhibitors of metalloproteinases: structure, function, and biochemistry. Circulation research. 2003;92(8):827-39.
- 120. Ouyang P, Peng W, Xu D, Lai W, Xu A. Green tea polyphenols inhibit advanced glycation end product-induced rat vascular smooth muscle cell proliferation. Di 1 jun yi da xue xue bao= Academic Journal of the First Medical College of PLA. 2004;24(3):247-51.
- 121. De Donatis A, Comito G, Buricchi F, Vinci MC, Parenti A, Caselli A, et al. Proliferation versus migration in platelet-derived growth factor signaling: the key role of endocytosis. Journal of Biological Chemistry. 2008;283(29):19948-56.
- 122. Yuan X, Zhang Z, Gong K, Zhao P, Qin J, Liu N. Inhibition of reactive oxygen species/extracellular signal-regulated kinases pathway by pioglitazone attenuates advanced glycation end products-induced proliferation of vascular smooth muscle cells in rats. Biological and Pharmaceutical Bulletin. 2011;34(5):618-23.
- 123. Yang J, Han Y, Sun H, Chen C, He D, Guo J, et al. (–)-Epigallocatechin gallate suppresses proliferation of vascular smooth muscle cells induced by high glucose by inhibition of PKC and ERK1/2 signalings. Journal of agricultural and food chemistry. 2011;59(21):11483-90.
- 124. Born G. Symposium on Thrombosis: Platelets in thrombogenesis: mechanism and inhibition of platelet aggregation. Annals of the Royal College of Surgeons of England. 1965;36(4):200.
- 125. Kang W-S, Chung K-H, Chung J-H, Lee J-Y, Park J-B, Zhang Y-H, et al. Antiplatelet activity of green tea catechins is mediated by inhibition of cytoplasmic calcium increase. Journal of cardiovascular pharmacology. 2001;38(6):875-84.
- 126. Di Michela M, Van Geet C, Freson K. Proteomics to unravel platelet-related diseases and identify novel anti-platelet drugs. Current medicinal chemistry. 2012;19(27):4662-70.
- 127. Luo K, Ma C, Xing S, An Y, Feng J, Dang H, et al. White tea and its active polyphenols lower cholesterol through reduction of very-low-density lipoprotein production and induction of LDLR expression. Biomedicine & Pharmacotherapy. 2020;127:110146.
- 128. Xu R, Yang K, Ding J, Chen G. Effect of green tea supplementation on blood pressure: A systematic review and meta-analysis of randomized controlled trials. Medicine. 2020 Feb;99(6).
- 129. Buguliskis JS. Pharmacogenomics serves as the critical driver for precision medicine. Clinical OMICs. 2015;2(6):12-4, 6.
- 130. Relling MV, Evans WE. Pharmacogenomics in the clinic. Nature. 2015;526(7573):343-50.
- 131. Mega JL, Close SL, Wiviott SD, Shen L, Walker JR, Simon T, et al. Genetic variants in ABCB1 and CYP2C19 and cardiovascular outcomes after treatment with clopidogrel and prasugrel in the TRITON–TIMI 38 trial: a pharmacogenetic analysis. The Lancet. 2010;376(9749):1312-9.

- 132. Hodges LM, Markova SM, Chinn LW, Gow JM, Kroetz DL, Klein TE, et al. Very important pharmacogene summary: ABCB1 (MDR1, P-glycoprotein). Pharmacogenetics and genomics. 2011;21(3):152.
- 133. Fukunaga K, Nakagawa H, Ishikawa T, Kubo M, Mushiroda T. ABCB1 polymorphism is associated with atorvastatin-induced liver injury in Japanese population. BMC genetics. 2016;17(1):1-6.
- 134. Su J, Xu H, Yang J, Yu Q, Yang S, Zhang J, et al. ABCB1 C3435T polymorphism and the lipid-lowering response in hypercholesterolemic patients on statins: a meta-analysis. Lipids in health and disease. 2015;14(1):1-10.
- 135. Roden DM, Johnson JA, Kimmel SE, Krauss RM, Medina MW, Shuldiner A, et al. Cardiovascular pharmacogenomics. Circulation research. 2011;109(7):807-20.
- 136. Bruhn O, Cascorbi I. Polymorphisms of the drug transporters ABCB1, ABCG2, ABCC2 and ABCC3 and their impact on drug bioavailability and clinical relevance. Expert opinion on drug metabolism & toxicology. 2014;10(10):1337-54.
- 137. König Jr, Cui Y, Nies AT, Keppler D. A novel human organic anion transporting polypeptide localized to the basolateral hepatocyte membrane. American Journal of Physiology-Gastrointestinal and Liver Physiology. 2000;278(1):G156-G64.
- 138. Gong IY, Kim RB. Impact of genetic variation in OATP transporters to drug disposition and response. Drug metabolism and pharmacokinetics. 2012:DMPK-12-RV-099.
- 139. Lorenz M, Paul F, Moobed M, Baumann G, Zimmermann BF, Stangl K, et al. The activity of catechol-O-methyltransferase (COMT) is not impaired by high doses of epigallocatechin-3-gallate (EGCG) in vivo. European journal of pharmacology. 2014;740:645-51.
- 140. Brown A, Lane J, Holyoak C, Nicol B, Mayes AE, Dadd T. Health effects of green tea catechins in overweight and obese men: a randomised controlled cross-over trial. British journal of nutrition. 2011;106(12):1880-9.
- 141. Inoue-Choi M, Yuan J-M, Yang CS, Van Den Berg DJ, Lee M-J, Gao Y-T, et al. Genetic association between the COMT genotype and urinary levels of tea polyphenols and their metabolites among daily green tea drinkers. International journal of molecular epidemiology and genetics. 2010;1(2):114.
- 142. Wu AH, Tseng C-C, Van Den Berg D, Mimi CY. Tea intake, COMT genotype, and breast cancer in Asian-American women. Cancer research. 2003;63(21):7526-9.
- 143. Miller RJ, Jackson KG, Dadd T, Nicol B, Dick JL, Mayes AE, et al. A preliminary investigation of the impact of catechol-O-methyltransferase genotype on the absorption and metabolism of green tea catechins. European journal of nutrition. 2012;51(1):47-55.
- Fleming M, Lin GD, Li RW. Omics Technologies and Development of Anti-diabetic Therapies from Prospective Natural Products. Evidence Based Validation of Traditional Medicines. 2021:77-96.
- 145. Cascorbi I. Role of pharmacogenetics of ATP-binding cassette transporters in the pharmacokinetics of drugs. Pharmacology & therapeutics. 2006;112(2):457-73.

- 146. Roth M, Timmermann BN, Hagenbuch B. Interactions of green tea catechins with organic anion-transporting polypeptides. Drug Metabolism and Disposition. 2011;39(5):920-6.
- 147. Kadowaki M, Sugihara N, Tagashira T, Terao K, Furuno K. Presence or absence of a gallate moiety on catechins affects their cellular transport. Journal of Pharmacy and Pharmacology. 2008;60(9):1189-95.
- 148. Scarel-Caminaga RM, Cera FF, Pigossi SC, Finoti LS, Kim YJ, Viana AC, et al. Inducible nitric oxide synthase polymorphisms and nitric oxide levels in individuals with chronic periodontitis. International journal of molecular sciences. 2017;18(6):1128.
- 149. Tantisira KG, Lima J, Sylvia J, Klanderman B, Weiss ST. 5-lipoxygenase pharmacogenetics in asthma: overlap with CystLTR1 loci. Pharmacogenetics and genomics. 2009;19(3):244.
- 150. Cambria-Kiely JA, Gandhi PJ. Aspirin resistance and genetic polymorphisms. Journal of thrombosis and thrombolysis. 2002;14(1):51-8.
- 151. Lee SJ, Park MK, Shin D-S, Chun MH. Variability of the drug response to nonsteroidal antiinflammatory drugs according to cyclooxygenase-2 genetic polymorphism. Drug design, development and therapy. 2017;11:2727.
- 152. Silig Y, Tas A, Sahin-Bolukbasi S, Caglayan G, Sari I. Superoxide Dismutase 1 (SOD 1) A251G Polymorphism. Turkish Journal of Biochemistry. 2017;42(2):181-5.
- 153. Kodydková J, Vávrová L, Kocík M, Zak A. Human catalase, its polymorphisms, regulation and changes of its activity in different diseases. Folia biologica. 2014;60(4):153.
- 154. Hernández-Guerrero C, Parra-Carriedo A, Ruiz-de-Santiago D, Galicia-Castillo O, Buenrostro-Jáuregui M, Díaz-Gutiérrez C. Genetic polymorphisms of anti-oxidant enzymes CAT and SOD affect the outcome of clinical, biochemical, and anthropometric variables in people with obesity under a dietary intervention. Genes & nutrition. 2018;13(1):1-10.
- 155. Sabbatini AR, Barbaro NR, de Faria AP, Ritter AMV, Modolo R, Correa NB, et al. Matrix metalloproteinase-2- 735C/T polymorphism is associated with resistant hypertension in a specialized outpatient clinic in Brazil. Gene. 2017;620:23-9.
- 156. Cozma A, Fodor A, Orasan OH, Vulturar R, Samplelean D, Negrean V, et al. Pharmacogenetic Implications of eNOS Polymorphisms (Glu298Asp, T786C, 4b/4a) in Cardiovascular Drug Therapy. in vivo. 2019;33(4):1051-8.
- 157. Silva P, Fontana V, Luizon M, Lacchini R, Silva W, Biagi C, et al. eNOS and BDKRB2 genotypes affect the anti-hypertensive responses to enalapril. European journal of clinical pharmacology. 2013;69(2):167-77.
- 158. Liljedahl U, Karlsson J, Melhus H, Kurland L, Lindersson M, Kahan T, et al. A microarray minisequencing system for pharmacogenetic profiling of anti-hypertensive drug response. Pharmacogenetics and Genomics. 2003;13(1):7-17.
- 159. Turner ST, Chapman AB, Schwartz GL, Boerwinkle E. Effects of endothelial nitric oxide synthase, α-adducin, and other candidate gene polymorphisms on blood pressure response to hydrochlorothiazide. American journal of hypertension. 2003;16(10):834-9.

- 160. Eisenhardt A, Sperling H, Hauck E, Porst H, Stief C, Rübben H, et al. ACE gene I/D and NOS3 G894T polymorphisms and response to sildenafil in men with erectile dysfunction. Urology. 2003;62(1):152-7.
- 161. Ding Y, Vaziri ND. Nifedipine and diltiazem but not verapamil up-regulate endothelial nitricoxide synthase expression. Journal of Pharmacology and Experimental Therapeutics. 2000;292(2):606-9.
- 162. Taddei S, Virdis A, Ghiadoni L, Magagna A, Favilla S, Pompella A, et al. Restoration of nitric oxide availability after calcium antagonist treatment in essential hypertension. Hypertension. 2001;37(3):943-8.
- 163. Miller JA, Thai K, Scholey JW. Angiotensin II type 1 receptor gene polymorphism predicts response to losartan and angiotensin II. Kidney international. 1999;56(6):2173-80.

#### **Graphical abstract**



#### Highlights

- Cardiovascular disease (CVD) is a chronic severe multifactorial disease that adversely affects an individual's health, well-being, and lifestyle.
- Polyphenols of green tea elicit a beneficial role in cardiometabolic health. The mechanism by which green tea exerts cardio-metabolic effects includes anti-oxidant, antiinflammatory, vascular endothelial dysfunction, anti-proliferative, antiplatelet and antithrombotic activities.
- Green tea exhibits significant inter-individual variability on cardio-metabolic health based on many heterogeneously reported clinical interventions.
- Polymorphisms of *NOS, COX 1, COX 2, SOD, CAT* etc. has been modifying the efficacy of the conventional drug acting via cardiometabolic pathways.
- Polymorphic effects in individuals using green tea for cardio-metabolic effects and other benefits and the necessity to individualize green tea therapy. Personalizing GTC therapy in cardio-metabolic health can regulate the risk-to-benefit ratio that could result in better market viability.