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1 Full title

2 Processes underlying glycemic deterioration in type 2 diabetes: An IMI DIRECT study

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4 **Running title**

- 5 Glycemic deterioration in type 2 diabetes
- 6

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96 Abstract

97 *Objective*

98 We investigated the processes underlying glycemic deterioration in type 2 diabetes (T2D).

99 Research Design and Methods

100 732 recently diagnosed T2D patients from the IMI-DIRECT study were extensively phenotyped
101 over three years, including measures of insulin sensitivity (OGIS), β-cell glucose sensitivity (GS)
102 and insulin clearance (CLIm) from mixed meal tests, liver enzymes, lipid profiles, and baseline
103 regional fat from MRI. The associations between the longitudinal metabolic patterns and HbA_{1c}
104 deterioration, adjusted for changes in BMI and in diabetes medications, were assessed via stepwise
105 multivariable linear and logistic regression.

106 *Results*

Faster HbA_{1c} progression was independently associated with faster deterioration of OGIS and GS, 107 and increasing CLIm; visceral or liver fat, HDL-cholesterol and triglycerides had further 108 109 independent, though weaker, roles (R^2 =0.38). A subgroup of patients with a markedly higher progression rate (fast progressors) was clearly distinguishable considering these variables only 110 111 (discrimination capacity from AUROC=0.94). The proportion of fast progressors was reduced from 112 56% to 8-10% in subgroups in which only one trait among OGIS, GS and CLIm was relatively 113 stable (odds ratios 0.07 to 0.09). T2D polygenic risk score and baseline pancreatic fat, GLP-1, 114 glucagon, diet, and physical activity did not show an independent role.

115 *Conclusions*

Deteriorating insulin sensitivity and β-cell function, increasing insulin clearance, high visceral or
liver fat, and worsening of the lipid profile are the crucial factors mediating glycemic deterioration

- 118 of T2D patients in the initial phase of the disease. Stabilization of a single trait among insulin
- sensitivity, β -cell function, and insulin clearance may be relevant to prevent progression.

120 Maintaining glucose levels within appropriate limits in patients with type 2 diabetes (T2D) is a 121 crucial factor to prevent complications. Effective strategies to slow glycemic progression can be 122 supported by understanding the processes underlying deterioration of glucose control. 123 Few studies have assessed HbA_{1c} trajectories and the possible determinants of glycemic deterioration. An established finding is that β -cell function decline is an important factor (1,2), 124 125 while contradictory conclusions were drawn for insulin sensitivity (1,3–7). Whether heterogeneous 126 patterns between patients exist in β -cell function and insulin sensitivity decline has not been 127 clarified, an important question for patient stratification and personalized medicine. Other limitations of previous analyses include the incomplete characterization of the metabolic parameters 128 129 affecting glucose homeostasis (derived using fasting data only (2,4)), the restricted set of traits investigated together, and the lack of potentially relevant measures such as ectopic fat, insulin 130 131 clearance, or lifestyle. No study has assessed the relationships between the longitudinal trajectories of HbA_{1c} and those of the other metabolic traits. 132

133 In this analysis, we have used data from the cohort of recently diagnosed and extensively

134 phenotyped T2D patients of the DIRECT study (8,9) to elucidate the processes underlying glycemic 135 deterioration. Specific features of the DIRECT study are the detailed assessment of the glucose 136 homeostasis parameters, and patients all being in the initial phase of the disease. We determined the 137 patterns over a 3-year period of HbA_{1c}, β -cell function, insulin sensitivity and other relevant 138 laboratory, clinical and functional parameters, and assessed their relevance in the deterioration of

- 139 glucose control.
- 140 Research Design and Methods

141 Subjects and protocol

The IMI-DIRECT (Innovative Medicines Initiative - Diabetes Research on Patient Stratification)
project is a multicenter prospective study on northern European adults (8,9) (ClinicalTrials.gov

identifier NCT03814915). The present analysis considers the DIRECT cohort of recently diagnosed 144 145 T2D patients, who were recruited according to the following criteria: white race, T2D diagnosis 146 according to the American Diabetes Association 2011 criteria (10) not less than 6 months and not more than 24 months before baseline examination, previous treatment via lifestyle measures with or 147 without metformin therapy, age between 35 and 74 years, BMI between 20 and 50 kg/m², estimated 148 glomerular filtration rate >50 ml/min, and HbA_{1c} concentration <7.64 % (60.0 mmol/mol) within 149 150 the previous 3 months. Participants were studied at baseline (month 0) and at months 9, 18 and 36. Subjects with HbA_{1c} available at least in two visits were included in this analysis (N=750). 151

All participants provided written informed consent and the study protocol was approved by the
regional research ethics review boards. The research conformed to the ethical principles for medical
research involving human participants outlined in the declaration of Helsinki.

155 *Collected data*

Anthropometric data, HbA_{1c}, blood lipids and liver enzymes were collected at all visits. A 27-month 156 HbA_{1c} sample was collected in 39 patients. A standardized mixed meal test (8) (MMTT) was 157 performed at months 0, 18 and 36 to calculate indices of insulin sensitivity (in fasting conditions, 158 QUICKI (11), and post-MMTT, OGIS (12)), β -cell function (13) (glucose sensitivity, GS, and rate 159 160 sensitivity), and insulin clearance (in fasting conditions, and post-MMTT, CLIm). From the baseline visit we collected glucagon, proinsulin and glucagon-like peptide 1 (GLP-1), measures of 161 regional fat from MRI (8) (available in 561 participants), of physical activity from accelerometer 162 163 (8), and of self-reported 24-hour nutrient intake (8), and we computed the fatty liver index (FLI) (14) and a T2D polygenic risk score (PRS) (15). The whole set of traits considered in this study is 164 165 described in detail in the Supplemental Material (DATA, METHODS, and Table S2).

166 Assessment of progression rates

167 We computed the progression rates for HbA_{1c} and several traits available at follow up 168 (Supplemental Table S4). Each trajectory was described with a conditional linear mixed-effect 169 model (16), in which the longitudinal component of the data was described as a proportional 170 function of time, with normally distributed slopes describing individual progression rates. HbA_{1c} 171 progression was adjusted for changes in BMI and diabetes medications, which were recorded at all 172 visits (as dosage and start and end of treatment). The adjustments were assumed to be 1) 173 proportional to BMI; 2) linearly related to the metformin dose, expressed as percentage of a 174 maximal dose of 3 grams; 3) linearly related to the cumulative dose for the other antidiabetic drugs (insulin excluded), expressed as sum of the percentages of the maximum dose of each drug; 4) 175 176 constant under insulin treatment. A proportional effect of delay in HbA_{1c} assay, i.e. of the difference 177 between the time of measurement and the time of sample collection, was also introduced. 178 Medications were considered to be effective if taken at least 30 days before HbA1c measurement. 179 OGIS and QUICKI trajectories were adjusted for changes in BMI. Further details about the 180 conditional linear mixed-effect models are provided in the Supplemental Material (METHODS). 181 Statistical analysis

Results are presented for participants (*N*=732) with GAD <11 U/ml and islet antigen-2 antibodies
(IA-2) <7.5 U/ml, to exclude other possible forms of diabetes (17). Distributions are described as
mean ± standard deviation. Pairwise associations between continuous variables were assessed using
the Spearman correlation coefficient; differences between groups were assessed using the Wilcoxon
signed rank test (for two groups) and Kruskal-Wallis test (for three or more groups).

187 We used stepwise multivariable linear regression to determine the set of variables, as baseline

values (Table S2) and progression rates (Table S4), independently associated with the HbA_{1c}

189 progression rate, with adjustment for center, sex and age. For baseline variables, both

190 untransformed and transformed values were considered; transformations were logarithmic, or logit

191 when variables where constrained within an interval. The independent variables were included in

the regression model when their effects had p<0.05 and produced an increment in the adjusted R^2 value. Two stepwise analyses were performed: one on all participants, excluding MRI variables from the analysis, and one on the subset of participants with MRI data, including this data in the analysis. Standardized coefficients were computed per standard deviation of the underlying data distribution.

197 Since the distribution of HbA_{1c} progression rates was skewed to the right with a group of patients 198 with high values, we split the subjects into average and fast progressors according to a progression 199 rate threshold (see Results). We used multivariable logistic regression to assess the odds ratios of 200 average vs fast progression, using the independent variables identified in the multiple linear 201 regression analysis of HbA_{1c} progression. The logistic analysis provided values for AUROC, 202 sensitivity, specificity and accuracy, to be used as measures of the discrimination capacity of the investigated independent variables over fast vs average progressors. These parameters must not be 203 204 interpreted as measures of predictive capacity.

205 Role of the funding source

The funders had no role in study design, in collection, analysis, and interpretation of data, in writing of the report, or in the decision to submit the paper for publication. The corresponding author had full access to all data and had final responsibility for the decision to submit for publication.

209 Results

210 Subjects' baseline characteristics

At baseline, the participants had age of 62 ± 8 years, were moderately obese (30.4 ± 4.9 kg/m² BMI),

and had HbA_{1c} of 6.41 ± 0.53 % (46.5 ± 5.8 mmol/mol) and fasting glucose of 7.1 ± 1.4 mmol/l. (Table

S2). 34% of the subjects were treated with metformin at baseline, the rest was treatment naïve.

214 *Progression rates of HbA*_{1c} and other traits

The individual HbA_{1c} progression rates (Supplemental Figure S1), adjusted for changes in BMI and in diabetes medications, were on average only slightly positive and mostly distributed close to their median (median, first and ninth deciles were 0.041, -0.038 and 0.185 %/year (0.45, -0.41 and 2.02 mmol mol⁻¹ year⁻¹), respectively). However, the distribution showed a heavy right tail with values up to 0.897 %/year (9.8 mmol mol⁻¹ year⁻¹). The adjustment of progression rates for BMI changes implied a standardized coefficient for the BMI effect of 0.37.

All the other investigated traits had a mean progression rate per year smaller, in absolute value, than
5% of the corresponding baseline average (see Table S5 for details). On average, waist

circumference, but not BMI, increased very slightly. Insulin sensitivity (as OGIS) and most of the

β-cell function parameters decreased. Fasting, but not post-meal, insulin clearance decreased. Total
cholesterol did not change, while its fractions showed opposite changes, with HDL increasing and
LDL decreasing; TG increased. Creatinine and ALT did not change, while AST and AST/ALT

increased.

228 Several pairwise associations were observed between HbA_{1c} progression rate and laboratory,

clinical, and functional parameters (Supplemental Figure S2). In particular, HbA_{1c} progression rate

230 was clearly associated (p<0.01) with some baseline traits (positively with BMI, waist

circumference, triglycerides, glucagon, liver and visceral fat; inversely with age, HDL, insulin

sensitivity, and β -cell function) and some progression rates (positively with those of triglycerides

and liver enzymes; inversely with those of insulin sensitivity, β -cell function, AST/ALT ratio, and

234 HDL).

Several pairwise associations were also observed between the progression rates of the investigated
traits (Figure S2, panel B). GS and OGIS progression rates were independent of one another despite
HbA_{1c} progression rate being associated with both of them.

238 Variables associated with HbA_{1c} progression rate: multivariable linear analysis

239 In multivariable linear analysis of HbA_{1c} progression rate in all patients, the baseline values and the 240 progression rates of several traits provided an independent contribution (adjusted R^2 0.38; Figure 1, panel A). Faster HbA_{1c} progression was independently associated with lower baseline values and 241 242 faster deterioration of insulin sensitivity (as OGIS) and β -cell function (mostly as glucose sensitivity, GS), with higher baseline values of MMTT insulin clearance, CLIm, and with its 243 244 increase (all p-values <0.001). Faster HbA_{1c} progression was also independently associated with 245 lower baseline HDL (p < 0.05) or its slower increase (p < 0.001), with a quicker increase of TG 246 (p<0.001), as well as with higher baseline values of BMI (p<0.01) and lower baseline values of HbA_{1c} (p<0.001). The variables with strongest effects were the baseline OGIS value and the 247 248 progression rates of OGIS, GS and CLIm (standardized coefficients, in absolute value, between 0.24 and 0.57). 249

In multivariable analysis of the subset of patients with baseline MRI measurements (adjusted R^2 0.40; Figure 1, panel B), baseline visceral fat was positively and independently correlated with HbA_{1c} progression rate; moreover, female sex and younger age independently predicted faster HbA_{1c} progression. The role of the other key metabolic parameters, OGIS, GS and CLIm, remained similar. Replacing visceral fat with liver fat produced similar results (standardized coefficient equal to 0.15 for visceral fat, to 0.11 for liver fat); when both visceral and liver fat were included in the model, the latter was not independently associated with HbA_{1c} progression.

No independent effects were detected for smoking status, family history, T2D polygenic risk score,
baseline values of diet, physical activity, pancreatic fat, GLP-1 (total and intact at fasting, total at 60 min), glucagon, and 60-min proinsulin, baseline values and progression rates of AST and ALT.

Further details on the multivariable linear analysis are reported in the Supplemental Material(RESULTS).

262 Variables associated with HbA_{1c} progression rate: multivariable logistic analysis

263 The threshold selected to separate the heavy right tail of the distribution of HbA_{1c} progression rates 264 was 0.255 %/year (2.79 mmol mol⁻¹ year⁻¹). This threshold split the subjects into average 265 progressors (N=699), with a progression rate of 0.044±0.076 %/year (0.48±0.83 mmol mol⁻¹ year⁻¹), 266 and fast progressors (N=33), with a ~10-fold mean progression rate (0.460±0.185 %/year, 5.03±2.02 mmol mol⁻¹ year⁻¹) (Figure 2). 267 268 We found that the trajectories of most variables independently affecting HbA_{1c} progression as from the linear analysis were clearly different (p < 0.001) in the two groups (Figure 2): in fast progressors, 269 270 OGIS and GS strongly declined and TG and CLIm markedly increased. At baseline, fast

progressors had lower OGIS (p<0.05), CLIm (p<0.01) and HDL (p<0.001), and higher BMI

272 (*p*<0.01).

273 Logistic analysis substantially confirmed the results of linear regression (Figure 1), with half the

investigated variables still contributing (p < 0.05) to distinguish average and fast progressors (Figure

275 3): fast HbA_{1c} progression independently associated with stronger deterioration and a lower

276 baseline value of OGIS and GS, CLIm increase, and HDL reduction. The discrimination capacity of

the logistic model, computed as AUROC, was 0.94 (95% CI between 0.86 and 0.98).

278 Similar outcomes were obtained using lower HbA_{1c} progression rate thresholds, which resulted in

279 larger numbers of patients classified as fast progressors (Supplemental Material - RESULTS,

Figures S1 and S3).

281 At baseline, the percentage of patients treated with metformin were not different between fast

282 progressors (39.4% [24.7-56.3%, 95% CI]) and average progressors (33.9% [30.5-37.5%], *p* =

283 0.64). At the last visit, the percentage of patients treated with any diabetes medication was

somewhat higher in fast progressors, as expected (p = 0.048, details provided in the Supplemental

285 Material - RESULTS). Only 7 average progressors were on insulin at the last visit.

286 Impact of stable OGIS, GS or CLIm on proportion of fast HbA_{1c} progressors

Because HbA_{1c} progression was associated with worsening of three main factors, OGIS, GS and CLIm, we have evaluated the possible importance of maintaining one of these key traits relatively stable in order to avoid fast progression. For this purpose, we considered each trait as deteriorating if its progression rate fell within its worst tertile (the bottom tertile for OGIS and GS, the top one for CLIm), and as stable if it fell in the other two tertiles. We examined the subgroups of patients in which none or only one of these key traits was relatively stable (Table 1).

293 We found that the proportion of fast progressors was 56% in the patient subgroup where GS, OGIS 294 and CLIm were all deteriorating, and decreased to 8-10% in the subgroups where a single trait, 295 either GS, OGIS or CLIm, was stable. All proportions were different from 0 at 90% confidence 296 level, stressing that fast progression did not imply quick changes for each of the three considered 297 traits. All differences in proportions (one stable trait vs none) had p < 0.001, and were associated to 298 odds ratio for fast vs average progression below 0.1 (Table 1); thus, relatively stable progression 299 rate of one single trait among GS, OGIS and CLIm was strongly associated to reduced glycemic 300 deterioration.

301 Conclusions

Leveraging on the detailed participant characterization of the DIRECT study, we have been able to 302 303 elucidate the processes underlying glycemic deterioration in T2D patients in the initial phase of the disease. We found that HbA_{1c} deterioration was independently associated with 1) a decrease in 304 insulin sensitivity; 2) a decrease in β -cell function (primarily β -cell glucose sensitivity); 3) an 305 306 increase in insulin clearance; 4) lower values of insulin sensitivity and glucose sensitivity and higher values of insulin clearance at baseline. Further variables independently associated with faster 307 308 HbA_{1c} progression were declining HDL, increasing TG and high baseline visceral or liver fat. 309 The variables identified by multivariable linear analysis also explained the rapid HbA_{1c} 310 deterioration detected in a subset of patients (identified as fast progressors), the strongest predicting

311 variables of the multivariable linear model being significant also with logistic analysis. Clear

312 differences were evident between fast and average HbA_{1c} progressors (Figure 2), consistent with the 313 associations derived from the multivariable linear analysis. The high discrimination capacity of the 314 logistic analysis suggests that the selected variables capture the most relevant pathophysiological 315 factors underlying glycemic deterioration.

The independent associations with HbA_{1c} progression of several variables, in particular the progression rates of insulin sensitivity, β -cell function and insulin clearance, and the existence of fast HbA_{1c} progressors with relatively stable conditions for any of these three traits (Table 1), indicates 1) that the processes of glycemic deterioration are heterogeneous in this population of T2D patients; 2) that fast progression does not imply quick deterioration of a specific trait, e.g. insulin sensitivity or β -cell function.

The dichotomous analysis shows that the odds for fast *vs* average progression are substantially reduced when either glucose sensitivity, insulin sensitivity or insulin clearance is relatively stable. Although these findings do not demonstrate causality, they suggest that preventing either high degradation rates of glucose sensitivity or insulin sensitivity, or high increase rates of insulin clearance, may be an effective strategy to slow down glycemic deterioration in the initial phase of the disease. This reemphasizes the importance of lifestyle interventions aiming at controlling insulin resistance, as preventing deterioration of the other traits currently appears more difficult.

329 This study also shows that insulin resistance plays a major role in glycemic deterioration in these T2D patients. In particular, we show associations of glycemic deterioration with baseline insulin 330 331 sensitivity and its longitudinal change that the Belfast Diet Study (1), UKPDS (4,18) and ADOPT (6) could not identify, possibly due to differences in subject selection or to the use of post-MMTT 332 333 vs fasting insulin sensitivity indices. We also demonstrate that the associations between glycemic 334 deterioration and insulin sensitivity are independent from both the baseline value and the progression rate of the β -cell function, and that insulin resistance progresses independently from β -335 cell glucose sensitivity. Since in our analysis both HbA_{1c} and insulin sensitivity trajectories were 336

337 adjusted for BMI changes and BMI did not increase on average, we can conclude that worsening of 338 insulin resistance in T2D and the associated glycemic deterioration are partly independent from 339 BMI changes. Whether the observed average increases in TG and AST (whose progression rates were inversely correlated with OGIS progression rate) have a role in insulin sensitivity deterioration 340 (19), and whether this is mediated by ectopic fat accumulation (20), deserves further study. 341 342 UKPDS 25 and 26 (4,18), the Belfast Diet Study (1) and the ADOPT study (6) identified baseline 343 HOMA-%B as a predictor of glycemic deterioration (insulin requirement within 6 years for 344 UKPDS, time of failure to dietary therapy for the Belfast Diet Study, and monotherapy failure before 4 years for ADOPT). Our study confirms the role of β -cell dysfunction as driver of glycemic 345 346 deterioration using a dynamic β -cell function assessment based on a glucose challenge, rather than on fasting data only. We show that both baseline β -cell dysfunction (especially β -cell glucose 347 sensitivity) and its deterioration over time are independently associated with HbA_{1c} worsening. 348 349 Moreover, we demonstrate that patients with limited or absent deterioration in β -cell function have 350 considerably lower odds of rapid glycemic deterioration.

351 Another novel finding is the strong and independent association between HbA_{1c} progression and insulin clearance during the MMTT, CLIm. To our knowledge, this is the first study examining 352 353 insulin clearance trajectories after T2D onset. We found that higher baseline CLIm and faster CLIm 354 increase over time independently associate with faster HbA_{1c} progression. This is consistent with the glucose homeostasis mechanisms, as higher CLIm reduces the average insulin levels. Notably, 355 we found a positive correlation between insulin sensitivity and insulin clearance, considering both 356 357 the baseline values of the two traits, in agreement with previous findings (21), and their progression rates (Figure S2). However, on average, in spite of a decrease in insulin sensitivity, insulin 358 359 clearance did not decrease. These findings show that, while in pre-diabetic subjects insulin 360 clearance reduction may be a way to mitigate the effects of insulin resistance (22), in T2D patients this compensation appears present but impaired and contributing to glycemic deterioration. The 361

362 reasons underlying these results remain elusive. The lack of decrease in insulin clearance may be
363 explained by the decrease of total MMTT insulin secretion and consequent desaturation of insulin
364 utilization (23) only in fast progressors, as in average progressors total insulin secretion slightly
365 increased (Figure 2). Whether hepatic or extrahepatic mechanisms underlie these findings cannot be
366 determined from this study and deserves further investigation.

367 Our results on TG and HDL effects were partially anticipated by a study of the Genetics of Diabetes 368 Audit and Research (GoDARTS) (24), where the outcome was the risk of progression to insulin 369 treatment. The study identified baseline TG and HDL (besides BMI, sex, and age, year and HbA_{1c} 370 at diagnosis) as independent determinants. A later study on the same data (25), investigating the 371 baseline determinants of HbA_{1c} progression rate over about 9 years, confirmed an independent 372 effect of HDL (together with age, BMI and year at diagnosis) but not of TG. The FIELD study in T2D patients on lifestyle measures only revealed that the HDL effect on initiation of oral 373 374 hypoglycemic agents survives the adjustment for HOMA-IR (26). Compared to previous studies 375 (24-26) our analysis includes the progression rates of plasma lipid components and baseline MRI assessment of regional fat. We show that baseline HDL and BMI, and the progression rates of TG 376 377 and HDL are associated with HbA_{1c} progression, even after accounting for the effects of the three 378 main determinants of glucose homeostasis, i.e. insulin sensitivity, β -cell function and insulin 379 clearance. In the subset of participants with MRI data, baseline visceral fat or liver fat was independently correlated with HbA_{1c} progression rate, a further novel observation. These findings 380 381 suggest that additional lipid-dependent factors contribute to HbA_{1c} deterioration, possible candidates being fat accumulation in the viscera (with excessive supply of fatty acids to the liver 382 383 (27)), liver fat and consequent hepatic insulin resistance (28), or glucose overproduction (29). The 384 role of visceral/liver fat supports interventions to reduce ectopic fat as a possible way for slowing 385 future glycemic progression.

Previous studies have reported an inverse correlation between baseline age and HbA_{1c} progression (1,4,6,24,25,30). In our analysis, baseline age does not have a clear independent role in the multivariable model, most likely because the age range is relatively narrow relative to other studies, or because the stronger predictors of HbA_{1c} progression are correlated with age. The latter explanation would suggest that the age univariate effect on glycemic deterioration is indirect. We do not find a clear sex effect in glycemic deterioration, in agreement with most previous studies (1,4,6,24,25).

In the multivariable model, baseline HbA_{1c} was independently and inversely correlated with HbA_{1c} progression rate, in contrast with previous findings (1,4,6,24,30). However, baseline HbA_{1c} was not significant in the logistic model. The most likely explanation of this finding is regression to the mean: indeed, a random decrease in baseline HbA_{1c} can produce a higher estimate of HbA_{1c} progression rate, particularly when the follow-up period is not long, as in our study. Tight glycemic control, an inclusion criterion, may have enhanced this effect.

This study does not find a relevant role of other variables often associated with glucose control. In particular, we did not find an effect of smoking status (reported in GPRD (30)), T2D polygenic risk score (in agreement with GoDARTS (24)), baseline values of diet, physical activity, pancreatic fat, GLP-1, and glucagon. Several of these variables were not associated with HbA_{1c} progression rate even in simple correlation analysis (Figure S2). The lack of association for pancreatic fat is particularly relevant, and contributes to the ongoing discussion on the role of pancreas fat in T2D management (31).

In spite of the unique extensive phenotyping of our study and the consistent results, a significant limitation is the relatively short follow-up period (3 years). The accuracy of the estimated HbA_{1c} progression rate over this time frame may be limited, and in a longer time period the factors contributing to progression may differ. In this study, we could not assess the changes over time of relevant variables such as regional fat by MRI, diet and physical activity. MRI measurements were available only for a subset of subjects. Insulin sensitivity was not derived from the gold standard
euglycemic clamp. As the cohort included only patients of white race, our findings are not
generalizable to other racial/ethnic groups. Causal relationships could not be inferred from our
regression analyses. The study of the mechanisms underlying the deterioration of the factors
affecting HbA_{1c} progression, an important aspect to envisage optimal treatment strategies, also
requires further investigation.

In summary, based on the extensively phenotyped cohort of white European diabetic patients of the DIRECT study, we identified decreasing insulin sensitivity, deteriorating β -cell function, increasing insulin clearance, high liver or visceral fat, and worsening of the lipid profile as the most important factors independently associated with HbA_{1c} deterioration in the early phase of the disease. We also showed that patients with a relatively stable value over time of at least one of insulin sensitivity, β cell glucose sensitivity, or insulin clearance have considerably reduced odds of fast HbA_{1c} increase. This study contributes to the understanding of the factors underlying diabetes progression,

424 elucidating the processes that might be targeted for personalized treatments.

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435 **Duality of Interest.**

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451 Author Contributions.

- 452 R.B. and A.M. designed the analysis, analyzed the data, and wrote the manuscript. R.B., C.J.,
- 453 A.G.J., M.W., E.R.P. and A.M. interpreted the results. E.R.P. and A.M. supervised the analysis.
- 454 C.J., A.G.J., A.K., M.W. and E.R.P. reviewed the manuscript. All authors were involved in the
- 455 DIRECT study at different levels, and were essential for the production, release and management of
- 456 the data analyzed here. R.B. is the guarantor of this work and, as such, takes full responsibility for
- 457 the work as a whole, including the study design, access to data, and the decision to submit and
- 458 publish the manuscript.

459 **References**

- Levy J, Atkinson AB, Bell PM, McCance DR, Hadden DR. Beta-cell deterioration determines
 the onset and rate of progression of secondary dietary failure in Type 2 diabetes mellitus: the
 10-year follow-up of the Belfast Diet Study. Diabet Med. 1998 Apr;15(4):290–6.
- 463 2. U.K. Prospective Diabetes Study Group. U.K. Prospective Diabetes Study 16: Overview of 6
 464 Years' Therapy of Type II Diabetes: A Progressive Disease. Diabetes. 1995 Nov
 465 1;44(11):1249–58.
- Best JD, Drury PL, Davis TME, Taskinen M-R, Kesäniemi YA, Scott R, et al. Glycemic
 Control Over 5 Years in 4,900 People With Type 2 Diabetes. Diabetes Care. 2012
 May;35(5):1165–70.
- 469 4. Matthews DR, Cull CA, Stratton IM, Holman RR, Turner RC. UKPDS 26: sulphonylurea
 470 failure in non-insulin-dependent diabetic patients over six years. Diabet Med. 1998;15(4):297–
 471 303.
- Kahn SE, Haffner SM, Heise MA, Herman WH, Holman RR, Jones NP, et al. Glycemic
 Durability of Rosiglitazone, Metformin, or Glyburide Monotherapy. N Engl J Med. 2006 Dec
 7;355(23):2427–43.
- Kahn SE, Lachin JM, Zinman B, Haffner SM, Aftring RP, Paul G, et al. Effects of
 Rosiglitazone, Glyburide, and Metformin on β-Cell Function and Insulin Sensitivity in
 ADOPT. Diabetes. 2011 May;60(5):1552–60.
- Festa A, Williams K, D'Agostino R, Wagenknecht LE, Haffner SM. The Natural Course of
 beta-Cell Function in Nondiabetic and Diabetic Individuals: The Insulin Resistance
 Atherosclerosis Study. Diabetes. 2006 Apr 1;55(4):1114–20.
- 481 8. Koivula RW, Heggie A, Barnett A, Cederberg H, Hansen TH, Koopman AD, et al. Discovery
 482 of biomarkers for glycaemic deterioration before and after the onset of type 2 diabetes:
 483 rationale and design of the epidemiological studies within the IMI DIRECT Consortium.
 484 Diabetologia. 2014 Jun;57(6):1132–42.
- 485 9. Koivula RW, Forgie IM, Kurbasic A, Viñuela A, Heggie A, Giordano GN, et al. Discovery of
 486 biomarkers for glycaemic deterioration before and after the onset of type 2 diabetes:
 487 descriptive characteristics of the epidemiological studies within the IMI DIRECT Consortium.
 488 Diabetologia. 2019 Sep;62(9):1601–15.
- 489 10. American Diabetes Association. Standards of Medical Care in Diabetes. Diabetes Care.
 490 2011;34(Suppl 1):S11-61.
- 491 11. Katz A, Nambi SS, Mather K, Baron AD, Follmann DA, Sullivan G, et al. Quantitative insulin sensitivity check index: a simple, accurate method for assessing insulin sensitivity in humans.
 493 J Clin Endocrinol Metab. 2000 Jul;85(7):2402–10.
- 494 12. Mari A, Pacini G, Murphy E, Ludvik B, Nolan JJ. A Model-Based Method for Assessing
 495 Insulin Sensitivity From the Oral Glucose Tolerance Test. Diabetes Care. 2001 Mar
 496 1;24(3):539–48.
- 497 13. Mari A, Tura A, Gastaldelli A, Ferrannini E. Assessing Insulin Secretion by Modeling in
 498 Multiple-Meal Tests: Role of Potentiation. Diabetes. 2002 Feb 1;51(Supplement 1):S221–6.

- 499 14. Bedogni G, Bellentani S, Miglioli L, Masutti F, Passalacqua M, Castiglione A, et al. The Fatty
 500 Liver Index: a simple and accurate predictor of hepatic steatosis in the general population.
 501 BMC Gastroenterol. 2006 Dec;6(1):33.
- 502 15. Mahajan A, Taliun D, Thurner M, Robertson NR, Torres JM, Rayner NW, et al. Fine-mapping
 503 type 2 diabetes loci to single-variant resolution using high-density imputation and islet504 specific epigenome maps. Nat Genet. 2018 Nov;50(11):1505–13.
- 505 16. Verbeke G. Conditional Linear Mixed Models. Am Stat. 2001;55(1):25–34.
- 506 17. Hitman GA. The message for MODY. Diabet Med J Br Diabet Assoc. 2011 Sep;28(9):1009.
- 18. Turner R, Stratton I, Horton V, Manley S, Zimmet P, Mackay IR, et al. UKPDS 25:
 autoantibodies to islet-cell cytoplasm and glutamic acid decarboxylase for prediction of insulin
 requirement in type 2 diabetes. The Lancet. 1997 Nov 1;350(9087):1288–93.
- 510 19. Ginsberg HN, Zhang Y-L, Hernandez-Ono A. Regulation of Plasma Triglycerides in Insulin
 511 Resistance and Diabetes. Arch Med Res. 2005 May 1;36(3):232–40.
- 512 20. Kotronen A, Juurinen L, Tiikkainen M, Vehkavaara S, Yki–Järvinen H. Increased Liver Fat,
 513 Impaired Insulin Clearance, and Hepatic and Adipose Tissue Insulin Resistance in Type 2
 514 Diabetes. Gastroenterology. 2008 Jul 1;135(1):122–30.
- Lorenzo C, Hanley AJG, Wagenknecht LE, Rewers MJ, Stefanovski D, Goodarzi MO, et al.
 Relationship of Insulin Sensitivity, Insulin Secretion, and Adiposity With Insulin Clearance in a Multiethnic Population. Diabetes Care. 2013 Jan;36(1):101–3.
- 518 22. Jung S-H, Jung C-H, Reaven GM, Kim SH. Adapting to insulin resistance in obesity: role of insulin secretion and clearance. Diabetologia. 2018 Mar;61(3):681–7.
- 520 23. Ferrannini E, Cobelli C. The kinetics of insulin in man. II. Role of the liver. Diabetes Metab
 521 Rev. 1987 Apr 1;3(2):365–97.
- 522 24. Zhou K, Donnelly LA, Morris AD, Franks PW, Jennison C, Palmer CNA, et al. Clinical and
 523 Genetic Determinants of Progression of Type 2 Diabetes: A DIRECT Study. Diabetes Care.
 524 2014 Mar;37(3):718–24.
- 525 25. Donnelly LA, Zhou K, Doney ASF, Jennison C, Franks PW, Pearson ER. Rates of glycaemic deterioration in a real-world population with type 2 diabetes. Diabetologia. 2018;61(3):607–
 527 15.
- 528 26. Waldman B, Jenkins AJ, Davis TME, Taskinen M-R, Scott R, O'Connell RL, et al. HDL-C
 529 and HDL-C/ApoA-I Predict Long-Term Progression of Glycemia in Established Type 2
 530 Diabetes. Diabetes Care. 2014 Aug 1;37(8):2351–8.
- Sattar N, McConnachie A, Ford I, Gaw A, Cleland SJ, Forouhi NG, et al. Serial metabolic
 measurements and conversion to type 2 diabetes in the west of Scotland coronary prevention
 study: specific elevations in alanine aminotransferase and triglycerides suggest hepatic fat
 accumulation as a potential contributing factor. Diabetes. 2007 Apr;56(4):984–91.
- 535 28. Birkenfeld AL, Shulman GI. Non Alcoholic Fatty Liver Disease, Hepatic Insulin Resistance
 536 and Type 2 Diabetes. Hepatol Baltim Md. 2014 Feb;59(2):713–23.

- Samuel VT, Liu Z-X, Qu X, Elder BD, Bilz S, Befroy D, et al. Mechanism of hepatic insulin
 resistance in non-alcoholic fatty liver disease. J Biol Chem. 2004 Jul 30;279(31):32345–53.
- 30. Cook MN, Girman CJ, Stein PP, Alexander CM, Holman RR. Glycemic Control Continues to
 Deteriorate After Sulfonylureas Are Added to Metformin Among Patients With Type 2
 Diabetes. Diabetes Care. 2005 May 1;28(5):995–1000.
- 542 31. Lim EL, Hollingsworth KG, Aribisala BS, Chen MJ, Mathers JC, Taylor R. Reversal of type 2
 543 diabetes: normalisation of beta cell function in association with decreased pancreas and liver
 544 triacylglycerol. Diabetologia. 2011 Oct 1;54(10):2506–14.

Table 1. Proportion of fast HbA1c progressors with different combinations of stable/deteriorating conditions for GS, OGIS and CLIm progression
 rates.

	Condition*	-					
GS	OGIS	CLIm	Average progressors (N)	Fast progressors (N)	Fast progressors (%) [95% CI]	Odds ratio [95% CI]	p-value [†]
Deteriorating	Deteriorating	Stable	47	5	9.6 [4.2,20.6]	0.09 [0.02,0.32]	2E-4
Deteriorating	Stable	Deteriorating	56	6	9.7 [4.5,19.5]	0.09 [0.02,0.30]	8E-5
Stable	Deteriorating	Deteriorating	34	3	8.1 [2.8,21.3]	0.07 [0.02,0.32]	4E-4
Deteriorating	Deteriorating	Deteriorating	8	10	55.6 [33.7,75.4]	-	-

^{*}The progression rate thresholds dividing stable and deteriorating traits for OGIS, GS and CLIm are -16.68 ml min⁻¹ m⁻² year⁻¹, -4.07 pmol min⁻¹ m⁻² mmol⁻¹ 1

549 year⁻¹ and $0.0184 \, l \, min^{-1} \, m^{-2}$ year⁻¹, respectively.

[†] Two-sided Chi-square test (α =0.05), with Yates continuity correction, on the proportion of fast progressors in the row compared to the same proportion in the last row.

552 GS: β -cell glucose sensitivity; OGIS: oral insulin sensitivity; CLIm: mixed meal test insulin clearance.

553 Figure legends

554 Figure 1. Variables independently associated with HbA_{1c} progression rate from multivariable linear analysis. Panel A: all subjects are included in the analysis (625 with all variables), and MRI 555 measurements are not considered; panel B: only subjects with MRI are included in the analysis (374 556 557 with all variables), and MRI measurements are taken into consideration. For each variable, the figure shows the standardized coefficients \pm 95% CI of the effect. Age and HDL were log-558 559 transformed. OGIS: oral insulin sensitivity; CLIm: mixed meal test insulin clearance; GS: β -cell glucose sensitivity; TG: fasting triacylglycerol; HDL: fasting HDL-cholesterol; RS: β -cell rate 560 sensitivity; progr: progression rate; bas: baseline value; *: p<0.05; **: p<0.01; ***: p<0.001. 561 Figure 2. Temporal trajectories or baseline values (bar graphs) of HbA_{1c} and other key traits in fast 562 (red lines) and average (blue lines) progressors. Data are mean \pm standard error. Simple 563 564 comparisons between fast and average progressors (Wilcoxon rank sum test) are shown for baseline 565 values (asterisks at month 0) and progression rates (asterisks at month 18). These comparisons may 566 differ from the results of the multivariable analyses (Figures 2 and 4). Sex is not included in the 567 figure: males were 42% and 36% in average and fast progressors, respectively (non-significant, 568 Chi-squared test). HbA_{1c} values at 27 months are not displayed as they were collected in a subgroup of individuals. In average progressors, HbA_{1c} increases from 46.4±0.2 mmol/mol to 46.7±0.3 569 570 mmol/mol; in fast progressors, from 48.9±1.21 mmol/mol to 75.7±2.5 mmol/mol. OGIS: insulin 571 sensitivity; CLIm: mixed meal test insulin clearance; GS: β -cell glucose sensitivity; RS: β -cell rate 572 sensitivity; TG: fasting triacylglycerol; HDL: fasting HDL-cholesterol; ISRtot: total mixed meal test insulin secretion; bas: baseline value; *: *p*<0.05; **: *p*<0.01; ***: *p*<0.001. 573 574 Figure 3. Odds ratios \pm 95% CI from the multivariable logistic analysis of fast vs average HbA_{1c} 575 progressors. The independent variables are those identified by multivariable linear analysis of 576 HbA_{1c} progression, excluding MRI variables (*N*=625, with 32 fast progressors and 593 average

577 progressors). Age and HDL were log-transformed. Values for sensitivity, specificity and accuracy

- 578 were derived via maximization of balanced accuracy. OGIS: insulin sensitivity; CLIm: mixed meal
- 579 test insulin clearance; GS: β -cell glucose sensitivity; TG: fasting triacylglycerol; HDL: fasting
- 580 HDL-cholesterol; RS: β -cell rate sensitivity; progr: progression rate; bas: baseline value; AUROC:
- area under the receiver operating characteristics; *: p < 0.05; **: p < 0.01; ***: p < 0.001.