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### An assessment of the effects of neurokinin<sub>1</sub> receptor antagonism against nausea and vomiting: Relative efficacy, sites of action and lessons for future drug development

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**Funding information** There was no specific funding for the publication. A broad-spectrum anti-vomiting effect of neurokinin<sub>1</sub> receptor antagonists (NK<sub>1</sub>RA), shown in pre-clinical animal studies, has been supported by a more limited range of clinical studies in different indications. However, this review suggests that compared with vomiting, the self-reported sensation of nausea is less affected or possibly unaffected (depending on the stimulus) by NK<sub>1</sub> receptor antagonism, a common finding for anti-emetics. The stimulus-independent effects of NK1RAs against vomiting are explicable by actions within the central pattern generator (ventral brainstem) and the nucleus tractus solitarius (NTS; dorsal brainstem), with additional effects on vagal afferent activity for certain stimuli (e.g., highly emetogenic chemotherapy). The central pattern generator and NTS neurones are multifunctional so the notable lack of obvious effects of NK<sub>1</sub>RAs on other reflexes mediated by the same neurones suggests that their anti-vomiting action is dependent on the activation state of the pathway leading to vomiting. Nausea requires activation of cerebral pathways by projection of information from the NTS. Although NK<sub>1</sub> receptors are present in cerebral nuclei implicated in nausea, and imaging studies show very high receptor occupancy at clinically used doses, the variable or limited ability of NK1RAs to inhibit nausea emphasizes: (i) our inadequate understanding of the mechanisms of nausea; and (ii) that classification of a drug as an anti-emetic may give a false impression of efficacy against nausea vs. vomiting. We discuss the potential mechanisms for the differential efficacy of NK<sub>1</sub>RA and the implications for future development of drugs that can effectively treat nausea, an area of unmet clinical need.

#### KEYWORDS

anti-cancer chemotherapy, gastroparesis, motion sickness, nausea, neurokinin\_1, substance P, vomiting

#### 1 | INTRODUCTION

Drugs treating nausea and vomiting as disease symptoms or as adverse effects of therapy are usually classified as *anti-emetics*.

However, the term *emetic* refers to a substance that causes vomiting (or retching). Emesis does not mean nausea. Further, increasing evidence indicates differential efficacy of *anti-emetic* drugs against nausea vs. vomiting. Seifert and Alexander<sup>1</sup> proposed a *rational drug class* 

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*terminology* based on a drug's pharmacological actions rather than its therapeutic orientation (e.g., anti-emetic). Applying this terminology to nausea and vomiting means that the term *anti-emetic* must be written in italics to denote the fact that efficacy against nausea and vomiting should not be assumed to be the same.<sup>2</sup> Here, we emphasize the importance of differentiating between nausea, a self-reported aversive sensation involving cortical and sub-cortical brain regions<sup>3–6</sup> and the mechanical events of retching and vomiting involving multiple brainstem nuclei.<sup>7</sup>

The introduction of neurokinin<sub>1</sub> receptor antagonists (NK<sub>1</sub>RAs) further improved control of *chemotherapy-induced nausea and vomiting* (CINV) and *post-operative nausea and vomiting* (PONV).<sup>8</sup> In addition, a potential expansion of indications may be appropriate, to include, for example, motion sickness.<sup>9</sup> If confirmed, this would point towards a relatively wide spectrum of *anti-emetic* activity for the NK<sub>1</sub>RAs in humans, as suggested by animal studies (see below). However, originating primarily from studies of CINV, there has been a concern that nausea is less well treated than vomiting<sup>10</sup> and this concern persists, as reflected in the comment by Aapro<sup>11</sup> that "Perhaps the greatest unmet need in CINV is the lack of complete nausea control." Accordingly, in an attempt to understand the nausea vs. vomiting question in relation to NK<sub>1</sub>RAs, from both clinical and basic science perspectives, we identified five key questions:

- 1. Has the broad spectrum of activity of NK<sub>1</sub>RAs suggested by animal studies of vomiting translated to humans?
- 2. Where do NK<sub>1</sub>RAs act to inhibit vomiting?
- 3. To what extent do NK<sub>1</sub>RAs inhibit nausea as compared to vomiting?
- 4. If NK<sub>1</sub>RAs have a differential effect against nausea compared to vomiting, what is the explanation?
- 5. What are the implications of the answers to the above questions in terms of patient satisfaction and for future development of drugs to treat nausea?

Different emetic stimuli signal to the brain via different routes. This is why it is first necessary to determine if the broad-spectrum ability of  $NK_1RAs$  to prevent vomiting in animals translates to humans in a similar manner; such a profile directs the discussion on potential mechanism of action against vomiting and nausea. Accordingly, we begin by briefly describing the  $NK_1RA$  studies in animals and then review the effects of  $NK_1RAs$  against vomiting and nausea in different clinical indications (see below for selection criteria), identifying differences in efficacy between these different indications.

#### 2 | ANIMAL STUDIES: SPECTRUM OF NK<sub>1</sub>RA EFFECTS AGAINST VOMITING AND NAUSEA-LIKE BEHAVIOURS

In this section we consider only data from species with a vomiting reflex (ferret, dog, cat, house musk shrew [*Suncus murinus*] and least shrew [*Cryptotis parva*]). To simplify comparisons between species

and between the effects of drugs on vomiting and nausea, we have not considered *nausea-like* behaviour data from rodents, which cannot vomit.<sup>12,13</sup>

#### 2.1 | Vomiting

Studies in multiple animal species (Table 1) have demonstrated *broad-spectrum* effects of NK<sub>1</sub>RAs, markedly reducing/blocking retching and/or vomiting induced by diverse stimuli acting via three key inputs to the brainstem (Figure 1; <sup>7,8</sup> for references).

#### 2.2 | Nausea-like behaviours

Administration to animals of substances inducing nausea and vomiting in humans evoke behavioural changes (often referred to as *nausealike*), but their significance and relevance to the human sensation of nausea is contentious (Chapter  $11^{7,10}$ ). In summary, and in contrast to the clear effects of NK<sub>1</sub>RA on vomiting, effects on nausea-like behaviours (as defined in individual papers) are absent or inconsistent (e.g., Table S1). Given this lack of clarity and since the relevance of these behaviours to the human experience is unknown, they will not be considered further (Chapter 11,<sup>7,10</sup> for detailed discussion).

#### 3 | HUMAN STUDIES: SPECTRUM OF NK1RA EFFECTS AGAINST VOMITING AND NAUSEA

It is important to determine if the broad-spectrum ability of NK1RAs to prevent vomiting in animals translates to the vomiting and nausea of humans. Accordingly, we searched either the name of individual antagonists and/or the therapeutic area (e.g., motion sickness, CINV, PONV, gastroparesis and cyclical vomiting syndrome). For CINV and PONV, where there has been more extensive investigation of NK1RAs anti-emetic efficacy, we initially reviewed systematic reviews/meta-analyses and then analysed data in selected original papers. As our focus was on the relative efficacy of NK1RAs against nausea and vomiting, we included papers where data on both vomiting and nausea were presented and in particular where adequate information was provided in the methods about how each was quantified, with data presented in a form allowing comparison. We note that few studies have given an NK<sub>1</sub>RA alone, n values can be small (e.g., in PONV the *n* value for seven studies of aprepitant included in a meta-analysis ranged from 30 to 55<sup>60</sup>) and some studies are uncontrolled. Nausea is often a secondary outcome with methodological variations in its assessment complicating inter-study comparisons (see below).

Sections 3.1 to 3.7 describe the results of studies investigating the effects of NK<sub>1</sub>RAs against different emetic challenges. Section 3.8 then provides an overview of the spectrum of efficacy against nausea and vomiting.



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**TABLE 1** A summary of pre-clinical studies investigating the efficacy of neurokinin<sub>1</sub> receptor antagonists against retching/vomiting induced by a range of stimuli across multiple species. The studies show the *broad-spectrum* effect of the neurokinin<sub>1</sub> receptor antagonists against stimuli acting via the vestibular system, area postrema or abdominal vagal afferents. See text for discussion of mechanisms.

Species	Neurokinin <sub>1</sub> receptor antagonist	Stimulus details	References
		Cytotoxic anti-cancer drugs	
Acute phase of cispl	atin		
Ferret	CJ-11974 CJ-17493 CP-99994 CP-122721 GR203040 L-742694 L-741671 Netupitant SCH 619734	Given either i.p or i.v.	14-19 20
Dog	FK886 Maropitant		21-24
Suncus	GR203040		25
Doxorubicin emesis	(5 days)		
Dog	Maropitant	i.v.	26
Delayed phase of cis	splatin		
Ferret	CJ-11974 Netupitant SCH619734	Given either i.p or i.v.	14,16,27
Cyclophosphamide			
Ferret	GR203040 GR205171	Given i.p.	25,28
		Pharmacological agents	
Apomorphine			
Dog	CP-99994 FK886 Maropitant	Given s.c.	22,29,20
Ferret	CP-99994 Netupitant SCH619734		14,16,30,20
Brimonidine			
Cat	Maropitant	Sedative given as eye drops	31
Copper sulphate			
Dog	CP-99994	Given p.o.	32,20
Ferret	CP-99994 Netupitant	Given p.o.	16,20
Ethanol			
Suncus	CP-99994	Given i.p.	33
FPL64176			
Least shrew	Netupitant	L-type Ca <sup>++</sup> channel agonist	34
GR73632			
Least shrew	CP-99994	Neurokinin <sub>1</sub> receptor agonist; given i.p.	35
Halothane/ $N_2O$			
Suncus	GR205171	Inhaled	36

(Continues)



#### TABLE 1 (Continued)

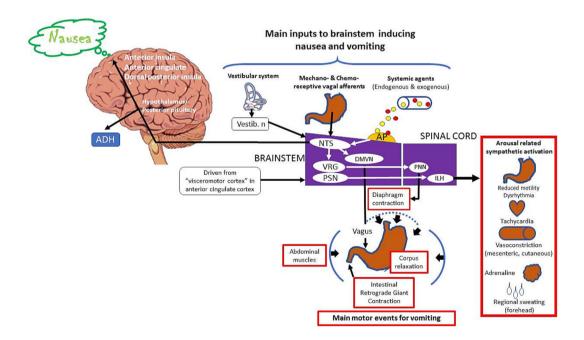
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TABLE 1 (Continu	ied)		
	Neurokinin <sub>1</sub> receptor		
Species	antagonist	Stimulus details	References
Ipecacuanha			
Ferret	CP-99994 CP-122721 GR205171 GR203040 Netupitant R116301	Given p.o.	25,28,37,38,20
Dog	GR203040 GR205171 Maropitant	Given p.o.	25,28,29
Lycorine			
Dog	Maropitant	Alkaloid from daffodils; given s.c.	39
2-methyl 5-hydroxytr	yptamine		
Least shrew	CP-99994	5-HT <sub>3</sub> receptor agonist; given i.p. Note no significant effect of CP-99994 given at same dose that blocked NK <sub>1</sub> agonist (GR73632; see above)	35
Naloxone			
Suncus	CP-99994	Given s.c.	40
Nicotine			
Suncus	CP-99994 CP-122721 RP67580	Given s.c.	30,40
Opiate receptor agon	ists		
Ferret	CP-99994	Loperamide; s.c.	41
Ferret	GR203041	Morphine; s.c.	25
Dog	Maropitant	Morphine; s.c.	42,43
Dog	Maropitant	Morphine; s.c.	44
Dog	Maropitant	Hydromorphone; i.m.	45
Dog	Maropitant	Hydromorphone; i.m.	46
Dog	Maropitant	Hydromorphone; i.m. $+$ acepromazine; i.m.	47
Cat	Maropitant	Dexmedetomidine + morphine; i.m.	48
Phosphodieseterase I			
Ferret	CP-99994	R-rolipram, CT-2450, RS14203; given p.o.	49
Prostaglandin E <sub>2</sub>			
Ferret	CP-99994	Given i.p.	50
Pyrogallol			
Ferret	CP-99994	Reactive oxygen species donor; given i.p.	Andrews and Matsuki, unpublished
Resiniferatoxin			54
Suncus	CP-99994	Given s.c.	51
Tranexamic acid			50
Dog	Maropitant	Fibrinolytic	52
U46619			53
Suncus	CP-99994	TP agonist; given i.p.	
Xylazine			38
Cat	R116301	Given s.c.	50

#### TABLE 1 (Continued)

Species	Neurokinin <sub>1</sub> receptor antagonist	Stimulus details	References
		Non-pharmacological stimuli	
Motion			
Cat	CP-99994	Ferris wheel	54
Dog	Maropitant	Car journey	55
Suncus	GR203040 Netupitant	Horizontal motion	25,16
Total Body Radiation			
Ferret	GR203040 GR205171	X-radiation	25,28
Ferret	CP-99994	X-radiation (3-week post-abdominal vagotomy and greater splanchnic nerve section)	Andrews and Watson, unpublished
Electrical stimulation of v	vagal afferents		
Dog (decerebrate)	GR205171	Stimulation either at the level of the terminal	56,57
Ferret (urethane anaesthesia)	CP-99994	thoracic oesophagus or abdomen; fictive emesis measured in the decerebrate dog.	20
Parvoviral enteritis-induc	ced vomiting		
Dog	Maropitant		58
Post-neurosurgery vomit	ting		
Macaca fascicularis Macaca mulatta	Maropitant		59

Abbreviations: i.m., intramuscularly; i.p, intraperitoneally; i.v., intravenously; p.o., orally; s.c. subcutaneously.



**FIGURE 1** A summary of the major pathways implicated in the motor events of vomiting and the sensation of nausea. The diagram shows the major inputs (vestibular system, abdominal vagal afferents, area postrema) to the nucleus tractus solitarius (NTS) in the brainstem by which both nausea and vomiting are evoked. The mechanical events of vomiting only require activation of brainstem and spinal cord nuclei. Most notable are the dorsal motor vagal nucleus (DMVN) projecting vagal efferents to the digestive tract to induce gastric relaxation and intestinal retrograde giant contraction, and the ventral respiratory group (VRG) of neurones driving the spinal phrenic nerve nucleus (PNN) responsible for contraction of the costal diaphragm, which together with the anterior abdominal muscles (not shown) provides the main force compressing the stomach and leading to forceful oral ejection of contents. Nausea requires activation of cerebral structures and is associated with the secretion of high concentrations vasopressin (AVP/ADH) from the hypothalamic/pituitary axis but other hormones are also released (e.g., cortisol). The main sympathetic motor outputs associated with nausea are shown in the right-hand red rectangle and are a consequence of descending pathways from the *visceromotor cortex* activating the pre-sympathetic nuclei (PSN) in the brainstem, which in turn drive the pre-ganglionic sympathetic neurones in the spinal cord (ILH). For details and references see text. Adapted and modified from previous studies.<sup>6</sup>

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### | Motion sickness

Studies in humans are limited as ethical considerations usually dictate that vomiting endpoints cannot be used in laboratory-based studies inducing motion sickness in healthy human volunteers. Two laboratory-based studies employed the well proven method of highly provocative whole-body rotational motion with head movements to induce motion sickness (so-called cross-coupled motion). These studies showed no significant efficacy of an NK<sub>1</sub>RA (GR205171 [vofopitant]; L-758298) using the degree of motion exposure tolerated before onset of nausea as the endpoint; this suggests no efficacy against nausea.<sup>61,62</sup> A study of healthy human volunteers using inescapable motion at sea investigated the NK1RA tradipitant (VLY-686/ LY686017)<sup>9</sup> and unlike laboratory-based trials, it was possible to measure both vomiting and nausea. Tradipitant was significantly effective (placebo comparator) in protecting against vomiting, but less effective against nausea, using the motion sickness severity scale as an index (Figure 2). Only for selected data obtained during rough seas did the NK<sub>1</sub>RA provide any protection against nausea compared to vomiting in this sub-group (Figure 2). By contrast, well proven muscarinic acetylcholine receptor antagonists such as scopolamine (hyoscine), provided protection against both nausea<sup>63,64</sup> and vomiting.<sup>65</sup> More detailed studies are now required, investigating, for example, the effects of NK<sub>1</sub>RA on the physiological changes accompanying motion sickness such as the reduced gastric antral contractile activity,<sup>66</sup> a pathway of potential relevance to understanding the effects of NK<sub>1</sub>RAs in gastrointestinal conditions associated with nausea, such as gastroparesis (see below).

From these very limited data, we tentatively conclude that NK<sub>1</sub>RAs are effective against *vomiting* induced by abnormal motion but are less effective against *nausea*.

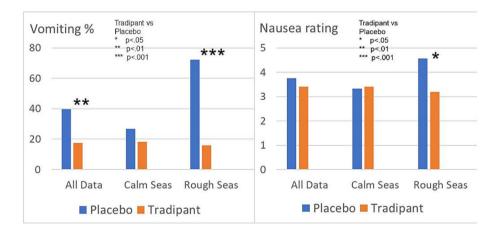
#### 3.2 | CINV

We focus on  $NK_1RA$  use in the acute and delayed phases of highly emetogenic chemotherapy (HEC) discussing their effects against vomiting before effects against nausea.

A study of CINV in seven patients given CP-122721 alone showed that in the acute phase (first 24 h) of HEC five patients had ≤2 episodes vs. 7 episodes of *emesis* in an historic control group and in the delayed phase, six had no emesis.<sup>67</sup> A larger study with L-758298 (the prodrug for the NK<sub>1</sub>RA, aprepitant [L-754030]) showed that 37% of patients (n = 30) had no vomiting or retching in the acute phase, compared with 52% of patients in an ondansetron (5-hydroxytryptamine<sub>3</sub> receptor antagonist; 5-HT<sub>3</sub>RA) group (n = 23; not significantly different).<sup>68</sup> However, confining analysis to the first 8 h following cisplatin showed 37% of patients had no vomiting or retching in the NK<sub>1</sub>RA group compared to 83% in the 5-HT<sub>3</sub>RA group (P = .001) but, in the delayed phase, 72% of patients were without vomiting or retching in the NK<sub>1</sub>RA group vs. 30% in the ondansetron group (P = .005).<sup>68</sup> This study suggests a shift in the relative involvement of 5-HT<sub>3</sub> and NK<sub>1</sub> receptors driving retching and vomiting between the acute and delayed phases following cisplatin, a finding confirmed by detailed time course analysis of the efficacy of aprepitant, L-758298, ondansetron and granisetron in treatment of CINV.<sup>69</sup>

Recent meta-analyses demonstrate additional protection against vomiting when NK<sub>1</sub>RAs are given with a 5-HT<sub>3</sub>RA and dexamethasone during both acute and delayed phases in HEC ( $\sim$ 15–20% more complete protection), with a greater effect in the delayed phase.<sup>70–72</sup>

Overall, and despite an ability of NK<sub>1</sub>RAs to further reduce the incidence of vomiting during the acute phase when combined with a 5-HT<sub>3</sub>RA and dexamethasone, the incidence of nausea is not further reduced during this phase. For example, an initial study with L-754030



**FIGURE 2** The effects of the NK<sub>1</sub> receptor antagonist (NK<sub>1</sub>RA) tradipitant vs. placebo on motion sickness signs and symptoms, are shown for vomiting (left) and for nausea (right). Motion sickness was provoked by motion at sea. Voyages inevitably varied in terms of the weather and roughness of waves; consequently, the data are presented in terms of all data (i.e. all voyages combined) and split by lower wave motion *calm seas* and higher wave motion *rough seas*. Vomiting is shown as % incidence. Nausea is shown as the mean sickness rating scale, with higher scores indicating more severe nausea. Note the differences in levels of statistical significance for the different comparisons. Data were adapted from previous studies.<sup>9</sup>

showed an additional effect on vomiting in the acute phase (day 1) following cisplatin when added to a 5-HT<sub>3</sub>RA/dexamethasone regimen,<sup>73</sup> but no significant reduction in nausea frequency. An analysis of the Phase III studies of NK<sub>1</sub>RAs added to a 5-HT<sub>3</sub>RA and dexamethasone regime in HEC, found no consistent evidence for an improvement in the incidence of no significant nausea (NSN) or no nausea (NN) in the acute phase.<sup>74</sup> For example, the percentage of patients experiencing NN in the NK<sub>1</sub>RA arm vs. placebo in the acute phase was 53.6 vs. 52%,<sup>75</sup> 65 vs. 66%,<sup>76</sup> 68 vs. 61% (Study 2<sup>77</sup>) and 73 vs. 68% (Study 1<sup>77</sup>). A pooled analysis of studies with rolapitant showed a small but statistically significant increase in the percentage of patients reporting NN (respectively, 64 and 70%) in the acute phase of HEC.<sup>74</sup> Saito et al.<sup>78</sup> found a tendency for the incidence of NSN to increase (90.2 vs 84.9%) when using intravenous **fosaprepitant** (150 mg + granisetron/dexamethasone) in patients receiving high-dose cisplatin, although the difference was not statistically significant and the NN incidence was unchanged (67.6 vs. 67.5%) compared to placebo.

Some, but not all, studies reported that during the delayed phase the addition of an NK<sub>1</sub>RA significantly increased the percentage of patients reporting NN or NSN. In the initial study with daily L-754030 (±placebo + granisetron/dexamethasone<sup>73</sup>) the median nausea score was reduced on a 100 mm visual analogue scale (higher score indicating more severe nausea) from 19 to 1 mm on day 2 and over days 2–5 from 10 to 1 mm. Similarly, others reported that the percentage of patients experiencing NN in the NK<sub>1</sub>RA arm vs. placebo/comparator arm in the delayed phase increased significantly: 52.7 vs. 39.9%,<sup>79</sup> 53 vs. 42% (Study 1<sup>77</sup>) and 58 vs. 47% (Study 2<sup>77</sup>). However, some showed no statistically significant change in NN (e.g., 28 vs. 17% days 2–7 but day 2 only, 52 vs. 22%,<sup>68</sup>; 43.9 vs. 49.1%,<sup>75</sup>; 71.4 vs. 73%,<sup>80</sup>; 48 vs. 45%,<sup>76</sup>). A pooled analysis of studies using rolapitant showed a significant 12% increase in the NN percentage (44 vs. 56%) in the delayed phase.<sup>74</sup>

A recent meta-analysis investigated the addition of aprepitant to a 5-HT<sub>3</sub>RA/dexamethasone regimen in patients receiving HEC treatments for lung cancer.<sup>81</sup> While the overall complete response rate (no vomiting/no rescue medication) was significantly better when aprepitant was given, the NN rate was not statistically significantly different.

In summary, there are insufficient data to compare different NK<sub>1</sub>RAs, but it is possible to draw general conclusions about their efficacy in HEC:

- i. NK<sub>1</sub>RAs further reduce the incidence of vomiting during the acute phase when combined with a 5-HT<sub>3</sub>RA and dexamethasone, but the effect is more marked in the delayed phase of HEC.
- When added to a 5-HT<sub>3</sub>RA/dexamethasone regime, the ability of NK<sub>1</sub>RAs to further reduce the incidence of nausea appears inconsistent.

#### 3.3 | Post-operative nausea and vomiting

Table 2 summarizes the effects of  $NK_1RAs$  in PONV using the outcome from studies reporting nausea and vomiting separately to



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illustrate the efficacy differences. Overall, several NK<sub>1</sub>RAs show efficacy against post-operative *vomiting* in a proportion of patients but the block is not complete in all patients and, the efficacy against *nausea* is inconsistent (e.g., small changes in incidence, inconsistent change in intensity, Table 2) and lower than against vomiting. A Cochrane meta-analysis examined the efficacy of diverse pharmacological agents in treating *vomiting* in the first 24 h<sup>95</sup> and concluded that *single* NK<sub>1</sub>RAs were as effective as other *drug combinations*. The analysis did not compare efficacy against nausea.

Assessment of the overall efficacy of NK<sub>1</sub>RAs against PONV is complicated by the variety of types or surgery (e.g., open abdomen, laparoscopic) and anaesthesia/analgesia protocols. A further issue is that in studies where a range of doses has been investigated the relationship between NK<sub>1</sub>RA dose and efficacy against either nausea or vomiting is not always clear (e.g., **casopitant**,<sup>90</sup> rolapitant,<sup>96</sup> vestepitant<sup>88</sup>).

#### 3.4 | Cyclical vomiting syndrome

An open-label uncontrolled trial of aprepitant in a paediatric population refractory to conventional treatment showed reduction in the number of cyclic vomiting episodes/year and number of vomits/h.<sup>97</sup> Although nausea is a feature of cyclical vomiting syndrome it was not assessed in this study.

## 3.5 | Paediatric patients with life-limiting conditions

A case series showed aprepitant (2.0–2.5 mg/kg, intravenously) was effective in complete resolution of nausea (parental reports of impact on mobility and feeding used as proxy efficacy markers) in paediatric patients receiving palliative care, with different diagnoses and unresponsive to at least two drugs classified as *anti-emetics* (e.g., cyclizine, ondansetron, metoclopramide, levomepromazine<sup>98</sup>). Additionally, aprepitant increased the ability to tolerate feeds as might be expected from the proposal that food refusal in children could be used as a surrogate marker for nausea,<sup>99</sup> although NK<sub>1</sub>RA-induced changes in gastric accommodation<sup>100</sup> offers an alternative explanation.

## 3.6 | Gastric distension-induced sensations and gastroparesis

In healthy human volunteers a single dose of aprepitant (80 or 125 mg) had no effect on gastric compliance or sensitivity to distension.<sup>101</sup> Also, in healthy volunteers, aprepitant (125 mg orally [p.o.] day 1 + 80 mg p.o. days 2–5) did not affect gastric emptying of liquids or solids, intestinal or colonic transit.<sup>102</sup> Using the same repeat dosing schedule but following a *dyspeptogenic* meal, Jacob et al.<sup>100</sup> confirmed no change in gastric emptying with aprepitant but found a modest increase in fasting (~10%), postprandial (~9%) and gastric accommodation (~5%)

**TABLE 2** A summary of studies investigating the efficacy of neurokinin<sub>1</sub> receptor antagonists against post-operative nausea and vomiting (PONV). Studies are selected to show the relative efficacy against nausea and vomiting and hence only studies in which they were assessed independently are included. See text for further discussion.

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Compound	Efficacy against nausea in PONV	Reference
CP-122721 (100 mg, 200 mg, p.o.)	In patients undergoing abdominal hysterectomy the maximum nausea score appeared to be reduced by CP-122721 in both dose groups compared to placebo but any effect was not statistically significant ( $n = 20-24$ ). Visual analogue scale nausea score did not differ between ondansetron, CP-122721 and combination groups ( $n = 52-53$ ).	<sup>82</sup> (abstract), <sup>83</sup>
Vofopitant (GR-205171) (25 mg, i.v.)	In patients undergoing major gynaecological surgery vofopitant showed superiority compared to placebo ( $n = 18$ in both groups) for the percentage of patients without nausea (2-h complete control nausea: 55 vs. 20%) and reduced the severity of nausea over the entire 24-h post-operative observation period.	84
Aprepitant (L-754030) (40 mg/125, p.o.)	Peak nausea score distribution (interquartile range) was significantly lower ( $P < .05$ , $n = 280-293$ ) for both aprepitant groups (40/125 mg) compared to ondansetron (4 mg) but the percentage of patients reporting no significant nausea was only significantly higher than that ondansetron for 40-mg aprepitant (62 vs. 53%). For vomiting both doses of aprepitant were superior to ondansetron and blocked vomiting in ~85% of patients. Open abdominal surgery.	85
Aprepitant (L-754030) (80 mg p.o.)	In patients undergoing laparoscopic gynaecological surgery nausea intensity was significantly lower with aprepitant compared to palonosetron on arrival in the recovery room ( $11.2 \pm 2.1 \text{ vs. } 19.0 \pm 2.2$ ) and at 2 h ( $9.7 \pm 2.1 \text{ vs. } 19.4 \pm 3.5$ ) but not in the subsequent 46 h. The complete response rate over 48 h did not differ (74 vs. 77%)	86
Aprepitant (L-754030) (40 mg p.o.)	In patients undergoing plastic surgery compared to placebo (+ondansetron) the severity of nausea was lower ( $P = .014$ , $n = 75$ /arm) in the aprepitant group (+ondansetron) between 0-48-h post-surgery. Vomiting incidence was also significantly lower in the aprepitant group (7/75 vs. 22/75).	87
Vestepitant (6-36 mg, i.v.)	Non-emergency surgery under general anaesthesia in patients failing prophylaxis with pre-surgery ondansetron. Nausea numerical rating scale median values did not differ between ondansetron (4 mg) alone and any dose of vestepitant ( $n = 7-15$ /group) given subsequently but overall vestepitant was superior to ondansetron (10.1–22.9%) improvement except at a dose of 18 mg when there was a $-1.2\%$ difference.	88
Fosaprepitant (150 mg, i.v.)	In patients undergoing surgery requiring general anaesthesia the percentage of patients vomiting was significantly lower with fosaprepitant ( $n = 82$ ) than with ondansetron ( $n = 89$ ) at 0–2 h (2 vs. 17%), 0–24 h (2 vs. 28%) and at 0–48 h (2 vs. 29%). However, the percentage of patients reporting nausea in the fosaprepitant was higher than for vomiting at all time points (e.g., at 0–2 h, nausea 41 vs. vomiting 2%).	89
Casopitant (GW679769) (50 100 150 mg, p.o.)	Only female patients, laparoscopic/laparotomic gynaecological procedure or laparoscopic cholecystectomy. All doses of casopitant further reduced the percentage of patients with vomiting at both 0-24 h (ondansetron 28.6 vs. casopitant + ondansetron 4.3–9.3%) 0–48-h time points (ondansetron 32.9 vs. casopitant + ondansetron 6.4–12.9%). There was no difference in the % of patients reporting nausea between ondansetron and casopitant + ondansetron groups. The % of patients experiencing nausea was higher that the % experiencing vomiting for all three doses of casopitant + ondansetron (casopitant 50 mg, nausea 70.0 vs. vomiting 9.3%; 100 mg, nausea 63.6 vs. vomiting 4.3%; 150 mg, nausea 66.4 vs. vomiting 7.1%). The intensity of nausea did not differ between the three casopitant doses.	90
Aprepitant (L-754030) (40 mg, p.o.)	Craniotomy patients. No difference between nausea scores, incidence or significant nausea between aprepitant and ondansetron (4 mg) up to 48-h post-surgery but the study may not have been sufficiently powered to see statistical differences at all time points.	91
Aprepitant (L-754030) (80 mg, p.o.)	In patients undergoing bariatric surgery aprepitant increased the number of patients without nausea and vomiting (42.18 vs. 36.67%) compared to ondansetron alone and nausea scores were unaffected by aprepitant.	92

#### TABLE 2 (Continued)

Compound	Efficacy against nausea in PONV	Reference
Aprepitant (L-754030) (80 mg, p.o.)	Laparoscopic gynaecological surgery. Significant ( $P = .014$ ) additional reduction in nausea incidence (24 h) when aprepitant was given with ondansetron but no change in severity of nausea or incidence of vomiting.	93
Aprepitant/ rolapitant/ casopitant	Systematic review and meta-analysis of 14 randomized control trials of three neurokinin <sub>1</sub> receptor agonists in patients undergoing mainly either abdominal or gynaecological surgery including open abdomen approaches. Aprepitant (80 mg) showed an additional increase of 31% in the patients protected from nausea compared to placebo.	<sup>94</sup> Note that table 2 in this paper contains a detailed summary of results from all studies included.
Aprepitant (L-754030) (80 mg, p.o.)	Systematic review and meta-analysis of seven randomized control trials of aprepitant (80 mg) in patients undergoing laparoscopic procedures. Risk ratio for nausea 0.56 vs. 0.2 for vomiting compared to placebo or no anti- emetic therapy. Risk of vomiting reduced by 80% in first 2 h post-operatively vs. 44% for nausea.	<sup>60</sup> Note that table 2 in this paper contains a detailed summary of results from all studies included.

Abbreviations: i.v., intravenously; p.o., orally.

volumes, and a tendency to increase maximal tolerated volume ( $\sim$ 25%). Interestingly, the aggregate symptoms, nausea and pain scores (but not bloating or fullness) increased significantly following the dyspeptogenic meal in the aprepitant group compared to placebo (median 36 vs. 4).

A 4-week placebo-controlled study of aprepitant (125 mg/day, p.o.) involving 126 patients failed to demonstrate an improvement in the primary outcome measure of nausea,<sup>103</sup> in a population with 57% gastroparesis patients and the remainder with chronic unexplained nausea and vomiting. The study also used the Gastroparesis Clinical Symptom Index<sup>104</sup> to assess symptom severity as a secondary outcome and this showed significant reductions in overall symptom score (1.3 vs. 0.7), vomiting (1.6 vs. 0.5 [69% decrease]) and nausea (1.8 vs. 1 [44% decrease]). The number of hours per day when nausea was experienced was reduced and the proportion of nausea-free days increased ( $\sim$  twofold).

A placebo-controlled trial of 152 patients with idiopathic or diabetic gastroparesis and moderate-to-severe nausea, investigated tradipitant (85 mg p.o.) twice daily (daily total 170 mg) for 4 weeks.<sup>105</sup> The trial met the primary outcome measure of a reduction in average daily diary nausea score measured using the Gastroparesis Clinical Symptom Index Daily Diary with a difference in score reduction between placebo and tradipitant of  $\sim$ 10%. Nausea severity appeared to begin decreasing by week 2 and this was statistically significant by week 3. Additionally, tradipitant increased secondary outcomes of nausea-free days ( $\sim$ 14% > placebo) and nausea response rate ( $\sim$ 21% > placebo). Patients who responded to tradipitant with a reduction in nausea also had improved early satiety, excessive fullness, bloating and upper abdominal pain, compared to placebo. Two case reports involving single patients with gastroparesis report stoppage of previously intractable nausea<sup>106</sup> or vomiting<sup>107</sup> on administration of aprepitant.

A recent systematic review and network meta-analysis of drugs used to treat gastroparesis showed that NK<sub>1</sub>RAs were efficacious (risk ratio = 0.69) using global symptom score. When individual symptoms were assessed tradipitant was more effective than placebo in treating nausea (tradipitant risk ratio = 0.77; 95% Cl 0.65-0.91).<sup>108</sup> By contrast, a recent phase III trial of tradipitant in gastroparesis showed no difference from placebo in the change of severity of nausea (prespecified primary endpoint) over a 12-week period.<sup>109</sup>

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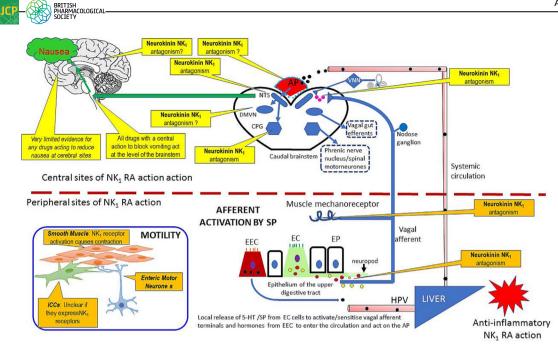
## 3.7 | Overview of clinical efficacy against nausea vs. vomiting

Summarizing Sections 3.1 to 3.7, NK<sub>1</sub>RAs can block vomiting induced by HEC ( $\pm$  5HT<sub>3</sub>RA and dexamethasone) and PONV, and with much more limited evidence perhaps also the vomiting associated with cyclical vomiting syndrome and motion-induced vomiting. NK<sub>1</sub>RAs do not block vomiting in all patients/subjects exposed to a given stimulus and for CINV the efficacy may depend on the phase (potentially, delayed>acute). When nausea is assessed, several studies report no significant benefit although there is some evidence that even if not completely blocking nausea NK<sub>1</sub>RAs may reduce its intensity (e.g., see PONV data, Table 2). Overall, however, the NK<sub>1</sub>RAs are less efficacious or have more variable efficacy against nausea than vomiting over the same range of stimuli but more quantitative data are needed.

We now attempt to explain this differential effect by a detailed analysis of the sites at which  $NK_1RAs$  could act to affect vomiting (Section 4) and nausea (Section 5).

#### 4 | POTENTIAL SITE(S) OF ACTION OF NK<sub>1</sub>RA AGAINST RETCHING AND VOMITING (FIGURE 3)

The sites at which NK<sub>1</sub>RA block retching and vomiting have been investigated in animals (primarily dog and ferret). The findings of these studies are included here because the afferent, integrative and motor pathways responsible for vomiting are comparable between animals



**FIGURE 3** A diagrammatic summary of the central and peripheral sites at which NK<sub>1</sub>RA could act to reduce nausea and vomiting. AP = area postrema; CPG = central pattern generator for vomiting; DMVN = dorsal motor vagal nucleus EC = enterochromaffin cell; EEC = enteroendocrine cell; EP = epithelial cell; HPV = hepatic portal vein; ICC = interstitial cells of Cajal; NK<sub>1</sub>RA = neurokinin<sub>1</sub> receptor antagonist; NTS = nucleus tractus solitarius; VNN = vestibular nerve nucleus. In the periphery, NK<sub>1</sub> receptors located on the gastric smooth muscle, the enteric neurones and possibly the ICCs could modulate motility contributing to a reduction in nausea when disordered motility is implicated (e.g., gastroparesis). NK<sub>1</sub>RA can prevent activation/sensitisation of both muscle mechanoreceptors and epithelial *chemoreceptive* vagal afferents driving nausea and vomiting by locally released SP. The latter are particularly implicated in nausea and vomiting induced by anti-cancer chemotherapy, gastric irritant and some infections (e.g., rotavirus). NK<sub>1</sub> receptors are also implicated in inflammation the reduction of which by NK<sub>1</sub>RA could also contribute to reducing afferent drive. The sites at which vomiting can be blocked all reside in the brainstem (particularly the NTS and CPG), although it is unclear if the AP is a site of action other than when vomiting is induced by an NK<sub>1</sub> receptor agonist. Induction of nausea requires activation of *higher* brain regions and although NK<sub>1</sub> receptors are present at multiple sites in the mid-brain and cerebral hemispheres the data implicating them in anti-nausea effects is circumstantial. See text for details and references.

(e.g., dog, ferret<sup>110</sup>) and humans.<sup>7</sup> For each potential site of action, we will consider whether it could account for a broad-spectrum effect against vomiting or whether it can only explain an action against vomiting induced by a specific stimulus or pathway. This analysis also provides an essential background for understanding the differential effects against nausea.

#### 4.1 | Vestibular system

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The vestibular system is essential for induction of nausea and vomiting caused by abnormal body motion. From an evolutionary perspective the vestibular system is considered a component of the mechanisms protecting the body against ingested toxins (see<sup>111-114</sup>). Although sensitivity to motion sickness is a predictive factor for both CINV and PONV,<sup>115,116</sup> there is no evidence that the vestibular system (including vestibular nuclei) is directly implicated in the induction of either. During motion sickness, the motor pathways for vomiting are activated via projections of the vestibular nuclei to the medial and caudal *nucleus tractus solitarius* (NTS; studies in the cat<sup>117,118</sup>). There is no evidence that NK<sub>1</sub>RAs affect transmission in the pathway between the vestibular system, the vestibular nuclei

and the NTS, to block induction of vomiting. This contrasts with the actions on this pathway of  $H_1$  and muscarinic acetylcholine  $(M_3/M_5)$  receptor antagonists, used to treat motion sickness.<sup>63,64,119</sup> An action of NK<sub>1</sub>RAs within the NTS or at a site(s) deeper in the brainstem is therefore the most likely site for effects against motion-induced vomiting.

#### 4.2 | Area postrema

The area postrema (AP) projects to neurones in the medial NTS (mNTS) which can be activated by emetic stimuli applied to the AP (e.g., **apomorphine**, **L**-glutamate) and by vagal afferent stimulation (dog studies<sup>120</sup>). However, the evidence that NK<sub>1</sub> receptors occur within the AP is weak, and their functional relevance uncertain. For example, low levels of [<sup>3</sup>H]-substance P binding displaced by CP-99994 (0.1–100 nM) were found in the ferret AP, as compared to the NTS (particularly subnucleus gelatinosus).<sup>20</sup> Ariumi et al.<sup>121</sup> reported dense <sup>3</sup>H-substance P binding in the AP and NTS of ferret but displacement by an NK<sub>1</sub>RA was not studied. Comparable evidence is available for *S. murinus* and rat,<sup>122,123</sup> lontophoretic application of substance P (SP) activated ~50% of AP neurones tested (dog<sup>124</sup>), but

although assumed to play a role during vomiting induced by intravenously administered SP (dog<sup>125</sup>), the receptor type activated by the applied concentration of SP and the link between activation and vomiting was not identified. In the ferret, application of SP to the AP can evoke vomiting<sup>123</sup> but microinjection studies<sup>126</sup> suggest that this response was probably due to SP penetration to the subjacent NTS as the blood-brain barrier between these two areas may have some permeability. A similar explanation of leak into the NTS may account for the block in morphine (subcutaneously) and reduction in copper sulphate (intragastric)-induced vomiting in the ferret by administration of the NK<sub>1</sub>RAs CP-99994 or HSP-117 into the AP.<sup>121</sup>

It is a possibility that NK<sub>1</sub> receptors in the AP could be activated if SP (or other tachykinins) are released from gut enteroendocrine cells (EEC)<sup>127</sup> to enter the blood circulation in addition to acting more locally. However, the evidence for this possibility in response to emetic stimuli is weak. Thus, in patients undergoing chemotherapy, the elevation of serum concentrations of SP during the delayed phase of vomiting was inconsistent<sup>128-132</sup> although this is the phase during which NK<sub>1</sub>RA are most effective (see above).

Another possibility is that SP could arise from neurones intrinsic to the AP following direct activation by endogenous or exogenous emetic substances or by abdominal vagal afferents projecting to the AP. However, SP-like immunoreactivity (SP-Li) was absent in the AP of a human infant,<sup>133</sup> consistent with the absence of SP-Li cell bodies in the AP of adult cat, rat<sup>134</sup> and ferret.<sup>135</sup> Previously, extraction studies in humans found some SP in the AP<sup>136,137</sup> and radioligand binding showed a *moderate* uptake of an NK<sub>1</sub>RA by the human AP.<sup>138</sup> Sparse SP-Li nerve fibres have been found in the AP (cat, rat) but their origin is most likely to be from either vagal nerve afferents terminating there or from the NTS<sup>134</sup>; this is consistent with the finding of high densities of SP immunoreactive fibres in lateral borders of the AP in the ferret.<sup>135</sup> However, in the least shrew SP-Li fibres and puncta were present at a moderate level in the AP.<sup>139</sup>

Finally, it is worth noting that the concept of the AP as a site at which systemic agents act to induce nausea and vomiting was originally derived from studies showing abolition of vomiting induced by apomorphine (a **dopamine D**<sub>2</sub> **receptor** agonist), following surgical ablation of the AP including in humans,<sup>140,141</sup> Similarly, other exogenously administered agents (e.g., morphine, **loperamide**, cisplatin) can induce emesis via the AP.<sup>142–144</sup> However, there is only limited evidence that systemic endogenous agents which can induce vomiting (e.g., **adrenaline**, **cholecystokinin**, **GDF15**, arginine**vasopressin**), act via the AP, with alternative sites of action suggested.<sup>143,145,146</sup> The above discussion suggests that SP, acting via NK<sub>1</sub> receptors in the AP should be added to the list of systemic endogenous emetic agents.

#### 4.3 | Abdominal vagal afferents

There are two sites at which vagal afferent activation by emetic stimuli could be affected by an NK<sub>1</sub>RA; they are not mutually exclusive (Figure 3).

#### 4.3.1 | The peripheral transduction mechanism

A potential ability of SP from enterochromaffin cells (ECs) to induce vomiting by acting on vagal afferents was hypothesised >30 years ago (<sup>147</sup>; for details see<sup>8</sup>). Potentially, such a mechanism would be similar to that for 5-HT, which is released from ECs in response to chemotherapeutic agents (e.g., cisplatin) and other emetic stimuli (e.g., rotavirus), causing vomiting by stimulating and sensitizing abdominal vagal afferent terminals via 5-HT<sub>3</sub> receptor activation (<sup>8,148</sup>; for reviews). In rats, treatment with methotrexate or cisplatin increased the number of SP-containing ECs within the intestine, 24 h after administration<sup>149,205</sup> but studies have not yet looked for local release of SP from ECs in response to anti-cancer chemotherapeutic agents or other emetic stimuli. By analogy with 5-HT (see above), any release of SP might be expected to activate vagal nerve terminals. Recently, SP (1 µM)-induced depolarisation of human isolated vagus was shown to be blocked by aprepitant.<sup>150</sup> However, the authors used a concentration (10 µM) at least 10 000× the human NK<sub>1</sub> receptor binding half-maximal inhibitory concentration, at or above the concentrations examined for selectivity of action.<sup>151</sup> and now understood to also activate the mechanosensitive t-pore domain potassium channel. TRAAK (encoded by the KCNK4 gene).<sup>152</sup> Interestingly. recordings from abdominal vagal afferents of ferrets show an interaction between 5-HT and SP<sup>153</sup> and cross talk has been demonstrated between NK1 and 5-HT3 receptors in relation to the anti-emetic effect of palonosetron.<sup>154</sup>

#### 4.3.2 | Vagal afferent to NTS transmission

Abdominal vagal afferents terminate in the mNTS.<sup>155</sup> There is evidence that SP is a transmitter from vagal afferents to NTS neurones (cat<sup>156</sup>; dog<sup>157</sup>) and for activation of NTS neurones by iontophoretically applied SP (ferret<sup>158</sup>; rat<sup>122</sup>). However, any action of NK<sub>1</sub>RA on vagal to NTS transmission must be selective for afferents involved in induction of vomiting as NK<sub>1</sub>RAs do not block the gag reflex, the cardiac or respiratory components of the von Bezold-Jarisch reflex or apnoea induced by cervical vagal afferent stimulation.<sup>20,56</sup> Additionally, while systemic administration of the NK1RA, CP-99994 in the anaesthetized ferret blocked licking, swallowing and retching induced by electrical stimulation of the abdominal vagal afferents, the accompanying rise in blood pressure was unaffected.<sup>20</sup> This makes it unlikely that vagal to NTS transmission per se is blocked and suggests that the block is either within the NTS integrative pathways that initiate vomiting or on the output side of the system in the central pattern generator (CPG) for vomiting located in the reticular formation dorsomedial to the retrofacial nucleus (Bötzinger complex) in the region of the NA (compact region) and the associated prodromal sign centre (PSC in the semi-compact area of the nucleus ambiguus).<sup>155,159,160</sup> Further support for a specific activity on some but not all vagal functions comes from studies in the decerebrate dog where the NK1RA, GR-205171 (intravenously) blocked fictive retching, the accompanying antral contractile response (probably the

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extension of the Retrograde Giant Contraction [RGC] that originates in the small intestine and immediately precedes the onset of retching mediated by vagal efferents; see<sup>161,162</sup>), and reduced the hypersalivation (mediated by PSC) evoked by vagal afferent stimulation, but not the accompanying vagal efferent mediated relaxation of the proximal stomach.<sup>57</sup>

It is self-evident that blockade of vagal afferent activation at a peripheral site or vagal afferent transmission to the mNTS would only contribute to the anti-vomiting effects of NK<sub>1</sub>RAs when the primary stimulus activates the vagus (e.g., acute phase of CINV, possibly gas-troparesis<sup>163</sup>). Therefore, a vagal site of action would not account for block of stimuli acting only either via the AP or the vestibular system so additional site(s) of action need to be considered.

## 4.4 | Brainstem integrative mechanism and the drive to the visceral and somatic motor outputs

The selective effects of NK<sub>1</sub>RA on reflex responses to vagal afferent stimulation (as above) show that actions of NK<sub>1</sub>RA within the brain stem integrative pathways (i.e. NTS, CPG, ventral respiratory group [VRG]) are selective to neurones involved in the *vomiting motor programme* occurring as a result of reconfiguration of the pattern of activity in the multifunctional respiratory neurones (<sup>164,206</sup>) (c.f. cough, yawn, sneeze). These same sets of neurones can also be driven to evoke vomiting by stimuli acting on the vestibular system and the AP (Figure 4). Thus, the effects of NK<sub>1</sub>RAs on the brainstem pathways are *state dependent* and this could explain the selectivity of effects against vomiting; when the brainstem is involved in baseline respiration and some respiratory reflexes there is probably little critical dependence on SP as a transmitter but when the pathway reconfigures and is highly active as occurs for vomiting then it becomes critically dependent on SP.

Overall, there is evidence for either the presence of SP positive neurones and/or  $NK_1$  receptors in the key brainstem sites implicated in vomiting.

#### 4.4.1 | Nucleus tractus solitarius

SP-like immunoreactive neurones are present in the human NTS, particularly subnucleus gelatinosus, and this is consistent with studies in both the cat and ferret.<sup>135,165</sup> A human brain PET study using a fluorine<sup>18</sup>-labelled NK<sub>1</sub>RA reported moderate uptake in the NTS, the nucleus ambiguus and *other nuclei of the vagus* (not specified).<sup>138</sup>

A site of action within the NTS is supported by studies showing microinjection of CP-99994 in the *region of the NTS* inhibited, but did not completely block, cisplatin-induced acute retching and vomiting in the ferret.<sup>19,126</sup> An important point is that the NK<sub>1</sub>RA was injected after retching/vomiting began showing that the antagonist was block-ing a pathway driven by ongoing NK<sub>1</sub> receptor activation. The peptide NK<sub>1</sub>RA, GR-82334 was infective against cisplatin-induced retching/ vomiting when given intravenously but was effective (77% reduction)

when given into the NTS.<sup>126</sup> Rupniak et al.<sup>17</sup> correlated anti-emetic activity against cisplatin in the ferret with central penetration using a range of NK<sub>1</sub>RAs with differing brain penetration. These studies argued strongly that central penetration (at least to the NTS) is required for the acute anti-emetic effect of an NK<sub>1</sub>RA. Further support for an action of NK<sub>1</sub>RA in the NTS comes from inhibition of SP (1  $\mu$ M)-induced discharge in NTS slices by HSP-117 (10  $\mu$ M), without affecting baseline spontaneous neuronal discharge (ferret<sup>158</sup>).

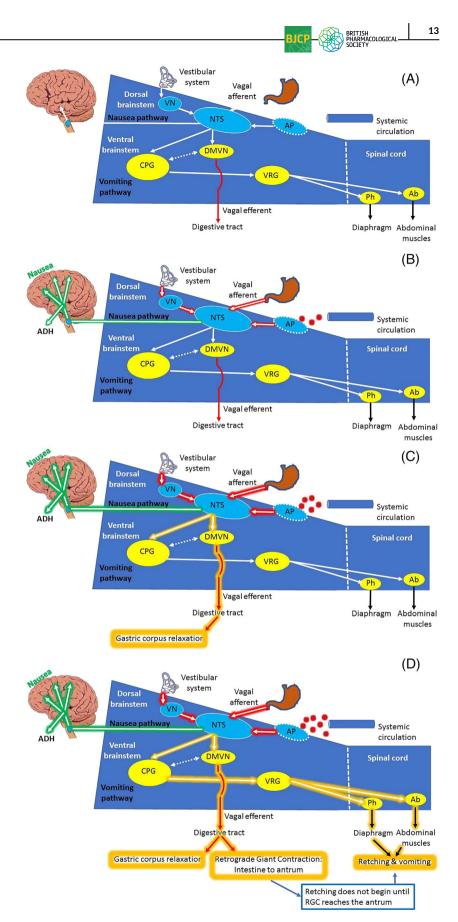
#### 4.4.2 | Dorsal motor vagal nucleus

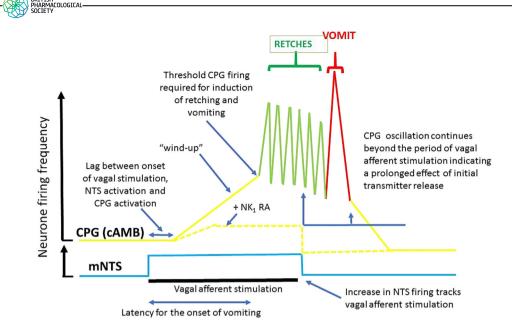
NK<sub>1</sub> receptors are present in the dorsal motor vagal nucleus (DMVN; ferret<sup>20</sup>), the site of origin of vagal efferents supplying the upper digestive tract and regulating the proximal gastric relaxation and RGC prior to the onset of retching and vomiting.<sup>162</sup> In the rat, neurones in the DMVN responsive to gastric distension ±24 h post cisplatin had their baseline activity altered by CP-99994 (5  $\mu$ M)<sup>166</sup> but the results should be interpreted with caution as the efferent projection (e.g., the stomach) of the neurones was not identified (e.g., using antidromic collision<sup>167</sup>) and the effects of CP-99994 were not controlled for by using its less potent 2R, 3R enantiomer, CP-100263.<sup>20</sup> Although these studies show that the DMVN is a potential target for NK<sub>1</sub>RA it should be noted that preventing the gastric relaxation and RGC will not block retching and vomiting as they can occur even in the absence of the stomach<sup>168</sup> and when the RGC is blocked by atropine.<sup>161</sup> An action of NK<sub>1</sub>RA on the DMVN is therefore unlikely to explain their anti-vomiting action.

#### 4.4.3 | Ventral brainstem

Neurophysiological studies of fictive emesis in the dog implicate nuclei in the ventral brainstem.<sup>110,155,159,160</sup> When administered systemically, the NK<sub>1</sub>RA, GR-205171 reduces vagal afferent activation (via the mNTS) of the CPG for vomiting and/or in the pathway linking the NTS to the CPG via the PSC<sup>155,159</sup>; immunohistochemistry has demonstrated the presence of NK<sub>1</sub> receptors in both regions of the dog ventral brainstem.<sup>160</sup> The CPG connects with the VRG, the location of the neurones driving the phrenic and abdominal motor neurones involved in normal respiration as well as retching and vomiting (Figure 4).

Total block of transmission at either the NTS or CPG is probably not required to stop induction of vomiting; a *reduction* in transmission at either site is likely to be sufficient as triggering vomiting requires a higher frequency stimulus, which also lasts for an extended time (e.g., ~20s of vagal afferent stimulation is required in dog<sup>120</sup> and ferret<sup>169</sup>), presumably to prevent inappropriate triggering. It is particularly notable that NK<sub>1</sub>RAs prevent the *wind-up* of CPG neurones induced by vagal afferent stimulation and blunts the rise in firing frequency when continuous vagal afferent stimulation is used, preventing the CPG reaching a threshold for induction of the oscillatory activity required for retching and vomiting<sup>55,159</sup> (Figure 5). FIGURE 4 (A-D) Diagrammatic representation of a longitudinal section through the brainstem showing the key nuclei and pathways implicated in retching, vomiting and nausea. AP = area postrema; CPG = central pattern generator responsible for the generation of the oscillatory pattern of activity driving the somato-motor pathways for retching and vomiting in the VRG; DMVN = dorsal motor nucleus of the vagus, origin of pre-ganglionic efferents to the digestive tract; NTS = nucleus tractus solitarius; VRG = ventral respiratory group of neurones; Ph = phrenic nerve nucleus in cervical (C3-C-5)spinal cord: ab = abdominal muscle motorneurones in ventrolateral thoracic and lumbar spinal cord. See text for further explanation and references. (A) Resting state; (B) low level of activation of pathways inputting to the NTS resulting in activation of NTS and ascending pathways inducing nausea including secretion of anti-diuretic hormone (ADH/AVP) from the posterior pituitary; (C and D) more intense activation of the inputs results in more intense nausea and proximal gastric relaxation, a preparatory action to accommodate refluxed material resulting from the retrograde giant contraction originating in the small intestine when the input is sufficient to exceed the threshold for induction of retching and vomiting when the phrenic and abdominal motor neurones are activated. Note that the CPG and the DMV outputs must be coordinated (dotted arrow) as retching does not begin until the retrograde giant contraction reaches the gastric antrum.





**FIGURE 5** Diagrammatic representation of the likely neuronal discharge pattern in the medial nucleus tractus solitarius (mNTS) and the central pattern generator (located in the compact part of the nucleus ambiguus, cAMB) in response to electrical stimulation of infra-cardiac vagal afferents based on neurophysiological studies in the dog reported in previous works.<sup>110,155,160</sup> Vagal afferent stimulation results in a uniform increase in NTS firing frequency, which ceases at the end of stimulation. NTS activation results in central pattern generator (CPG) activation after a lag period and is followed by a progressive increase in frequency which is due to *wind-up*. The CPG firing frequency reaches at threshold at which the pattern becomes oscillatory with the output driving the ventral respiratory group of neurones (VRG) which in turn drive the phrenic and abdominal motor neurones responsible for the mechanical events of retching a vomiting. The CPG oscillations causing retching are shorter and smaller magnitude than the ultimate burst of activity resulting in vomiting and continue beyond the period of vagal afferent stimulation showing a protracted effect of the initial stimulation.

#### 4.5 | Overview of site(s) of action against vomiting

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The clinically used  $NK_1RAs$  are brain penetrant so when given systemically they can act at both the central and peripheral neuronal sites involved in retching and vomiting:

- i. For vomiting induced by abnormal motion, the brainstem integrative pathways (NTS, CPG) are the most likely site of action.
- ii. For stimuli involving abdominal vagal afferents it is possible that NK<sub>1</sub>RA can: (a) block effects of any SP released from EEC cells onto NK<sub>1</sub> receptors on the peripheral afferent nerve terminals<sup>153</sup>; (b) reduce tachykininergic transmission between vagal afferents and the NTS<sup>123,160</sup>; or (c) modulate the brainstem integrative pathways (NTS, CPG) sufficiently to disrupt the signals encoding induct vomiting.<sup>20,123,159,160</sup> At present, the evidence for (b) and (c) is stronger.
- iii. For stimuli acting on the AP via the circulation (or cerebrospinal fluid) including exogenous emetics and endogenous substances released for example from the digestive tract because of damage/inflammation (e.g., during the delayed phase of CINV and chronic phases of infection; <sup>8,170,171</sup> for references), the brainstem integrative mechanisms (NTS, CPG) are the most likely sites at which vomiting is affected as there is little evidence for an action within the AP itself.

The NTS and CPG sites of action of NK<sub>1</sub>RA are common to all stimuli inducing vomiting. However, for stimuli where abdominal vagal

afferent activation occurs two additional sites of action are implicated, which, if operational, would block vagal afferent input and thereby make it unnecessary for NK<sub>1</sub>RA to act within the NTS and CPG. However, although the NK<sub>1</sub>RA are highly effective against vomiting in a number of clinical settings, NK1 receptors are not the only receptors involved in all of the pathways and this may explain why they may not always be fully effective in all patients. For example, SP is likely to cotransmit with a non-peptide (e.g., glutamate) with the former likely to be released by a higher frequency or different pattern of nerve firing.<sup>172</sup> Further, glutamate has been implicated in abdominal vagal afferent to mNTS transmission as NBQX blocked vagal afferentinduced retching in dog and ferret and the resulting mNTS activation in the dog.<sup>110,173</sup> Nevertheless peptides, as co-transmitters, are known to be involved in network reconfiguration with release determined by both neuronal firing pattern and time.<sup>174</sup> Variations in the predominant transmitters in the nausea and vomiting pathways, possibly as a response to disease, especially if chronic (e.g., in chronic visceral pain NK<sub>1</sub> receptor availability is downregulated<sup>175</sup>), may also contribute to NK<sub>1</sub>RAs spectrum of clinical efficacy.

#### 5 | THE POTENTIAL SITE(S) OF NK<sub>1</sub>RA ACTION AGAINST NAUSEA

Anti-emetics must not be assumed to equally affect both nausea and vomiting.<sup>2</sup> Accordingly, we discuss the relative effects of NK<sub>1</sub>RA against nausea and vomiting by considering specific questions about

the pathways involved; this also informs directions for development of novel drugs (Section 6). Direct experimental data are not available to answer all the questions raised, so some answers are speculative and hypothetical but experimentally testable.

## 5.1 | What information reaches the NTS from the abdominal vagal afferents in the presence of $NK_1RAs$ ?

This question is relevant to both CINV and gastroparesis where abdominal vagal afferents are implicated in genesis of nausea and vomiting.<sup>8,163</sup> Regardless of whether NK<sub>1</sub>RAs reduce vagal afferent firing by acting peripherally (e.g.,<sup>153</sup>) or centrally (e.g.,<sup>160</sup>), the degree of activation, and the pattern, frequency and duration of abdominal vagal afferent activity required for induction of nausea as compared to vomiting is unknown. It is, nevertheless, a reasonable assumption that nausea requires less intense activation of afferent pathways than vomiting (see<sup>176</sup> for discussion in relation to the vagus). The effects of NK<sub>1</sub>RAs on vagal afferent activity evoked by a wide range of stimulus intensities,  $\pm$  substances that may sensitize the afferents (e.g., 5-HT, prostaglandins) need to be investigated directly to answer the above question. The development of vagal afferent recording techniques in humans may eventually allow direct testing of this hypothesis.<sup>177</sup>

# 5.2 | Do differential effects of $NK_1RAs$ on the NTS account for the differential effects against nausea and vomiting?

NK<sub>1</sub>RA modulation of the vagal afferent drive to the NTS and/or transmission within the NTS (vagal, AP and vestibular inputs) could contribute to a *reduction* in nausea *intensity* by decreasing the drive from the NTS to supra-medullary structures implicated in the sensation of nausea. However, the evidence for such an action is poor, as discussed below.

# 5.3 | Are NK<sub>1</sub> receptors in the mid-brain and cerebral hemispheres involved in potential anti-nausea effects of NK<sub>1</sub>RA?

In contrast to vomiting, the brain pathways responsible for nausea are not well defined. The majority of brain imaging studies are in subjects reporting nausea induced by illusory-self motion (vection; visually induced motion sickness), with only single studies using *real* motion or a pharmacological challenge<sup>6</sup> making it difficult to assess whether the findings have general applicability. Cortical and sub-cortical areas consistently showing an increase in activity in healthy volunteers reporting nausea include the frontal lobe (e.g., anterior cingulate cortex), occipital lobe (e.g., posterior cingulate cortex), temporal lobe (e.g., amygdala, part of the *limbic cortex*) and basal ganglia (e.g., putamen).<sup>6</sup>

 $NK_1RA$  binding in the human brain using PET shows  $NK_1$  receptors in several brain areas implicated in nausea. For example,

aprepitant has receptor occupancy of 50% in the caudate and 90% in the putamen (basal ganglia) at plasma concentrations of  $\sim 2 \times 10^{-9}$  M and  $\sim 2 \times 10^{-8}$  M respectively.<sup>178</sup> Based on the striatal occupancy levels, the authors concluded that the recommended *anti-emetic* aprepitant regime of 125 mg on day 1 and 80 mg on the subsequent 2 days in CINV would result in an occupancy of >90%.<sup>177</sup> Hietala et al.,<sup>138</sup> using the same radioligand confirmed the highest uptake in the caudate and putamen and levels  $\sim$ 50% in regions of the occipital lobe (e.g., posterior cingulate cortex), temporal lobe (e.g., amygdala [forms the limbic cortex with the hippocampus]) and frontal lobe (anterior cingulate cortex) all of which have been implicated in nausea in brain imaging studies.<sup>6</sup>

Pharmacological MRI studies provide additional unexpected insights. Using fosaprepitant, the NK<sub>1</sub> receptor distribution profile identified in the above PET studies was confirmed but in addition identified *activation* of brain areas (e.g., cerebellum, red nucleus) where there were thought to not be any NK<sub>1</sub> receptors, an effect attributed to *downstream pharmacodynamic effects*<sup>206</sup> (see fig. <sup>179</sup>). Such effects demonstrate that in identifying brain sites of drug action we should not only consider regions that have their activity inhibited; activation of a pathway that itself is inhibitory on the function under consideration should not be overlooked. Brain imaging studies in nausea have identified areas with both *increased* and *decreased* activity.<sup>3</sup>

Although we focus on areas directly implicated in nausea, as nausea involves heightened anxiety, the potential anxiolytic effects of  $NK_1RA^{180}$  could indirectly contribute to reducing nausea scores especially in chronic conditions (e.g., gastroparesis).

Overall, NK<sub>1</sub>RAs do not appear to have a consistent ability to reduce nausea induced by multiple stimuli despite high levels of NK<sub>1</sub>RA binding in many of the relevant brain areas. Therefore, it is reasonable to conclude that NK<sub>1</sub> receptors do not have a major role in transmission in the *higher* brain regions currently implicated in nausea. We note that NK<sub>1</sub>RA efficacy in depression (e.g.,<sup>181,182</sup>), panic disorder,<sup>183</sup> pain<sup>179</sup> and anxiety<sup>180</sup> are also variable and less than might be anticipated from NK<sub>1</sub> receptor distribution.

#### 5.4 | Do NK<sub>1</sub> RA reduce vasopressin secretion?

Relatively high plasma concentrations of arginine vasopressin (AVP) are associated with nausea induced by stimuli activating the vestibular system, AP and abdominal vagal afferents.<sup>146</sup> A causal link between AVP and nausea is not proven, but a credible possibility in at least some clinical scenarios involves the actions of low concentrations of AVP on gastric pacemaker activity (the interstitial cells of Cajal; ICC), synergising with actions of other nauseagenic stimuli to disrupt motility and hence, initiate vagal afferent discharge; the demonstration of synergy between two different nauseagenic stimuli (adrenaline + AVP) was used to argue that antagonism of one alone (e.g., the effects of vasopressin) might reduce but not prevent the symptom of nausea.<sup>146</sup> In dogs, following cisplatin administration, the NK<sub>1</sub>RA maropitant was without significant effect on the peak AVP concentration

or the area under the curve whereas both were significantly reduced by ondansetron.<sup>23</sup> In human patients treated with cisplatin the acute rise in AVP concentration was blocked by ondansetron<sup>184</sup> as in the dog, but as far as we are aware similar patient studies have not been

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performed with an NK<sub>1</sub>RA.

## 5.5 | Do NK<sub>1</sub>RA have a role in treating nausea by gastric motility modulation?

The presence of SP in the digestive tract in nerve terminals and EEC<sup>185</sup> and of NK<sub>1</sub> receptors on smooth muscle cells and interstitial cells of Caial (ICCs)<sup>186-189</sup> makes the digestive tract a potential target for NK<sub>1</sub>RA. However, an ability of NK<sub>1</sub>RAs to affect nausea by a direct effect on gastric motility is unlikely. Thus, in healthy volunteers there is little evidence for an effect of NK<sub>1</sub>RA on digestive tract motility (assessed by gastric emptying or compliance, or small and large bowel propulsion).<sup>100–102,190</sup> Interestingly, after a dyspeptogenic meal, aprepitant (125 mg on day 1, then 80 mg on days 2-5) increased fasting, postprandial and accommodation gastric volume but increased aggregate symptoms, nausea and pain scores after ingestion of the maximum tolerated volume: the authors suggested that differences between these studies may be dependent on what is measured and on the application of acute- or longer term dosing with aprepitant<sup>100</sup> but activation of TRAAK channels (see above) should also be considered.

Dysrhythmic gastric electrical activity has been associated with nausea in disorders including gastroparesis, chronic unexplained nausea and vomiting, functional dyspepsia, gastro-esophageal reflux disease, all linked with loss of ICCs.<sup>191,192</sup> Thus, any ability of NK<sub>1</sub>RAs to affect ICC functions (see above) could, in theory, have an influence on *induction* of nausea although an effect on vagal afferent signalling or the NTS seems more likely based on current knowledge.

#### 6 | CONCLUDING COMMENTS

Irrespective of the stimulus, the effects of NK<sub>1</sub>RA against *vomiting* are explicable by a central action on the NTS and CPG in the brain stem with potential additional peripheral effects on vagal afferent activity when activated by an emetic stimulus (e.g., HEC, some ingested toxins). NK<sub>1</sub>RAs are not always 100% effective against vomiting in humans (c.f., pre-clinical studies, Table 1) implicating other transmitter/receptor systems and explaining why optimal *anti-vomiting* therapy may require drug combinations (e.g., netupitant + palonosetron + dexamethasone) in treating complex situations such as HEC. An additional role for other neurotransmitters/co-transmitters (e.g., glutamate) has not yet been fully explored.

A reduction in the projection of information from the NTS to the higher brain regions by suppression of NTS pathways and the drive from the abdominal vagal afferents is likely to contribute to any reduction of nausea by  $NK_1RAs$ , no matter how sub-optimal the current evidence suggests. It could be argued that the distribution of

 $NK_1$  receptors in cortical and sub-cortical structures implicated in nausea may predict efficacy against nausea, but it is also possible that these receptors are coupled to non-nauseagenic pathways, such as those involved in fear and/or anxiety (which nonetheless may contribute to the overall sensation of nausea).

Mechanistically, vomiting is well understood and studies with NK<sub>1</sub>RAs show that targeting the NTS/CPG in the brainstem is a valid approach and adverse effects on the respiratory, cardiovascular and digestive systems all regulated from the brainstem appear to be avoided. The apparent specificity of NK<sub>1</sub>RA blockade of vomiting probably reflects the functional reconfiguration of the neural network to coordinate retching/vomiting where tachykininergic signalling becomes critical (state dependence; see<sup>193</sup> for a study of NK<sub>1</sub> receptors and state dependent functions of pre-Bötzinger complex respiratory neurones). The NTS and CPG need investigating in emetic species using neurophysiological studies similar to those in rodents showing complex interaction between NK<sub>1</sub> receptor activation, glutamate and GABA release<sup>194</sup> to understand how NK<sub>1</sub>RAs are *functionally specific* for vomiting.

Nausea remains a challenge as there are major gaps in knowledge of the cerebral pathways involved and hence in identifying potential receptor targets to identify broad-spectrum anti-nausea drugs. As the insular cortex is the highest cortical region consistently activated in subjects reporting nausea,<sup>6</sup> this would be a logical place to target a drug to block nausea, although the associated physiological changes (e.g., regional cold sweating, AVP secretion) may not be blocked as they involve lower brain regions. An alternative approach is to selectively suppress transmission of nauseagenic signals from the NTS to the mid-brain with consideration being given to the parabrachial nucleus as a potential target. While this might be achieved by a combination of receptor antagonists the use of agonists (e.g., GABA<sub>B</sub>, CB<sub>1</sub>, 5-HT<sub>1A</sub>, ghrelin, opioid) may provide a more fruitful approach as this makes fewer assumptions about the nature of the nauseagenic stimulus.<sup>195</sup> A gastric inhibitory polypeptide-1 receptor agonist has been shown to block the acute vomiting induced by the chemotherapeutic agent cisplatin in the ferret, 196 further extending the list of receptor agonists with anti-emetic potential. The electroceutical approaches to treatment of gastrointestinal symptoms, including nausea,<sup>197,198</sup> may provide a route by which this system may be controlled but further study is needed to determine the pathways and cell types involved. A final approach is to target the abdominal vagal afferents at a peripheral site but this would only be applicable when a peripheral release of SP has been demonstrated and when the original signal originates from disordered upper digestive tract function (e.g., gastroparesis<sup>163</sup>). Research into the development of anti-nausea drugs is further hampered by the paucity of human volunteer studies using stimuli other than motion. Studies of anti-emetics have been undertaken in humans using apomorphine, ipecacuanha and morphine as challenges<sup>199-201</sup> and a wider range of challenges could be identified from the side effect profile of licenced drugs (e.g., GLP-1 receptor agonists). The final issue is quantification of nausea. The assessment tools widely used in clinical trials rely on an accurate classification of nausea by

the subject, an assumption that subjects are reporting the same sensation and reliable recollection as data may only be collected daily giving data with a low temporal resolution (see<sup>6</sup> Supporting Information). The heterogeneity of nausea assessment instruments was identified as an issue in a recent US Food and Drug Administration review of endpoints in CINV and PONV studies, which identified nausea assessment as an "opportunity for continued research and development".<sup>202</sup> A reliable, subject independent method for assessing nausea in real time is needed to ensure an accurate assessment of candidate drug efficacy.<sup>10</sup>

We close by dedicating this review to a colleague and friend Wes Miner who died while we were drafting this review. Wes was coauthor of the first paper demonstrating the remarkable anti-emetic effect of a 5-HT<sub>3</sub> receptor antagonist<sup>203</sup> and spent his career in the pharmaceutical industry. In a note to one of the authors (P.L.R.A.) in 1999 Wes made the following insightful comment of relevance to this review regarding the paper<sup>73</sup> reporting some of the earliest clinical data on NK<sub>1</sub>RA: "results are very, very good and I think this will just about wrap it up for pharmaceutical company interest in the N + V area for the next 20 years". As Wes predicted, there have indeed been no major advances in the development in drugs affecting vomiting and especially nausea in the last 20 plus years and as this review shows the accepted dogma that *anti-emetics* equally affect nausea and vomiting requires challenging; a view with which we are sure Wes would concur.

#### AUTHOR CONTRIBUTIONS

All authors made an equivalent contribution.

#### CONFLICT OF INTEREST STATEMENT

P.L.R.A. has no conflict of interest; J.F.G. advises DefenderPharma; G.J.S. advises BYOMass and Neurix.

#### DATA AVAILABILITY STATEMENT

Not applicable.

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#### REFERENCES

- Seifert R, Alexander S. Perspective article: a proposal for rational drug class terminology. Br J Pharmacol. 2022;179(17):4311-4314. doi:10.1111/bph.15916
- Sanger GJ, Andrews PLR. A proposal for rational drug class terminology: a gastrointestinal perspective. Br J Pharmacol. 2022;179(23): 5233-5234. doi:10.1111/bph.15946
- Farmer AD, Ban VF, Coen SJ, et al. Visually induced nausea causes characteristic changes in cerebral, autonomic and endocrine function in humans. J Physiol. 2015;593(5):1183-1196. doi:10.1113/jphysiol. 2014.284240
- Napadow V, Sheehan JD, LaCount LT, et al. The brain circuitry underlying the temporal evolution of nausea in humans. *Cereb Cor*tex. 2013;23(4):806-813. doi:10.1093/cercor/bhs073
- 5. Ruffle JK, Patel A, Giampietro V, et al. Functional brain networks and neuroanatomy underpinning nausea severity can predict nausea

susceptibility using machine learning. J Physiol. 2019;597(6):1517-1529. doi:10.1113/JP277474

- Varangot-Reille C, Sanger GJ, Andrews PLR, et al. Neural networks involved in nausea in adult humans: a systematic review. *Journal of the Autonomic Nervous System*. 2023;245:103059. doi:10.1016/j. autneu.2022.103059
- Stern RM, Koch KL, Andrews PLR. Nausea: mechanisms and management. Oxford University Press; 2011:462.
- Sanger GJ, Andrews PLR. A history of drug discovery for treatment of nausea and vomiting and the implications for future research. *Front Pharmacol.* 2018;9:913. doi:10.3389/fphar.2018.00913
- Polymeropoulos VM, Czeisler MÉ, Gibson MM, et al. Tradipitant in the treatment of motion sickness: a randomized, double blind, placebo-controlled study. *Front Neurol.* 2020;11:563373. doi:10. 3389/fneur.2020.563373
- Andrews PLR, Sanger GJ. Nausea and the quest for the perfect antiemetic. *Eur J Pharmacol.* 2014;722:108-121. doi:10.1016/j.ejphar. 2013.09.072
- Aapro M. CINV: still troubling patients after all these years. Support Care Cancer. 2018;26(Suppl 1):55-59. doi:10.1007/s00520-018-4131-3
- Horn CC, Kimball BA, Wang H, et al. Why can't rodents vomit? A comparative behavioral, anatomical, and physiological study. *PLoS ONE*. 2013;8(4):e60537. doi:10.1371/journal.pone.0060537
- Sanger GJ, Holbrook JD, Andrews PLR. The translational value of rodent gastrointestinal functions: a cautionary tale. *Trends Pharmacol Sci.* 2011;32(7):402-409. doi:10.1016/j.tips.2011.03.009
- Duffy RA, Morgan C, Naylor R, et al. Rolapitant (SCH 619734): a potent, selective and orally active neurokinin NK1 receptor antagonist with centrally-mediated antiemetic effects in ferrets. *Pharmacol Biochem Behav.* 2012;102(1):95-100. doi:10.1016/j.pbb.2012. 03.021
- Lau AHY, Kan KKW, Lai HW, et al. Action of ondansetron and CP-99, 994 to modify behavior and antagonize cisplatin-induced emesis in the ferret. *Eur J Pharmacol.* 2005;506(3):241-247. doi:10.1016/j. ejphar.2004.11.019
- Rudd JA, Ngan MP, Lu Z, et al. Profile of antiemetic activity of netupitant alone or in combination with palonosetron and dexamethasone in ferrets and *Suncus murinus* (house musk shrew). *Front Pharmacol.* 2016;7:263. doi:10.3389/fphar.2016.00263
- Rupniak NMJ, Tattersall FD, Williams AR, et al. In vitro and in vivo predictors of the anti-emetic activity of tachykinin NK1 receptor antagonists. *Eur J Pharmacol.* 1997;326(2–3):201-209. doi:10.1016/ S0014-2999(97)85415-5
- Shishido Y, Wakabayashi H, Koike H, et al. Discovery and stereoselective synthesis of the novel isochroman neurokinin-1 receptor antagonist CJ-17,493. *Bioorg Med Chem.* 2008;16(15):7193-7205. doi:10.1016/j.bmc.2008.06.047
- Tattersall FD, Rycroft W, Francis B, et al. Tachykinin NK<sub>1</sub> receptor antagonists act centrally to inhibit emesis induced by the chemotherapeutic agent cisplatin in ferrets. *Neuropharmacology*. 1996; 35(8):1121-1129. doi:10.1016/S0028-3908(96)00020-2
- Watson JW, Gonsalves SF, Fossa AA, et al. The anti-emetic effects of CP-99,994 in the ferret and the dog: role of NK<sub>1</sub> receptors. Br J Pharmacol. 1995;115(1):84-94. doi:10.1111/j.1476-5381.1995.tb16324.x
- de la Puente-Redondo VA, Tilt R, Rowan TG, Clemence RG. Efficacy of maropitant for treatment and prevention of emesis caused by intravenous infusion of cisplatin in dogs. *Am J Vet Res.* 2007;68(1): 48-56. doi:10.2460/ajvr.68.1.48
- Furukawa TY, Nakayama H, Kikuchi A, et al. Antiemetic effects of a potent and selective neurokinin-1 receptor antagonist, FK886, on cisplatin- and apomorphine-induced emesis in dogs. *Biol Pharm Bull*. 2013;36(6):974-979. doi:10.1248/bpb.b12-01079
- 23. Kenward H, Elliott J, Lee T, Pelligrand L. Anti-nausea effects and pharmacokinetics of ondansetron, maropitant and metoclopramide

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in a low-dose cisplatin model of nausea and vomiting in the dog: a blinded crossover study. *BMC Vet Res.* 2017;13(1):244. doi:10.1186/s12917-017-1156-7

- Vail DM, Rodabaugh HS, Conder GA, Boucher JF, Mathur S. Efficacy of injectable maropitant (Cerenia) in a randomized clinical trial for prevention and treatment of cisplatin-induced emesis in dogs presented as veterinary patients. *Vet Comp Oncol.* 2007;5(1):38-46. doi: 10.1111/j.1476-5829.2006.00123.x
- Gardner CJ, Twissell DJ, Gale JD, et al. The broad-spectrum antiemetic activity of the novel non-peptide tachykinin NK<sub>1</sub> receptor antagonist GR203040. Br J Pharmacol. 1995;116(8):3158-3163. doi: 10.1111/j.1476-5381.1995.tb15118.x
- Rau SE, Barber LG, Burgess KE. Efficacy of maropitant in the prevention of delayed vomiting associated with administration of doxorubicin in dogs. J Vet Intern Med. 2010;24(6):1452-1457. doi:10.1111/j. 1939-1676.2010.0611.x
- Tsuchiya M, Fujiwara Y, Kanai Y, et al. Anti-emetic activity of the novel nonpeptide tachykinin NK1 receptor antagonist ezlopitant (CJ-11,974) against acute and delayed cisplatin-induced emesis in the ferret. *Pharmacology*. 2002;66(3):144-152. doi:10.1159/ 000063796
- Gardner CJ, Armour DR, Beattie DT, et al. GR205171: a novel antagonist with high affinity for the tachykinin NK<sub>1</sub> receptor, a potent broad-spectrum anti-emetic activity. *Regul Pept.* 1996;65(1):45-53. doi:10.1016/0167-0115(96)00071-7
- Sedlacek HS, Ramsy DS, Boucher JF, Eagleson JS, Conder GA, Clemence RG. Comparative efficacy of maropitant and selected drugs in preventing emesis induced by centrally or peripherally acting emetogens in dogs. J Vet Pharmacol Ther. 2008;31(6):533-537. doi:10.1111/j.1365-2885.2008.00991.x
- Tattersall FD, Rycroft W, Marmont N, Cascieri M, Hill RG, Hargreaves RJ. Enantiospecific inhibition of emesis induced by nicotine in the house musk shrew (*Suncus murinus*) by the neurokinin1 (NK1) receptor antagonist CP-99, 994. *Neuropharmacology*. 1995; 34(12):1697-1699. doi:10.1016/0028-3908(95)00164-6
- Kanda T, Toda C, Morimoto H, et al. Anti-emetic effect of oral maropitant treatment before the administration of brimonidine ophthalmic solution in healthy cats. J Feline Med Surg. 2020;22(6):557-563. doi:10.1177/1098612X19862687
- Andrews PLR, Kovacs M, Watson JW. The anti-emetic action of the neurokinin(1) receptor antagonist CP-99,994 does not require the presence of the area postrema in the dog. *Neurosci Lett.* 2001; 314(1-2):102-104. doi:10.1016/S0304-3940(01)02269-8
- Chen Y, Saito H, Matsuki N. Ethanol-induced emesis in the house musk shrew, Suncus murinus. Life Sci. 1996;60(4-5):253-261. doi:10. 1016/S0024-3205(96)00625-X
- Zhong W, Chebolu S, Darmani NA. Intracellular emetic signaling evoked by the L-type Ca<sup>2+</sup> channel agonist FPL64176 in the least shrew (*Cryptotis parva*). *Eur J Pharmacol*. 2018;834:157-168. doi:10. 1016/j.ejphar.2018.06.035
- Darmani NA, Chebolu S, Amos B, Alkam T. Synergistic antiemetic interactions between serotonergic 5-HT<sub>3</sub> and tachykininergic NK<sub>1</sub>receptor antagonists in the least shrew (*Cryptotis parva*). *Pharmacol Biochem Behav*. 2011;99(4):573-579. doi:10.1016/j.pbb.2011. 05.025
- Gardner C, Perrin M. Inhibition of anaesthetic-induced emesis by a NK<sub>1</sub> or 5-HT<sub>3</sub> receptor antagonist in the house musk shrew, *Suncus murinus*. *Neuropharmacology*. 1998;37(12):1643-1644. doi:10.1016/ S0028-3908(98)00133-6
- Gonsalves S, Watson J, Ashton C. Broad spectrum antiemetic effects of CP-122721, a tachykinin NK<sub>1</sub> receptor antagonist in ferrets. *Eur J Pharmacol.* 1996;305(1-3):181-185. doi:10.1016/0014-2999(96) 00216-6
- Megens AAHP, Ashton D, Vermeire JC, et al. Pharmacological profile of (2r-trans)-4-[1-[3,5-bis (trifluromethyl)benzoyl]-2-

(phenylmethyl)-4-piperidinyl]-n-(2,6-dimethylphenyl)-1-acetamide (s)hydroxybutanedioate (R116301), an orally and centrally active neurokinin-1 receptor antagonist. *J Pharmacol Exp Ther.* 2002;302(2): 696-709. doi:10.1124/jpet.102.034348

- Kretzing S, Abraham G, Seiwert B, et al. In vivo assessment of antiemetic drugs and mechanism of lycorine-induced nausea and emesis. *Arch Toxicol.* 2011;85(12):1565-1573. doi:10.1007/s00204-011-0719-9
- Rudd JA, Ngan MP, Wai MK. Inhibition of emesis by tachykinin NK1 receptor antagonists in *Suncus murinus* (house musk shrew). *Eur J Pharmacol.* 1999;366(2-3):243-252. doi:10.1016/S0014-2999(98) 00920-0
- Zaman S, Woods AJ, Watson JW, Reynolds DJM, Andrews PLR. The effect of the NK1 receptor antagonist CP-99,994 on emesis and c-fos protein induction by loperamide in the ferret. *Neuropharmacol*ogy. 2000;39(2):316-323. doi:10.1016/S0028-3908(99)00113-6
- Lorenzutti AM, Martin-Flores M, Literio NJ, Himelfarb MA, Invaldi SH, Zarazaga MP. Evaluation of the antiemetic efficacy of maropitant in dogs medicated with morphine and acepromazine. *Journal of Veterinary Anaesthesia*. 2016;43(2):195-198. doi:10.1111/ vaa.12286
- Lorenzutti AM, Martin-Flores M, Literio NJ, Himelfarb MA, Invaldi SH, Zarazaga MP. A comparison between maropitant and metoclopramide for the prevention of morphine-induced nausea and vomiting in dogs. *Can Vet J.* 2017;58(1):35-38.
- Koh RB, Isaza N, Xie H, Cooke K, Robertson SA. Effects of maropitant, acepromazine, and electroacupuncture on vomiting associated with administration of morphine in dogs. JAVMA. 2014;244(7):820-829. doi:10.2460/javma.244.7.820
- Claude AK, Dedeaux A, Chiavaccini L, Hinz S. Effects of maropitant citrate or acepromazine on the incidence of adverse events associated with hydromorphone premedication in dogs. J Vet Intern Med. 2014;28(5):1414-1417. doi:10.1111/jvim.12414
- Hay-Kraus BL. Efficacy of orally administered maropitant citrate in preventing vomiting associated with hydromorphone administration in dogs. J Am Vet Med Assoc. 2014;244(10):1164-1169. doi:10. 2460/javma.244.10.1164
- Johnson RA. Maropitant prevented vomiting but not gastroesophageal reflux in anesthetized dogs premedicated with acepromazinehydromorphone. Vet Anesth Analg. 2014;41(4):406-410. doi:10. 1111/vaa.12120
- Martin-Flores M, Sakai DM, Learn MM, et al. Effect of maropitant in cats receiving dexamethasone and morphine. J Am Vet Med Assoc. 2016;248(11):1257-1261. doi:10.2460/javma.248.11.1257
- Robichaud A, Tattersall FD, Choudry I, Rodger IW. Emesis induced by inhibitors of type IV cyclic nucleotide phosphodiesterase (PDE IV) in the ferret. *Neuropharmacology*. 1999;38(2):289-297. doi:10. 1016/S0028-3908(98)00190-7
- Kan KKW, Rudd JA, Wai MK. Differential action of anti-emetic drugs on defecation and emesis induced by prostaglandin E2 in the ferret. *Eur J Pharmacol.* 2006;544(1–3):153-159. doi:10.1016/j. ejphar.2006.06.034
- Andrews PLR, Okada F, Woods AJ, et al. The emetic and anti-emetic effects of the capsaicin analogue resiniferatoxin in *Suncus murinus*, the house musk shrew. Br J Pharmacol. 2000;130(6):1247-1254. doi: 10.1038/sj.bjp.0703428
- 52. Kantyka ME, Meira C, Bettschart-Wolfensberger R, Hartnack S, Kutter APN. Prospective, controlled, blinded, randomized crossover trial evaluating the effect of maropitant versus ondansetron on inhibiting tranexamic acid-evoked emesis. J Vet Emerg Crit Care. 2020; 30(4):436-441. doi:10.1111/vec.12954
- Kan KKW, Jones RL, Ngan M-P, Rudd JA, Wai M-K. Action of prostanoids on the emetic reflex of *Suncus murinus* (the house musk shrew). *Eur J Pharmacol*. 2003;477(3):247-251. doi:10.1016/j.ejphar. 2003.08.020

- 54. Lucot JB, Obach RS, McClean S, Watson JW. The effect of CP-99994 on the responses to provocative motion in the cat. Br J Pharmacol. 1997;120(1):116-120. doi:10.1038/sj.bjp.0700888
- 55. Conder GA, Sedlacek HS, Boucher JF, Clemence RG. Efficacy and safety of maropitant, a selective neurokinin 1 receptor antagonist, in to randomized clinical trials for prevention of vomiting due to motion sickness in dogs. J Vet Pharmacol Ther. 2008;31(6):528-532. doi:10.1111/j.1365-2885.2008.00990.x
- Fukuda H, Koga T, Furukawa N, Nakamura E, Shiroshita Y. The tachykinin NK1 receptor antagonist GR205171 abolishes the retching activity of neurones comprising the central pattern generator for vomiting in dogs. *Neurosci Res.* 1999;33(1):25-32. doi:10. 1016/S0168-0102(98)00106-0
- Furukawa N, Fukuda H, Hatano M, Koga T, Shiroshita Y. A neurokinin-1 receptor antagonists reduce hypersalivation and gastric contractility related to emesis in dogs. *Am J Physiol*. 1998;275(5): G1193-G1201. doi:10.1152/ajpgi.1998.275.5.G1193
- Yalcin E, Keser GO. Comparative efficacy of metoclopramide, ondansetron and maropitant in preventing parvoviral enteritisinduced emesis in dogs. J Vet Pharmacol Ther. 2017;40(6):599-603. doi:10.1111/jvp.12396
- Steinbach JR, MacGuire J, Chang S, Dierks E, Roble GS. Assessment of pre-operative maropitant citrate use in macaque (*Macaca fascicularis & Macaca mulatta*) neurosurgical procedures. J Med Primatol. 2018;47(3):178-184. doi:10.1111/jmp.12343
- 60. Cavaye J, Dai B, Gurunathan K, Weir RM, Yerkovich S, Gurunathan U. NK1 receptor antagonists versus other antiemetics in the prevention of postoperative nausea and vomiting following laparascopic surgical procedures: a systematic review and metaanalysis. J Anesthesiol Clin Pharmacol. 2021;38:35-37.
- 61. Reid K, Sciberras DG, Gertz BJ, et al. Comparison of a neurokinin-1 antagonist L758,298, and scopolamine with placebo in the prevention of motion-induced nausea in man. *Br J Clin Pharmacol*. 1998;45: 282P.
- 62. Reid K, Palmer JL, Wright RJ, et al. Comparison of the neurokinin-1 antagonist GR205171, alone and in combination with the 5-HT3 antagonist ondansetron, hyoscine and placebo in the prevention of motion-induced nausea in man. *Br J Clin Pharmacol.* 2000;50(1):61-64. doi:10.1046/j.1365-2125.2000.00221.x
- Golding JF, Wesnes KA, Leaker BR. The effects of the selective muscarinic M<sub>3</sub> receptor antagonist darifenacin, and of hyoscine (scopolamine), on motion sickness, skin conductance & cognitive function. Br J Clin Pharmacol. 2018;84(7):1535-1543. doi:10.1111/bcp.13579
- Golding JF, Stott JRR. Comparison of the effects of a selective muscarinic receptor antagonist and hyoscine (scopolamine) on motion sickness, skin conductance and heart rate. *Br J Clin Pharmacol*. 1997; 43(6):633-637. doi:10.1046/j.1365-2125.1997.00606.x
- Golding JF, Paillard AC, Normand H, Besnard S, Denise P. Prevalence, predictors & prevention of motion sickness in zero-G parabolic flights. Aerosp Med Hum Perform. 2017;88(1):3-9. doi:10.3357/ AMHP.4705.2017
- Faas H, Feinle C, Enck P, Grundy DG, Boesiger P. Modulation of gastric motor activity by centrally acting stimulus, circular vection, in humans. Am J Physiol Gastrointest Liver Physiol. 2001;280(5):G850-G857. doi:10.1152/ajpgi.2001.280.5.G850
- Kris MG, Radford JE, Pizzo BA, Inaninet R, Hesketh A, Hesketh PJ. Use of an NK<sub>1</sub> receptor antagonist to prevent delayed emesis after cisplatin. J Natl Cancer Inst. 1997;89(11):817-818. doi:10.1093/jnci/ 89.11.817
- Cocquyt V, Van Belle S, Reinhardt RR, et al. Comparison of L-758,298, a prodrug for the selective neurokinin-1 antagonist, L-754,030, with ondansetron for the prevention of cisplatin-induced emesis. *Eur J Cancer*. 2001;37(7):835-842. doi:10.1016/S0959-8049 (00)00416-0

- Hesketh PJ, Van Belle S, Aapro M, et al. Differential involvement of neurotransmitters through the time course of cisplatin-induced emesis revealed by therapy with specific receptor antagonists. *Eur J Cancer.* 2003;39(8):1074-1080. doi:10.1016/S0959-8049(02)00674-3
- Jordan K, Warr DG, Hinke A, Sun L, Hesketh PJ. Defining the efficacy of neurokinin-1 receptor antagonists in controlling chemotherapy-induced nausea and vomiting in different emetogenic settings—a meta-analysis. *Support Care Cancer*. 2016;24(5):1941-1954. doi:10.1007/s00520-015-2990-4
- Qiu T, Men P, Xu X, Zhai S, Cui X. Antiemetic regimen with aprepitant in the prevention of chemotherapy-induced nausea and vomiting. An updated systematic review and meta-analysis. *Medicine*. 2020;99(33):e21559. doi:10.1097/MD.00000000021559
- Yokoe T, Hayashida T, Nagayama A, et al. Effectiveness of anti-emetic regimens for highly emetogenic chemotherapy-induced nausea and vomiting: a systematic review and network meta-analysis. *Oncologist*. 2019;24(6):e347-e357. doi:10.1634/theoncologist.2018-0140
- Navari RM, Reinhardt RR, Gralla RJ, et al. Reduction of cisplatininduced emesis by a selective neurokinin –1 receptor antagonist. L-754,030 antiemetic trials group. N Engl J Med. 1999;340(3):190-195. doi:10.1056/NEJM199901213400304
- 74. Bošnjak SM, Gralla RJ, Schwartzberg L. Prevention of chemotherapy-induced nausea: the role of neurokinin-1 (NK<sub>1</sub>) receptor antagonists. *Support Care Cancer*. 2017;25(5):1661-1671. doi:10.1007/s00520-017-3585-z
- 75. Roila F, Ruggeri B, Ballatori E, Del Favero A, Tonato M. Aprepitant versus dexamethasone for preventing chemotherapy-induced delayed emesis in patients with breast cancer: a randomized doubleblind study. J Clin Oncol. 2014;32(2):101-106. doi:10.1200/JCO. 2013.51.4547
- 76. Schwartzberg LS, Modiano MR, Rapoport BL, et al. Safety and efficacy of rolapitant for prevention of chemotherapy-induced nausea and vomiting after administration of moderately emetogenic chemotherapy or anthracycline and cyclophosphamide regimens in patients with cancer: a randomised, active controlled, double-blind, phase 3 trial. *Lancet Oncol.* 2015;16(9):107-1078. doi:10.1016/S1470-2045(15)00034-0
- 77. Rapoport BL, Chasen MR, Gridelli C, et al. Safety and efficacy of rolapitant for prevention of chemotherapy-induced nausea and vomiting after administration of cisplatin-based highly emetogenic chemotherapy in patients with cancer: two randomised, activecontrolled, double blind, phase 3 trials. *Lancet Oncol.* 2015;16(9): 1079-1089. doi:10.1016/S1470-2045(15)00035-2
- Saito H, Yoshizawa H, Yoshimori K, et al. Efficacy and safety of singledose fosaprepitant in the prevention of chemotherapy-induced nausea and vomiting in patients receiving high-dose cisplatin: a multicentre, randomised, double blind, placebo-controlled phase 3 trial. Ann Oncol. 2013;24(4):1067-1073. doi:10.1093/annonc/mds541
- 79. Poli-Bigelli S, Rodrigues-Pereira J, Carides AD, et al. Addition of the neurokinin 1 receptor antagonist aprepitant to standard antiemetic therapy improves control of chemotherapy-induced nausea and vomiting. Results from a randomized, double-blind, placebocontrolled trial in Latin America. *Cancer.* 2003;97(12):3090-3098. doi:10.1002/cncr.11433
- Roila F, Ruggeri B, Ballatori E, et al. Aprepitant versus metoclopramide, both combines with dexamethasone, for the prevention of cisplatin-induced delayed emesis: a randomized, double-blind study. *Ann Oncol.* 2015;26(6):1248-1253. doi:10.1093/annonc/mdv132
- 81. He M, Xu R, Liu M, et al. Use of dexamethasone and a 5-HT3 receptor antagonist with or without aprepitant to prevent chemotherapy-induced nausea and vomiting among patients with lung cancer who are treated with platinum-based chemotherapy: a systematic review and meta-analysis of randomized controlled trials. *Ann Palliat Med.* 2021;10(4):4308-4319. doi:10.21037/apm-20-2290

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- Gesztesi ZS, Song D, White PF, et al. Comparison of a new NK-1 antagonist (CP122,721) to ondansetron in the prevention of postoperative nausea and vomiting. *Anesth Analg.* 1998;86(2S):32S. doi:10. 1097/00000539-199802001-00032
- Gesztesi Z, Scuderi PE, White PF, et al. Substance P (neurokinin-1) antagonists prevents postoperative vomiting after abdominal hysterectomy procedures. *Anesthesiology*. 2000;93(4):931-937. doi:10. 1097/00000542-200010000-00009
- Diemunsch P, Schoeffler P, Bryssine B, et al. Antiemetic activity of the NK<sub>1</sub> receptor antagonist GR205171 in the treatment of established postoperative nausea and vomiting after major gynaecological surgery. Br J Anaesth. 1999;82(2):274-276. doi:10.1093/bja/82. 2.274
- Diemunsch P, Gan TJ, Philip BK, et al. Single-dose aprepitant vs ondansetron for the prevention of postoperative nausea and vomiting: a randomized, double-blind phase III trial in patients undergoing open abdominal surgery. Br J Anaesth. 2007;99(2):202-211. doi:10. 1093/bja/aem133
- Moon HY, Baek CW, Choi GJ, et al. Palonosetron and aprepitant for the prevention of postoperative nausea and vomiting in patients indicated for laparascopic gynaecological surgery: a double-blind randomised trial. *BMC Anesthesiol.* 2014;14(1):68. doi:10.1186/ 1471-2253-14-68
- Vallejo MC, Phelps AL, Ibinson JW, et al. Aprepitant plus ondansetron compared with ondansetron alone in reducing postoperative nausea and vomiting in ambulatory patients undergoing plastic surgery. *Plast Reconstr Surg.* 2012;129(2):519-526. doi:10.1097/PRS. 0b013e31822b6932
- Kranke P, Thompson JP, Dalby PL, et al. Comparison of vestipitant with ondansetron for the treatment of breakthrough postoperative nausea and vomiting after failed prophylaxis with ondansetron. Br J Anaesth. 2015;114(3):423-429. doi:10.1093/bja/aeu376
- Murakami C, Kakuta N, Kume K, et al. A comparison of fosaprepitant and ondansetron for preventing postoperative nausea and vomiting in moderate to high risk patients: a retrospective database analysis. *Biomed Res Int.* 2017;2017:5703528. doi:10.1155/2017/5703528
- 90. Singla NK, Singla SK, Chung F, et al. Phase II study to evaluate the safety and efficacy of the oral neurokinin-1 receptor antagonist casopitant (GW679769) administered with ondansetron for the prevention of postoperative and post discharge nausea and vomiting in high-risk patients. *Anesthesiology*. 2010;113(1):74-82. doi:10.1097/ ALN.0b013e3181d7b13a
- Habib AS, Keifer JC, Borel CO, White WD, Gan TJ. A comparison of the combination of aprepitant and dexamethasone versus the combination of ondansetron and dexamethasone for the prevention of postoperative nausea and vomiting in patients undergoing craniotomy. *Anesth Analg.* 2011;112(4):813-818. doi:10.1213/ANE. 0b013e3181ff47e2
- Sinha AC, Singh PM, Williams NW, Ochroch EA, Goudra BG. Aprepitant's prophylactic efficacy in decreasing postoperative nausea and vomiting in morbidly obese patients undergoing bariatric surgery. *Obes Surg.* 2014;24(2):225-231. doi:10.1007/s11695-013-1065-1
- Ham SY, Shim YH, Kim EH, Son MJ, Park WS, Lee JS. Aprepitant for antiemesis after laparoscopic gynaecologcial surgery: a randomised controlled trial. *Eur J Anesthesiol.* 2016;33(2):90-95. doi:10.1097/ EJA.00000000000242
- Liu M, Zhang H, Du B-X, et al. Neurokinin-1 receptor antagonists in preventing postoperative nausea and vomiting. A systematic review and meta-analysis. *Medicine*. 2015;94:1-10.
- 95. Weibel S, Rücker G, Eberhart LH, et al. Drugs for preventing postoperative nausea and vomiting in adults after general anaesthesia: a network meta-analysis. *Cochrane Database Syst Rev.* 2020;10(11): CD012859. doi:10.1002/14651858.CD012859.pub2
- 96. Gan T, Gan J, Singla N, et al. Rolapitant for the prevention of postoperative nausea and vomiting: a prospective double-blinded, placebo

controlled randomized trial. *Anesth Analg.* 2011;112(4):804-812. doi: 10.1213/ANE.0b013e31820886c3

- 97. Cristofori F, Thapar N, Saliakellis E, et al. Efficacy of the neurokinin-1 receptor antagonist aprepitant in children with cyclical vomiting syndrome. *Aliment Pharmacol Ther.* 2014;40(3):309-317. doi:10.1111/apt.12822
- Patel B, Downie J, Baylis J, Stephenson A, Bluebond-Langer M. Long-term daily administration of aprepitant for the management of intractable nausea and vomiting in children with life-limiting conditions: a case series. J Pain Symptom Manage. 2021;62(3):e225-e231. doi:10.1016/j.jpainsymman.2021.02.007
- Richards CA, Andrews PLR. Food refusal: a sign of nausea? J Pediatr Gastroenterol Nutr. 2004;38(2):227-228. doi:10.1097/00005176-200402000-00025
- 100. Jacob D, Busciglio I, Burton D, et al. Effects of NK<sub>1</sub> receptors on gastric motor functions and satiation in healthy humans: results from a controlled trial with the NK<sub>1</sub> antagonist aprepitant. *Am J Physiol Gastrointest Liver Physiol*. 2017;313(5):G505-G510. doi: 10.1152/ajpgi.00197.2017
- Ang D, Pauwels A, Akyuz F, Vos R, Tack J. Influence of a neurokinin-1 receptor antagonist (aprepitant) on gastric sensorimotor function in healthy volunteers. *Neurogastroenterol Motil.* 2013; 25(12):e830-e838. doi:10.1111/nmo.12210
- 102. Madsen JL, Fuglsang S. A randomized, placebo-controlled, crossover, double blind trial of the NK1 receptor antagonist aprepitant on gastrointestinal motor functions in healthy humans. *Aliment Pharmacol Ther.* 2008;27(7):609-615. doi:10.1111/j.1365-2036.2008. 03618.x
- 103. Pasricha PJ, Yates KP, Sarosiek I, et al. Aprepitant has mixed effects on nausea and reduces other symptoms in patients with gastroparesis and related disorders. *Gastroenterology*. 2018;154(1):65-76. doi: 10.1053/j.gastro.2017.08.033
- 104. Revicki DA, Rentz AM, Dubois D, et al. Gastroparesis cardinal symptom index (GCSI): development and validation of a patient reported assessment of severity of gastroparesis symptoms. *Qual Life Res.* 2004;13(4):833-844. doi:10.1023/B:QURE.0000021689. 86296.e4
- 105. Carlin JL, Lieberman VR, Dahal A, et al. Efficacy and safety of tradipitant in patients with diabetic and idiopathic gastroparesis in a randomized, placebo-controlled trial. *Gastroenterology*. 2021;160(1):76-87.e4. doi:10.1053/j.gastro.2020.07.029
- Fahler J, Wall GC, Leman BI. Gastroparesis-associated refractory nausea treated with aprepitant. Ann Pharmacother. 2012;46(12):e38. doi:10.1345/aph.1R484
- Chong K, Dhatariya K. A case of severe, refractory diabetic gastroparesis managed by prolonged use of aprepitant. *Nat Rev Endocrinol*. 2009;5(5):285-288. doi:10.1038/nrendo.2009.50
- Ingrosso MR, Camilleri M, Tack J, Ianiro G, Black CJ, Ford AC. Efficacy and safety of drugs for gastroparesis: systematic review and network meta-analysis. *Gastroenterology*. 2023;164(4):642-654. doi: 10.1053/j.gastro.2022.12.014
- 109. Nasdaq. Phase III study of tradipitant in gastroparesis fails to meet prespecified primary endpoint. https://www.nasdaq.com/articles/ vanda-%3A-phase-iii-study-of-tradipitant-ingastroparesis-fails-tomeet-prespecified-primary. 2022.
- Onishi T, Mori T, Yanagihara M, Furukawa N, Fukuda H. Similarities of the neuronal circuit for the induction of fictive vomiting between ferrets and dogs. *Auton Neurosci.* 2007;136(1-2):20-30. doi:10. 1016/j.autneu.2007.03.002
- 111. Lackner MR. Motion sickness: more than nausea and vomiting. *Exp Brain Res.* 2014;232(8):2493-2510. doi:10.1007/s00221-014-4008-8
- 112. Money KE, Cheung BS. Another function of the inner ear: facilitation of the emetic response to poisons. *Aviat Space Environ Med*. 1983;54(3):208-211.

- 113. Oman CM. Are evolutionary hypotheses for motion sickness "just so stories"? *J Vestib Res.* 2012;22(2–3):117-127. doi:10.3233/VES-2011-0432
- 114. Treisman M. Motion sickness: an evolutionary hypothesis. *Science*. 1977;197(4302):493-495. doi:10.1126/science.301659
- 115. Gan TJ. Risk factors for postoperative nausea and vomiting. *Anesth Analg.* 2006;102(6):1884-1898. doi:10.1213/01.ANE.0000219597. 16143.4D
- 116. Warr D. Prognostic factors for chemotherapy induced nausea and vomiting. *Eur J Pharmacol*. 2014;722:192-196. doi:10.1016/j.ejphar. 2013.10.015
- 117. Sugiyama Y, Suzuki T, DeStefino VJ, Yates BJ. Integrative responses of neurons in nucleus tractus solitarius to visceral afferent stimulation and vestibular stimulation in vertical planes. *Am J Physiol Regul Integr Comp Physiol.* 2011;301(5):R1380-R1390. doi:10.1152/ ajpregu.00361.2011
- Yates BJ, Grelot L, Kerman IA, Balaban CD, Jakus J, Miller AD. Organization of vestibular inputs to nucleus tractus solitarius and adjacent structures in cat brain stem. Am J Physiol Regul Integr Comp Physiol. 1994;267(4):R974-R983. doi:10.1152/ajpregu.1994.267.4.R974
- 119. Soto E, Vega R. Neuropharmacology of vestibular system disorders. *Curr Neuropharmacol.* 2010;8(1):26-40. doi:10.2174/157015910 790909511
- 120. Koga T, Fukuda H. Neurons in the nucleus of the solitary tract mediating inputs from emetic vagal afferents and the area postrema to the pattern generator for the emetic act in dogs. *Neurosci Res.* 1992; 14(3):166-179. doi:10.1016/0168-0102(92)90078-Q
- 121. Ariumi H, Saito R, Nago S, Hyakusoku M, Takanon Y, Kamiya H-o. The role of tachykinin NK-1 receptors in the area postrema of ferrets in emesis. *Neurosci Lett.* 2000;286(2):123-126. doi:10.1016/ S0304-3940(00)01113-7
- 122. Maubach KA, Jones RS. Electrophysiological characterisation of tachykinin receptors in the rat nucleus of the solitary tract and dorsal motor nucleus of the vagus *in vitro*. Br J Pharmacol. 1997;122(6): 1151-1159. doi:10.1038/sj.bjp.0701482
- 123. Andrews PLR, Rudd JA. The role of tachykinins and the tachykinin NK<sub>1</sub> receptor in nausea and emesis. In: Holzer P, ed. *Tachykinins*. Springer; 2004:359-440. doi:10.1007/978-3-642-18891-6\_12
- Carpenter DO, Briggs DB, Knox AP, Strominger N. Excitation of area postrema neurons by transmitters, peptides, and cyclic nucleotides. *J Neurophysiol*. 1988;59(2):358-369. doi:10.1152/jn.1988.59.2.358
- 125. Carpenter DO, Briggs DB, Strominger N. Peptide-induced emesis in dogs. *Behav Brain Res.* 1984;11(3):277-281. doi:10.1016/0166-4328(84)90220-1
- 126. Gardner CJ, Bountra C, Bunce KT, et al. Anti-emetic activity of neurokinin  $NK_1$  receptor antagonists is mediated centrally in the ferret. Br J Pharmacol. 1994;112:516P.
- 127. Rezzani R, Franco C, Franceschetti L, Gianó M, Favero G. A focus on enterochromaffin cells among the enteroendocrine cells: localization, morphology, and role. *Int J Mol Sci.* 2022;23(7):3758. doi:10. 3390/ijms23073758
- 128. Higa GM, Auber ML, Altahoa R, et al. 5-hydroxyindoleacetic acid and substance P profiles in patients receiving emetogenic chemotherapy. J Oncol Pharm Pract. 2006;12(4):201-209. doi:10.1177/ 1078155206072080
- 129. Higa GM, Auber ML, Hobbs G. Identification of a novel marker associated with risk for delayed chemotherapy-induced vomiting. *Support Care Cancer*. 2012;20(11):2803-2809. doi:10.1007/s00520-012-1402-2
- 130. Matsumoto S, Kawasaki Y, Mikami M, et al. Relationship between cancer chemotherapeutic drug-induced delayed emesis and plasma levels of substance P in two patients with small cell lung cancer. *Gan to Kagaku Ryoho.* 1999;26(4):535-538.
- 131. Park HS, Won HS, An HJ, et al. Elevated serum substance P level as a predictive marker for moderately emetogenic chemotherapy-

induced nausea and vomiting: a prospective cohort study. *Cancer Med.* 2021;10(3):1057-1065. doi:10.1002/cam4.3693

- 132. Takahashi T, Nakamura Y, Tsuya A, Murakami H, Endo M, Yamamoto N. Pharmacokinetics of aprepitant and dexamethasone after administration of chemotherapeutic agents and effects of plasma substance P concentration on chemotherapy-induced nausea and vomiting in Japanese cancer patients. *Cancer Chemother Pharmacol.* 2011;68(3):653-659. doi:10.1007/s00280-010-1519-2
- Rikard-Bell GC, Törk I, Sullivan C, Scheibner T. Distribution of substance P-like immunoreactive fibres and terminals in the medulla oblongata of the human infant. *Neuroscience*. 1990;34(1):133-148. doi:10.1016/0306-4522(90)90308-Q
- Newton BW, Maley B, Traurig H. The distribution of substance P, enkephalin, and serotonin immunoreactivities in the area postrema of the rat and cat. J Comp Neurol. 1985;234(1):87-104. doi:10.1002/ cne.902340107
- 135. Boissonade FM, Davison JS, Egizii R. The dorsal vagal complex of the ferret: anatomical and immunohistochemical studies. *Neurogastroenterol Motil.* 1996;8(3):255-272. doi:10.1111/j.1365-2982.1996. tb00265.x
- Cooper PE, Fernstrom MH, Rorstad OP, Leeman SE, Martin JB. The regional distribution of somatostatin, substance P and neurotensin in human brain. Brain Res. 1981;218(1-2):219-232. doi:10.1016/ 0006-8993(81)91302-0
- Zettler G, Schlosser L. Über die Verteilung von Substanz P und Cholinacetylase im Gehirn. Naunyn-Schmiedeberg's Exp Pathol Pharmakol. 1955;224:159-175.
- Hietala J, Nyman MJ, Eskola O, et al. Visualization and quantification of neurokinin-1 (NK1) receptors in the huma brain. *Mol Imaging Biol.* 2005;7(4):262-272. doi:10.1007/s11307-005-7001-6
- 139. Ray AP, Darmani NA. A histologically derived stereotaxic atlas and substance P immunohistochemistry in the brain of the least shrew (*Cryptotis parva*) support its role as a model organism for behavioural and pharmacological research. *Brain Res.* 2007;1156:99-111. doi:10. 1016/j.brainres.2007.04.061
- 140. Lindstrom PA, Brizzee KR. Relief of intractable vomiting from surgical lesion in the area postrema. *J Neurosurg.* 1962;19(3):228-236. doi:10.3171/jns.1962.19.3.0228
- Borison HL, Wang SC. Physiology and pharmacology of vomiting. Pharmacol Rev. 1953;5(2):193-230.
- 142. Bhandari P, Bingham S, Andrews PLR. The neuropharmacology of loperamide-induced emesis in the ferret: the role of the area postrema, vagus, opiate and 5-HT<sub>3</sub> receptors. *Neuropharmacology*. 1992; 31(8):735-742. doi:10.1016/0028-3908(92)90034-M
- Borison HL. Area postrema: chemoreceptor circumventricular organ of the medulla oblongata. *Prog Neurobiol*. 1989;32(5):351-390. doi: 10.1016/0301-0082(89)90028-2
- 144. Percie du Sert N, Rudd JA, Moss R, Andrews PLR. The delayed phase of cisplatin-induced emesis is mediated by the area postrema and not the abdominal visceral innervation in the ferret. *Neurosci Lett.* 2009;465(1):16-20. doi:10.1016/j.neulet.2009.08.075
- 145. Borner T, Shaulson ED, Ghidewon MY, et al. GDF15 induces anorexia through nausea and emesis. *Cell Metab.* 2020;31(2):351-362. doi:10.1016/j.cmet.2019.12.004
- 146. Makwana R, Crawley E, Straface M, et al. Synergistic augmentation of rhythmic myogenic contractions of human stomach by [Arg<sup>8</sup>]vasopressin and adrenaline: implications for the induction of nausea. Br J Pharmacol. 2022;179(24):5305-5322. doi:10.1111/ bph.15943
- 147. Andrews PLR, Rapeport WG, Sanger GJ. Neuropharmacology of emesis induced by anti-cancer therapy. *Trends Pharmacol Sci.* 1988; 9(9):334-341. doi:10.1016/0165-6147(88)90106-X
- 148. Andrews PLR, Rudd JA. The physiology and pharmacology of nausea and vomiting induced by anti-cancer chemotherapy in humans. In: Navari RM, ed. *Management of chemotherapy-induced nausea and*

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BRITISH PHARMACOLOGICAI vomiting: new agents and new uses of current agents; 2016:5-44. doi: 10.1007/978-3-319-27016-6\_2

- 149. Obara Y, Machida T, Takano Y, et al. Cisplatin increases the number of enterochromaffin cells containing substance P in rat intestine. *Naunyn Schmiedebergs Arch Pharmacol.* 2018;391(8):847-858. doi: 10.1007/s00210-018-1493-5
- Smith JA, Harle A, Dockry R, et al. Aprepitant for cough in lung cancer: a randomised placebo-controlled trial and mechanistic insights. *Am J Respir Crit Care Med.* 2021;203(6):737-745. doi:10.1164/rccm. 202006-2359OC
- 151. Tattersall FD, Rycroft W, Cumberbatch M, et al. The novel NK<sub>1</sub> receptor antagonist MK-0869 (L-754,030) and its water soluble phosphoryl prodrug, L-758,298, inhibit acute and delayed cisplatin-induced emesis in ferrets. *Neuropharmacology*. 2000;39(4):652-663. doi:10.1016/S0028-3908(99)00172-0
- McCoull D, Veale EL, Walsh Y, et al. Aprepitant is a novel, selective activator of the K2P channel TRAAK. *Biochem Biophys Res Commun.* 2022;588:41-46. doi:10.1016/j.bbrc.2021.12.031
- Minami M, Endo T, Yokota H, et al. Effects of CP-99, 994, a tachykinin NK1 receptor antagonist, on abdominal afferent vagal activity in ferrets: evidence for involvement of NK<sub>1</sub> and 5-HT<sub>3</sub> receptors. *Eur J Pharmacol.* 2001;428(2):215-220. doi:10.1016/S0014-2999(01) 01297-3
- Rojas C, Raje M, Tsukamoto T, Slusher BS. Molecular mechanisms of 5-HT<sub>3</sub> and tachykinin NK<sub>1</sub> receptor antagonists in prevention of emesis. *Eur J Pharmacol.* 2014;722:26-37. doi:10.1016/j.ejphar. 2013.08.049
- 155. Fukuda H, Koga T. Non-respiratory neurons in the Bötzinger complex exhibiting appropriate firing patterns to generate the emetic act in dogs. *Neurosci Res.* 1992;14(3):180-194. doi:10.1016/0168-0102 (92)90079-R
- 156. Baude A, Lanoir J, Vernier P, Puizillout JJ. Substance P-immunoreactivity in the dorsal medial region of the medulla in the cat: effects of nodosectomy. *J Chem Neuroanat*. 1989;2(2):67-81.
- 157. Shiroshita Y, Koga T, Fukuda H. Capsaicin in the 4<sup>th</sup> ventricle abolishes retching and transmission of emetic vagal afferents to solitary nucleus neurons. *Eur J Pharmacol.* 1997;339(2-3):183-192. doi:10. 1016/S0014-2999(97)01370-8
- 158. Saito R, Suehiro Y, Ariumi H, et al. Anti-emetic effects of a novel NK-1 receptor antagonist HSP-117 in ferrets. *Neurosci Lett.* 1998; 254(3):169-172. doi:10.1016/S0304-3940(98)00682-X
- 159. Fukuda H, Koga T. The Bötzinger complex as the pattern generator for retching and vomiting in the dog. *Neurosci Res.* 1991;12(4):471-485. doi:10.1016/S0168-0102(09)80001-1
- 160. Fukuda H, Koga T, Furukawa N, Nakamura E, Hatano M, Yanagihara M. The site of the antiemetic action of NK1 receptor antagonists. In: Donnerer J, ed. Antiemetic therapy. Karger; 2003:33-77. doi:10.1159/000071410
- Lang IM, Sarna SK, Condon RE. Gastrointestinal motor correlates of vomiting in the dog: quantification and characterization as an independent phenomenon. *Gastroenterology*. 1986;90(1):40-47. doi:10. 1016/0016-5085(86)90072-7
- 162. Lang IM. Digestive tract motor correlates of vomiting and nausea. *Can J Physiol Pharmacol.* 1990;68(2):242-253. doi:10.1139/y90-038
- 163. Sanger GJ, Andrews PLR. An analysis of the pharmacological rationale for selecting drugs to inhibit vomiting or increase gastric emptying during treatment of gastroparesis. *Aliment Pharmacol Ther.* 2023; 57(9):962-978. doi:10.1111/apt.17466
- Grélot L, Bianchi AL. Mutifunctional medullary respiratory neurons. In: Miller AB, Bianchi AL, Bishop BP, eds. Neural control of the respiratory muscles. CRC Press; 2019:297-304. doi:10.1201/ 9780429277740-25
- 165. Leslie RA. Neuroactive substances in the dorsal vagal complex of the medulla oblongata: nucleus of the tractus solitarius, area

postrema, and dorsal motor nucleus of the vagus. *Neurochem Int.* 1985;7(2):191-211. doi:10.1016/0197-0186(85)90106-8

- 166. Sun X, Xu L, Guo F, Luo W, Gao S, Luan X. Neurokinin-1 receptor blocker CP-99994 improved emesis induced by cisplatin via regulating the activity of gastric distension responsive neurons in the dorsal motor nucleus of vagus and enhancing gastric motility in rats. *Neurogastroenterol Motil.* 2017;29(10):e13096. doi:10.1111/nmo.13096
- 167. Andrews PLR, Fussey IV, Scratcherd T. The spontaneous discharge in abdominal vagal efferents in the dog and ferret. *Pflügers Arch*. 1980;387(1):55-60. doi:10.1007/BF00580844
- 168. Magendie F. Mémoire sur le vomissement, lu à la première classe de l'Institut de France. (Ed.1813). Cez Crochard; 1813:48.
- 169. Andrews PLR, Davis CJ, Bingham S, Davidson HIM, Hawthorn J, Maskell L. The abdominal visceral innervation and the emetic reflex: pathways, pharmacology and plasticity. *Can J Physiol Pharmacol.* 1990;68(2):325-345. doi:10.1139/y90-047
- Andrews PLR, Cai W, Rudd JA, Sanger GJ. COVID-19, nausea and vomiting. J Gastroenterol Hepatol. 2021;36(3):646-656. doi:10. 1111/jgh.15261
- 171. Andrews PLR, Williams RSB, Sanger GJ. Anti-emetic effects of thalidomide: evidence, mechanism of action, and future directions. *Curr Res Pharmacol Drug Dis.* 2023;3:100138.
- 172. Svensson E, Apergis-Schoute J, Burnstock G, Nusbaum MP, Parker D, Schiöth HB. General principles of neuronal co-transmission: insights from multiple model systems. *Front Neural Circuits*. 2019;12:117. doi:10.3389/fncir.2018.00117
- 173. Furukawa N, Hatano M, Fukuda H. Glutaminergic vagal afferents may mediate both retching and gastric adaptive relaxation in dogs. *Auton Neurosci.* 2001;93(1-2):21-30. doi:10.1016/S1566-0702(01) 00322-8
- Cropper EC, Jing J, Vilim FS, Weiss KR. Peptide cotransmitters as dynamic, intrinsic modulators of network activity. *Front Neural Circuits*. 2018;12:78. doi:10.3389/fncir.2018.00078
- 175. Jarcho JM, Feier NA, Bert A, et al. Diminished neurokinin-1 receptor availability in patients with two forms of chronic visceral pain. *Pain*. 2013;154(7):987-996. doi:10.1016/j.pain.2013.02.026
- 176. Horn CC. Measuring the nausea-to-emesis continuum in nonhuman animals: refocusing on gastrointestinal vagal signaling. *Exp Brain Res.* 2014;232(8):2471-2481. doi:10.1007/s00221-014-3985-y
- 177. Ottaviani MM, Wright L, Dawood T, Macefield VG. In vivo recordings from the human vagus nerve using ultrasound guided microneurography. J Physiol. 2020;598(17):3569-3576. doi:10.1113/ JP280077
- 178. Bergstrom M, Hargreaves RJ, Burns HD, et al. Human positron emission tomography studies of brain neurokinin 1 receptor occupancy by aprepitant. *Biol Psychiatry*. 2004;55(10):1007-1012. doi:10.1016/j.biopsych.2004.02.007
- 179. Borsook D, Upadhay J, Klimas M, et al. Decision-making using fMRI in clinical drug development: revisiting NK-1 receptor antagonists for pain. *Drug Discov Today*. 2012;17(17–18):964-973. doi:10.1016/ j.drudis.2012.05.004
- 180. Hoppe JM, Frick A, Åhs F, et al. Association between amygdala neurokinin-1 receptor availability and anxiety-related personality traits. *Transl Psychiatry*. 2018;8(1):168. doi:10.1038/s41398-018-0163-1
- Keller M, Montgomery S, Ball W, et al. Lack of efficacy of the substance P (neurokinin receptor) antagonist aprepitant in the treatment of major depressive disorder. *Biol Psychiatry*. 2006;59(3): 216-223. doi:10.1016/j.biopsych.2005.07.013
- 182. Ratti E, Bettica P, Alexander R, et al. Full central neurokinin-1 receptor blockade is required for efficacy in depression: evidence from orvepitant clinical studies. J Psychopharmacol. 2013;27(5):424-434. doi:10.1177/0269881113480990

- 183. Fujimura Y, Yasuno F, Farris A, et al. Decreased neurokinin-1 (substance P) receptor binding in patients with panic disorder: positron emission tomography study with [18F]SPA-RQ. *Biol Psychiatry*. 2009;66(1):94-97. doi:10.1016/j.biopsych.2008.12.027
- Barreca T, Corsini G, Cataldi A, et al. Effect of the 5-HT3 receptor antagonist ondansetron on plasma AVP secretion: a study in cancer patients. *Biomed Pharmacother*. 1996;50(10):512-514. doi:10.1016/ S0753-3322(97)89285-2
- Sanger GJ. Neurokinin NK<sub>1</sub> and NK<sub>3</sub> receptors as targets for drugs to treat gastrointestinal motility disorders and pain. Br J Pharmacol. 2004;141(8):1303-1312. doi:10.1038/sj.bjp.0705742
- 186. Chen H, Redelman D, Ro S, Ward SM, Ordög T, Sanders S. Selective labelling and isolation of functional classes of interstitial cells of Cajal of human and murine small intestine. Am J Physiol Cell Physiol. 2007;292(1):C497-C507. doi:10.1152/ajpcell.00147.2006
- 187. Faussone-Pellegrini M-S. Relationships between neurokinin receptor-expressing interstitial cells of Cajal and tachykininergic nerves in the gut. J Cell Mol Med. 2006;10(1):20-32. doi:10.1111/j. 1582-4934.2006.tb00288.x
- Lavin S, Southwell BR, Murphy R, Jenkinson KM, Furness JB. Activation of neurokinin 1 receptors on interstitial cells of Cajal of the Guinea-pig small intestine by substance P. *Histochem Cell Biol.* 1998; 110(3):263-271. doi:10.1007/s004180050288
- Liu JYH, Rudd JA. Predicting drug adverse effects using a new gastro-intestinal pacemaker activity drug database (GIPADD). *Sci Rep.* 2023;13(1):6935. doi:10.1038/s41598-023-33655-5
- 190. Khanna L, Zheng T, Atieh J, et al. Clinical trial: a single-centre, randomised, controlled trial of tradipitant on satiation, gastric functions, and serum drug levels in healthy volunteers. *Alimentary Pharmacol Ther.* 2022;56(2):224-230. doi:10.1111/apt.17065
- 191. Koch KL. Gastric dysrhythmias: a potential objective measure of nausea. Exp Brain Res. 2014;232(8):2553-2561. doi:10.1007/ s00221-014-4007-9
- 192. O'Grady G, Gharibans AA, Du P, Huizinga JD. The gastric conduction system in health and disease: a translational review. *Am J Physiol Gastrointest Liver Physiol*. 2021;321(5):G527-G542. doi: 10.1152/ajpgi.00065.2021
- 193. Doi A, Ramirez J-M. State-dependent interactions between excitatory neuromodulators in the neuronal control of breathing. *J Neurosci.* 2010;30(24):8251-8262. doi:10.1523/JNEUROSCI. 5361-09.2010
- 194. Bailey CP, Maubach KA, Jones RSG. Neurokinin-1 receptors in th rat nucleus tractus solitarius: pre- and postsynaptic modulation of glutamate and GABA release. *Neuroscience*. 2004;127(2):467-479. doi:10.1016/j.neuroscience.2004.05.025
- Sanger GJ, Andrews PLR. Treatment of nausea and vomiting: gaps in our knowledge. Auton Neurosci. 2006;129(1-2):3-16. doi:10.1016/j. autneu.2006.07.009
- Borner T, Reiner BC, Crist RC, et al. GIP receptor agonism blocks chemotherapy-induced nausea and vomiting. *Mol Metab.* 2023;73: 101743. doi:10.1016/j.molmet.2023.101743

- Horn CC, Ardell JL, Fisher LE. Electroceutical targeting of the autonomic nervous system. *Phys Ther.* 2019;34(2):150-162. doi:10. 1152/physiol.00030.2018
- Ramadi KB, Srinivasan S, Traverso G. Electroceuticals in the gastrointestinal tract. *Trends Pharmacol Sci.* 2020;41(12):960-976. doi:10. 1016/j.tips.2020.09.014
- Minton N, Swift R, Lawlor C, Mant T, Henry J. Ipecacuanha-induced emesis: a human model for testing antiemetic drug activity. *Clin Pharmacol Ther*. 1993;54(1):53-57. doi:10.1038/clpt.1993.109
- 200. Proctor JD, Chremos AN, Evans EF, Wasserman AJ. An apomorphineinduced vomiting model for antiemetic studies. *J Clin Pharmacol.* 1978;18(2-3):95-99. doi:10.1002/j.1552-4604.1978.tb02427.x
- 201. Soergel DG, Subach RA, Burnham N, et al. Biased agonism of the μopioid receptor by TRV130 increases analgesia and reduces ontarget adverse effects versus morphine: a randomized, double blind, placebo-controlled, crossover study in healthy volunteers. *Pain*. 2014;155(9):1829-1835. doi:10.1016/j.pain.2014.06.011
- 202. Gabby ME, Bugin K, Lyons E. Review article: the evolution of endpoint assessments for chemotherapy-induced nausea and vomiting and post-operative nausea and vomiting—a perspective from the US Food and Drug Administration. *Aliment Pharmacol Ther.* 2021;54(1): 7-13. doi:10.1111/apt.16386
- Miner WD, Sanger GJ. Inhibition of cisplatin-induced vomiting by selective 5-hydroxytryptamine M-receptor antagonism. Br J Pharmacol. 1986;88(3):497-499. doi:10.1111/j.1476-5381.1986.tb10228.x
- Grelot L, Miller AD. Neural control of respiratory muscle activation during vomiting. In: Miller AB, Bianchi AL, Bishop BP, eds. *Neural control of the respiratory muscles*. CRC Press; 1997:239-248.
- 205. Machida T, Takano Y, lizuka K, Machida M, Hirafuji M. Methotrexate causes acute hyperplasia of enterochromaffin cells containing substance P in the intestinal mucosa of rats. J Pharmacol Sci. 2020; 133(3):190-193. doi:10.1016/j.jphs.2017.02.009
- 206. Upadhay J, Anderson J, Schwarz AJ, et al. Imaging of drugs with and without clinical efficacy. *Neuropsychopharmacology*. 2011;36(13): 2659-2673. doi:10.1038/npp.2011.156

#### SUPPORTING INFORMATION

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